

AGRICULTURE

A global synthesis reveals biodiversity-mediated benefits for crop production

Matteo Dainese^{1,2*}, Emily A. Martin², Marcelo A. Aizen³, Matthias Albrecht⁴, Ignasi Bartomeus⁵, Riccardo Bommarco⁶, Luisa G. Carvalheiro^{7,8}, Rebecca Chaplin-Kramer⁹, Vesna Gagic¹⁰, Lucas A. Garibaldi¹¹, Jaboury Ghazoul¹², Heather Grab¹³, Mattias Jonsson⁶, Daniel S. Karp¹⁴, Christina M. Kennedy¹⁵, David Kleijn¹⁶, Claire Kremen¹⁷, Douglas A. Landis¹⁸, Deborah K. Letourneau¹⁹, Lorenzo Marini²⁰, Katja Poveda¹³, Romina Rader²¹, Henrik G. Smith^{22,23}, Teja Tschamntke²⁴, Georg K. S. Andersson²², Isabelle Badenhauer^{25,26}, Svenja Baensch^{24,27}, Antonio Diego M. Bezerra²⁸, Felix J. J. A. Bianchi²⁹, Virginie Boreux^{12,30}, Vincent Bretagnolle³¹, Berta Caballero-Lopez³², Pablo Cavigliasso³³, Aleksandar Ćetković³⁴, Natacha P. Chacoff³⁵, Alice Classen², Sarah Cusser³⁶, Felipe D. da Silva e Silva^{37,38}, G. Arjen de Groot³⁹, Jan H. Dudenhöffer⁴⁰, Johan Ekroos²², Thijs Fijen¹⁶, Pierre Franck⁴¹, Breno M. Freitas²⁸, Michael P. D. Garratt⁴², Claudio Gratton⁴³, Juliana Hipólito^{11,44}, Andrea Holzschuh², Lauren Hunt⁴⁵, Aaron L. Iverson¹³, Shalene Jha⁴⁶, Tamar Keasar⁴⁷, Tania N. Kim⁴⁸, Miriam Kishinevsky⁴⁹, Björn K. Klatt^{23,24}, Alexandra-Maria Klein³⁰, Kristin M. Krewenka⁵⁰, Smitha Krishnan^{12,51,52}, Ashley E. Larsen⁵³, Claire Lavigne⁴¹, Heidi Liere⁵⁴, Bea Maas⁵⁵, Rachel E. Mallinger⁵⁶, Eliana Martinez Pachon⁵⁷, Alejandra Martínez-Salinas⁵⁸, Timothy D. Meehan⁵⁹, Matthew G. E. Mitchell⁶⁰, Gonzalo A. R. Molina⁶¹, Maike Nesper¹², Lovisa Nilsson²², Megan E. O'Rourke⁶², Marcell K. Peters², Milan Plečáček³⁴, Simon G. Potts⁴³, Davi de L. Ramos⁶³, Jay A. Rosenheim⁶⁴, Maj Rundlöf²³, Adrien Rusch⁶⁵, Agustín Sáez⁶⁶, Jeroen Scheper^{16,39}, Matthias Schleuning⁶⁷, Julia M. Schmack⁶⁸, Amber R. Sciligo⁶⁹, Colleen Seymour⁷⁰, Dara A. Stanley⁷¹, Rebecca Stewart²², Jane C. Stout⁷², Louis Sutter⁴, Mayura B. Takada⁷³, Hisatomo Taki⁷⁴, Giovanni Tamburini³⁰, Matthias Tschumi⁴, Blandina F. Viana⁷⁵, Catrin Westphal²⁷, Bryony K. Willcox²¹, Stephen D. Wratten⁷⁶, Akira Yoshioka⁷⁷, Carlos Zaragoza-Trello⁵, Wei Zhang⁷⁸, Yi Zou⁷⁹, Ingolf Steffan-Dewenter²

Human land use threatens global biodiversity and compromises multiple ecosystem functions critical to food production. Whether crop yield-related ecosystem services can be maintained by a few dominant species or rely on high richness remains unclear. Using a global database from 89 studies (with 1475 locations), we partition the relative importance of species richness, abundance, and dominance for pollination; biological pest control; and final yields in the context of ongoing land-use change. Pollinator and enemy richness directly supported ecosystem services in addition to and independent of abundance and dominance. Up to 50% of the negative effects of landscape simplification on ecosystem services was due to richness losses of service-providing organisms, with negative consequences for crop yields. Maintaining the biodiversity of ecosystem service providers is therefore vital to sustain the flow of key agroecosystem benefits to society.

INTRODUCTION

Natural and modified ecosystems contribute a multitude of functions and services that support human well-being (1, 2). It has long been recognized that biodiversity plays an important role in the functioning of ecosystems (3, 4), but the dependence of ecosystem services on biodiversity is under debate. An early synthesis revealed inconsistent results (5), whereas subsequent studies suggest that a few dominant species may supply the majority of ecosystem services (6, 7). It thus remains unclear whether a few dominant or many complementary species are needed to supply ecosystem services.

The interpretation of earlier studies has been controversial because multiple mechanisms underlying changes in ecosystem service response to biodiversity can operate in combination (8, 9). On one hand, communities with many species are likely to include species responsible for large community-wide effects due to statistical selection. On the other hand, such diverse communities may contain a particular combination

of species that complement each other in service provisioning. While these mechanisms imply positive effects of species richness on ecosystem service supply, total organism abundance or dominance of certain species may also drive the number of interactions benefiting ecosystem service supply. Depending on the relative importance of species complementarity, community abundance, and the role of dominant species, different relationships between species richness and ecosystem services can be expected (10).

In real-world ecosystems, natural communities consist of a few highly abundant (dominant species) and many rare ones. The importance of richness, abundance, and dominance is likely to be influenced by the extent to which relative abundance changes with species richness (11) and by differences in the effectiveness and degree of specialization of service-providing communities. However, these three aspects of diversity have typically been tested in isolation and mainly in small-scale experimental settings (12, 13), while a synthetic study contrasting their

Copyright © 2019
The Authors, some
rights reserved;
exclusive licensee
American Association
for the Advancement
of Science. No claim to
original U.S. Government
Works. Distributed
under a Creative
Commons Attribution
NonCommercial
License 4.0 (CC BY-NC).

relative importance in real-world ecosystems is still lacking. A major limitation to resolving these relationships is a lack of evidence from real-world human-driven biodiversity changes (14, 15), particularly for ecosystem services in agroecosystems. For instance, changes in richness and total or relative abundance of service-providing organisms in response to land-clearing for agriculture (16, 17) could alter the flow of benefits to people in different ways compared to experimental random loss of biodiversity.

Over the past half-century, the need to feed a growing world population has led to markedly expanded and intensified agricultural production, transforming many regions into simplified landscapes (18). This transformation not only has contributed to enhanced agricultural production but also has led to the degradation of the global environment. The loss of biodiversity can disrupt key intermediate services to agriculture, such as crop pollination (19) and biological pest control (20), which underpin the final provisioning service of crop

production (21). The recent stagnation or even decline of crop yields with ongoing intensification (22) indicates that alternative pathways are necessary to maintain future stable and sustainable crop production (23–25). An improved understanding of global biodiversity-driven ecosystem services in agroecosystems and their cascading effects on crop production is urgently needed to forecast future supplies of ecosystem services and to pursue strategies for sustainable management (15).

We compiled an extensive database comprising 89 studies that measured richness and abundance of pollinators, pest natural enemies, and associated ecosystem services at 1475 sampling locations around the world (Fig. 1A). We focused on the ecosystem services of pollination and biological pest control because these services are essential to crop production and have been the focus of much research in recent decades (26). We quantified pollinator and pest natural enemy richness as the number of unique taxa sampled from each location (field), abundance as the number of observed individuals, and evenness (or the complementary term,

¹Institute for Alpine Environment, Eurac Research, Viale Druso 1, 39100 Bozen/Bolzano, Italy. ²Department of Animal Ecology and Tropical Biology, Biocenter, University of Würzburg, Am Hubland, 97074 Würzburg, Germany. ³Grupo de Ecología de la Polinización, INIBIOMA, Universidad Nacional del Comahue, CONICET, 8400 Bariloche, Río Negro, Argentina. ⁴Agroecology and Environment, Agroscope, Reckenholzstrasse 191, 8046 Zurich, Switzerland. ⁵Estación Biológica de Doñana (EBD-CSIC), Integrative Ecology, E-41092 Sevilla, Spain. ⁶Swedish University of Agricultural Sciences, Department of Ecology, 750 07 Uppsala, Sweden. ⁷Departamento de Ecología, Universidade Federal de Goiás (UFG), Goiânia, Brazil. ⁸Faculdade de Ciências, Centre for Ecology, Evolution and Environmental Changes (CE3C), Universidade de Lisboa, Lisboa, Portugal. ⁹Natural Capital Project, Stanford University, Stanford, CA 94618, USA. ¹⁰CSIRO, GPO Box 2583, Brisbane, QLD 4001, Australia. ¹¹Instituto de Investigaciones en Recursos Naturales, Agroecología y Desarrollo Rural (INRAD), Sede Andina, Universidad Nacional de Río Negro (UNRN) y CONICET, Mitre 630, CP 8400 San Carlos de Bariloche, Río Negro, Argentina. ¹²Department of Environmental Systems Science, ETH Zurich, Universitätstrasse 16, 8092 Zurich, Switzerland. ¹³Department of Entomology, Cornell University, Ithaca, NY 14853, USA. ¹⁴Department of Wildlife, Fish and Conservation Biology, University of California Davis, Davis, CA 95616, USA. ¹⁵Global Lands Program, The Nature Conservancy, 117 E. Mountain Avenue, Fort Collins, CO 80524, USA. ¹⁶Plant Ecology and Nature Conservation Group, Wageningen University, Droevendaalsesteeg 3a, Wageningen 6708 PB, Netherlands. ¹⁷IREs and Biodiversity Research Centre, University of British Columbia, Vancouver, BC, Canada. ¹⁸Department of Entomology and Great Lakes Bioenergy Research Center, Michigan State University, 204 CIPS, 578 Wilson Ave, East Lansing, MI 48824, USA. ¹⁹Department of Environmental Studies, University of California, Santa Cruz, Santa Cruz, CA 95064, USA. ²⁰DAFNAE, University of Padova, via dell'Università 16, 35020 Legnaro, Padova, Italy. ²¹School of Environment and Rural Science, University of New England, Armidale, NSW 2350, Australia. ²²Centre for Environmental and Climate Research, Lund University, S-223 62 Lund, Sweden. ²³Department of Biology, Lund University, S-223 62 Lund, Sweden. ²⁴Agroecology, Department of Crop Sciences, University of Göttingen, D-37077 Göttingen, Germany. ²⁵USC1339 INRA-CNRS, CECB UMR 7372, CNRS and Université de La Rochelle, Beauvoir sur Niort 79360, France. ²⁶INRA, Unité de Recherche Pluridisciplinaire Prairies et Plantes Fourragères (URP3F), Lusignan 86600, France. ²⁷Functional Agroecology, Department of Crop Sciences, University of Göttingen, Germany. ²⁸Departamento de Zootecnia—CCA, Universidade Federal do Ceará, 60.356-000 Fortaleza, CE, Brazil. ²⁹Farming Systems Ecology, Wageningen University and Research, P.O. Box 430, 6700 AK Wageningen, Netherlands. ³⁰Chair of Nature Conservation and Landscape Ecology, University of Freiburg, Tennenbacher Straße 4, 79106 Freiburg, Germany. ³¹LTSE Zone Atelier Plaine and Val de Sevre, CECB UMR 7372, CNRS and Université de La Rochelle, Beauvoir sur Niort 79360, France. ³²Arthropods Department, Natural Sciences Museum of Barcelona, 08003 Barcelona, Spain. ³³Instituto Nacional de Tecnología Agropecuaria, Estación Experimental Concordia, Estación Yuqueri y vías del Ferrocarril s/n, 3200 Entre Ríos, Argentina. ³⁴Faculty of Biology, University of Belgrade, Studentski trg 16, 11000 Belgrade, Serbia. ³⁵Instituto de Ecología Regional (IER), Universidad Nacional de Tucumán, CONICET, 4107 Yerba Buena, Tucumán, Argentina. ³⁶W.K. Kellogg Biological Station, Michigan State University, Hickory Corners, MI 49060, USA. ³⁷Federal Institute of Education, Science and Technology of Mato Grosso, Campus of Barra do Garças/MT, 78600-000, Brazil. ³⁸Center of Sustainable Development, University of Brasília (UnB)—Campus Universitário Darcy Ribeiro, Asa Norte, Brasília-DF 70910-900, Brazil. ³⁹Wageningen Environmental Research, Wageningen University and Research, P.O. Box 47, 6700 AA Wageningen, Netherlands. ⁴⁰Natural Resources Institute, University of Greenwich, Central Avenue, Chatham Maritime, Kent ME44TB, UK. ⁴¹INRA, UR 1115, Plantes et Systèmes de culture Horticoles, 84000 Avignon, France. ⁴²Centre for Agri-Environmental Research, School of Agriculture, Policy and Development, Reading University, Reading RG6 6AR, UK. ⁴³Department of Entomology, University of Wisconsin, Madison, WI 53705, USA. ⁴⁴Instituto Nacional de Pesquisas da Amazônia (INPA), CEP 69.067-375 Manaus, Amazonas, Brazil. ⁴⁵Human-Environment Systems, Ecology, Evolution, and Behavior, Department of Biological Sciences, Boise State University, Boise, ID 83725, USA. ⁴⁶Department of Integrative Biology, University of Texas at Austin, 205 W 24th Street, 401 Biological Laboratories, Austin, TX 78712, USA. ⁴⁷Department of Biology and Environment, University of Haifa, Oranim, Tivon 36006, Israel. ⁴⁸Department of Entomology, Kansas State University, 125 Waters Hall, Manhattan, KS 66503, USA. ⁴⁹Department of Evolutionary and Environmental Biology, University of Haifa, 3498838 Haifa, Israel. ⁵⁰Institute for Plant Science and Microbiology, University of Hamburg, Hamburg, Germany. ⁵¹Biodiversity International, Bangalore 560 065, India. ⁵²Ashoka Trust for Research in Ecology and the Environment (ATREE), Bangalore, India. ⁵³Bren School of Environmental Science and Management, University of California, Santa Barbara, Santa Barbara, CA 93106-5131, USA. ⁵⁴Department of Environmental Studies, Seattle University, 901 12th Avenue, Seattle, WA 9812, USA. ⁵⁵Department of Botany and Biodiversity Research, Division of Conservation Biology, Vegetation Ecology and Landscape Ecology, University of Vienna, Rennweg 14, 1030 Vienna, Austria. ⁵⁶Department of Entomology and Nematology, University of Florida, 1881 Natural Area Drive, Gainesville, FL 32601, USA. ⁵⁷Agrosavia, Centro de Investigación Obonuco, Km 5 vía Obonuco, Pasto, Nariño, Colombia. ⁵⁸Agriculture, Livestock and Agroforestry Program, Tropical Agricultural Research and Higher Education Center (CATIE), Cartago, Turrialba 30501, Costa Rica. ⁵⁹National Audubon Society, Boulder, CO 80305, USA. ⁶⁰Institute for Resources, Environment and Sustainability, University of British Columbia, Vancouver, BC, Canada. ⁶¹Cátedra de Avicultura, Cunicultura y Apicultura, Facultad de Agronomía, Universidad de Buenos Aires, CABA C1417DSE, Argentina. ⁶²School of Plant and Environmental Sciences, Virginia Tech, Blacksburg, VA 24061, USA. ⁶³Department of Ecology, UnB—Campus Universitário Darcy Ribeiro, Brasília-DF 70910-900, Brazil. ⁶⁴Department of Entomology and Nematology, University of California, Davis, Davis, CA 95616, USA. ⁶⁵INRA, UMR 1065 Santé et Agroécologie du Vignoble, ISVV, Université de Bordeaux, Bordeaux Sciences Agro, F-33883 Villenave d'Ornon Cedex, France. ⁶⁶INIBIOMA, Universidad Nacional del Comahue, CONICET, Quintral 1250, 8400 Bariloche, Río Negro, Argentina. ⁶⁷Senckenberg Biodiversity and Climate Research Centre (SBK-F), Senckenberganlage 25, 60325 Frankfurt am Main, Germany. ⁶⁸Centre for Biodiversity and Biosecurity, University of Auckland, Auckland, New Zealand. ⁶⁹Department of Environmental Science, Policy, and Management, University of California Berkeley, Berkeley, CA, USA. ⁷⁰South African National Biodiversity Institute, Kirstenbosch Research Centre, Private Bag X7, Claremont 7735, South Africa. ⁷¹School of Agriculture and Food Science and Earth Institute, University College Dublin, Belfield, Dublin 4, Ireland. ⁷²School of Natural Sciences, Trinity College Dublin, Dublin 2, Ireland. ⁷³Institute for Sustainable Agro-ecosystem Services, School of Agriculture and Life Sciences, The University of Tokyo, 188-0002 Tokyo, Japan. ⁷⁴Forestry and Forest Products Research Institute, 1 Matsunosato, Tsukuba, Ibaraki 305-8687, Japan. ⁷⁵Instituto de Biologia, Universidade Federal da Bahia, 40170-210 Salvador, Brazil. ⁷⁶Bio-Protection Research Centre, Lincoln University, Lincoln, New Zealand. ⁷⁷Fukushima Branch, National Institute for Environmental Studies, 963-770 Fukushima, Japan. ⁷⁸Environment and Production Technology Division, International Food Policy Research Institute, Washington, DC 20005, USA. ⁷⁹Department of Health and Environmental Sciences, Xi'an Jiaotong-Liverpool University, 215123, Suzhou, China.

*Corresponding author. Email: matteo.dainese@eurac.edu

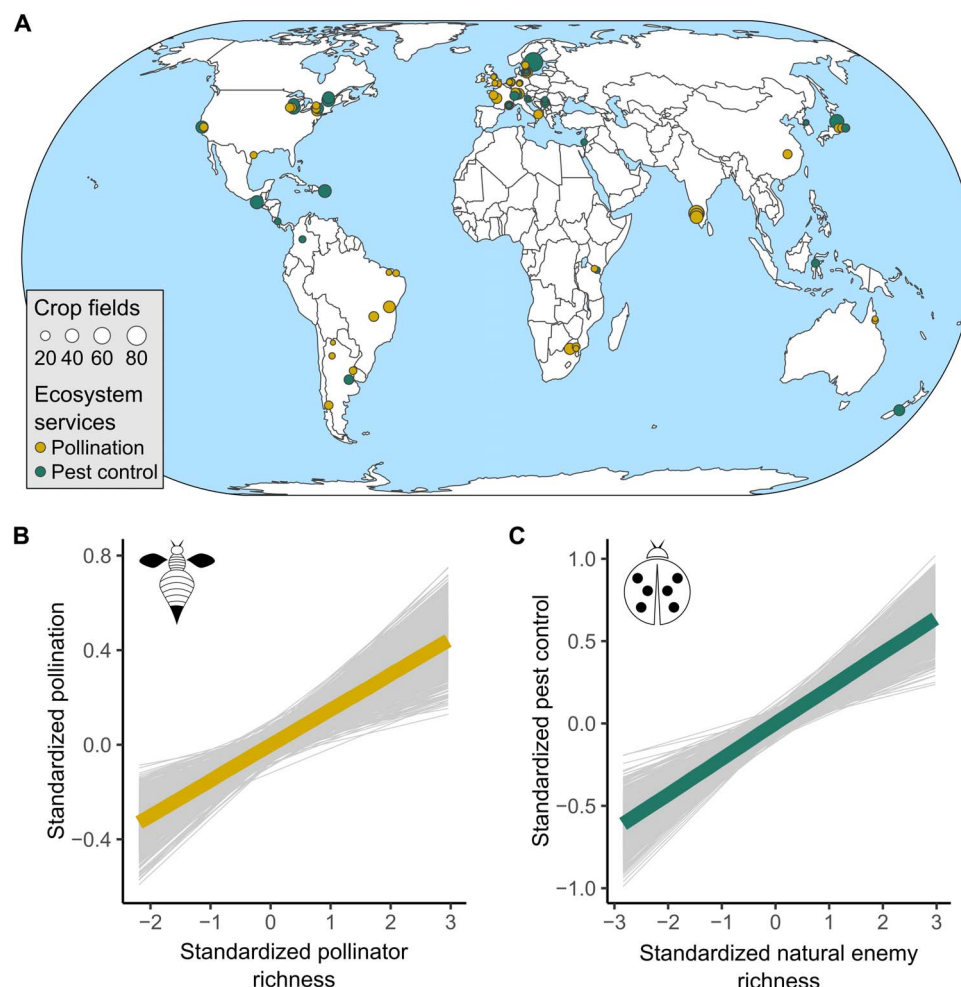


Fig. 1. Distribution of analyzed studies and effects of richness on ecosystem services provisioning. (A) Map showing the size (number of crop fields sampled) and location of the 89 studies (further details of studies are given in table S1). (B) Global effect of pollinator richness on pollination ($n = 821$ fields of 52 studies). (C) Global effect of natural enemy richness on pest control ($n = 654$ fields of 37 studies). The thick line in each plot represents the median of the posterior distribution of the model. Light gray lines represent 1000 random draws from the posterior. The lines are included to depict uncertainty of the modeled relationship.

dominance) as the E_{var} index (27) that reflects the relative abundance of those taxa. We derived a standardized index of pollination services using measures of pollination success (fruit set or seed set) and plant reproduction (pollen limitation) and of pest control services using measures of natural enemy activity (predation or parasitism rates) and crop damage (pest activity) (see Materials and Methods). We z -transformed each measure separately to remove the effect of differences in measurement scale between indices and inverted values for those measures where low values indicate positive contribution to ecosystem service (i.e., pest activity). We also characterized the 1-km-radius landscape surrounding each field by measuring the percentage of cropland from high-resolution land-use maps. This landscape metric has been used as a relevant proxy for characterizing landscape simplification (28, 29) and is often correlated with other indicators of landscape complexity (30, 31).

Using a Bayesian multilevel modeling approach, we addressed three fundamental yet unresolved questions in the biodiversity-ecosystem function framework: (i) Do richness, abundance, and dominance sustain ecosystem services in real-world ecosystems? (ii) Does landscape simplification indirectly affect ecosystem services mediated by a loss

of local community diversity? (iii) How strong are the cascading effects of landscape simplification on final crop production?

RESULTS AND DISCUSSION

We found clear evidence that richness of service-providing organisms positively influenced ecosystem service delivery. This was detected for both pollination (Fig. 1B and table S2) and pest control (Fig. 1C and table S2) and in almost all studies (figs. S1 and S2). As different methods were used in different studies to quantify richness and ecosystem services, we tested the sensitivity of our results to methodological differences. The bivariate relationships between richness and ecosystem services were robust to the taxonomic resolution to which each organism was identified, the sampling methods used to collect pollinators and natural enemies, and the pollination and pest control service measures used (see Materials and Methods).

The positive contribution of richness to service supply was also confirmed in a series of path analyses where we partitioned the relative importance of richness, total abundance, and evenness in driving biodiversity-ecosystem services relationships (Fig. 2). Although richness

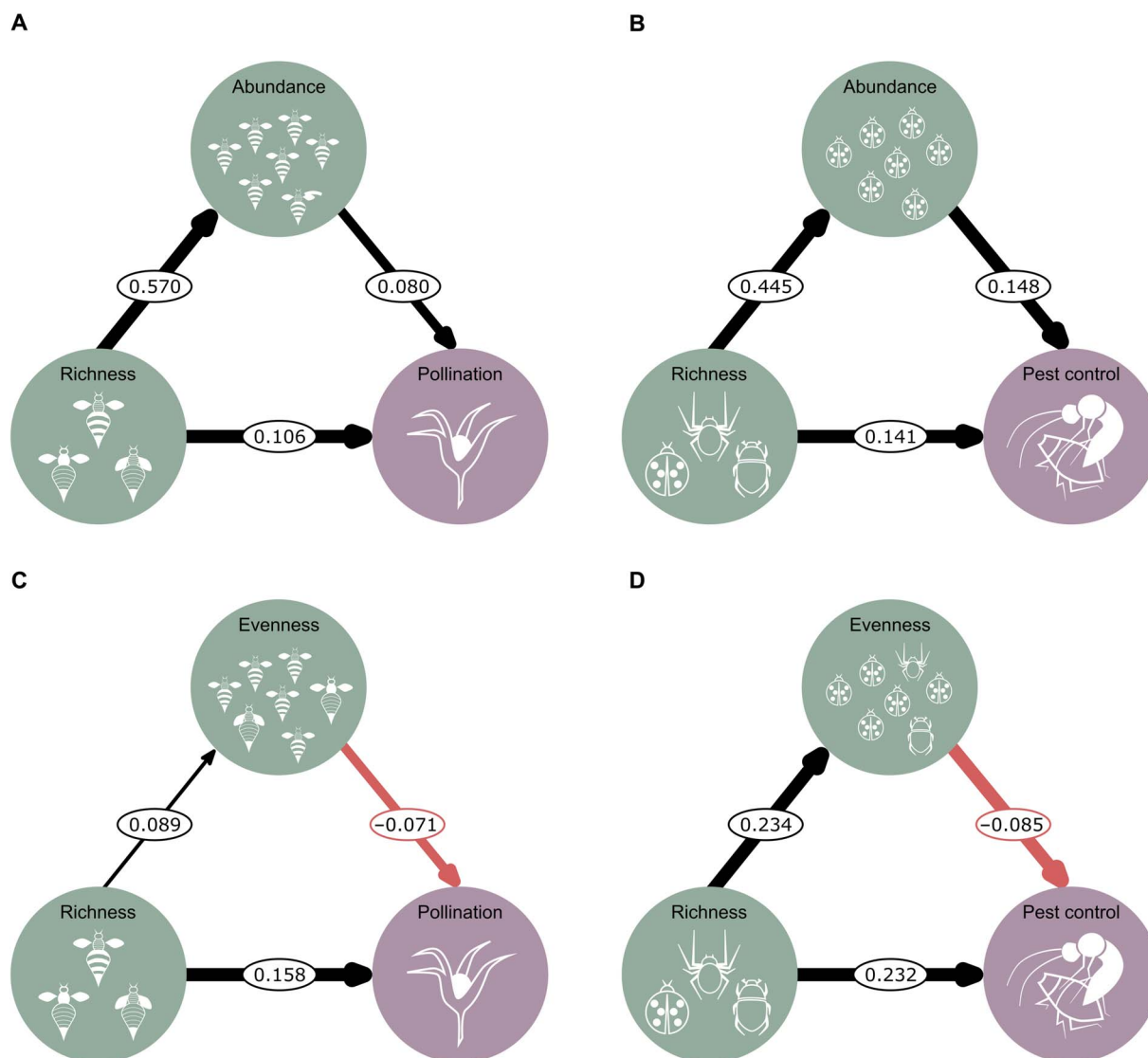


Fig. 2. Direct and indirect effects of richness, total abundance, and evenness on ecosystem services. (A) Path model of pollinator richness as a predictor of pollination, mediated by pollinator abundance. (B) Path model of natural enemy richness as a predictor of pest control, mediated by natural enemy abundance. (C) Path model of pollinator richness as a predictor of pollination, mediated by pollinator evenness. (D) Path model of natural enemy richness as a predictor of pest control, mediated by natural enemy evenness. Pollination model, $n = 821$ fields of 52 studies; pest control model, $n = 654$ fields of 37 studies. Path coefficients are effect sizes estimated from the median of the posterior distribution of the model. Black and red arrows represent positive or negative effects, respectively. Arrow widths are proportional to highest density intervals (HDIs).

showed the larger contribution in both models, we also found a direct effect of total abundance (Fig. 2, A and B) and evenness (Fig. 2, C and D) on ecosystem service delivery. While both richness and total abundance showed a positive effect, evenness had a negative effect, suggesting that dominant species contribute more strongly to service supply. This integrative assessment of different aspects of community structure revealed a more multilayered relationship between biodiversity and ecosystem services than has been previously acknowledged. Our results complement previous findings for pollination (7, 21, 32) and pest control (33) and indicate that richness and total and relative abundance are not mutually exclusive but concurrently contribute to support these two key ecosystem services to agriculture. Hence, we find strong support for the role of species-rich communities in supporting pollination and pest control services.

Furthermore, we found that landscape simplification indirectly affected ecosystem services by reducing the richness of service-providing organisms. Roughly a third of the negative effects of landscape simplification on pollination were due to a loss in pollinator richness (Fig. 3A and table S5). This effect was even greater for pest control where natural enemy richness mediated about 50% of the total effect of landscape simplification (Fig. 3B and table S5). A similar pattern was also found considering abundance in addition to richness. In this case, landscape simplification indirectly affected ecosystem services by reducing both richness and abundance of service-providing organisms (fig. S4, A and B, and table S6). A consistent richness-mediated effect was also confirmed when we tested the direct and indirect effects of landscape simplification on ecosystem services via changes in both richness and evenness (fig. S4, C and D, and table S7). However, contrary to our ex-

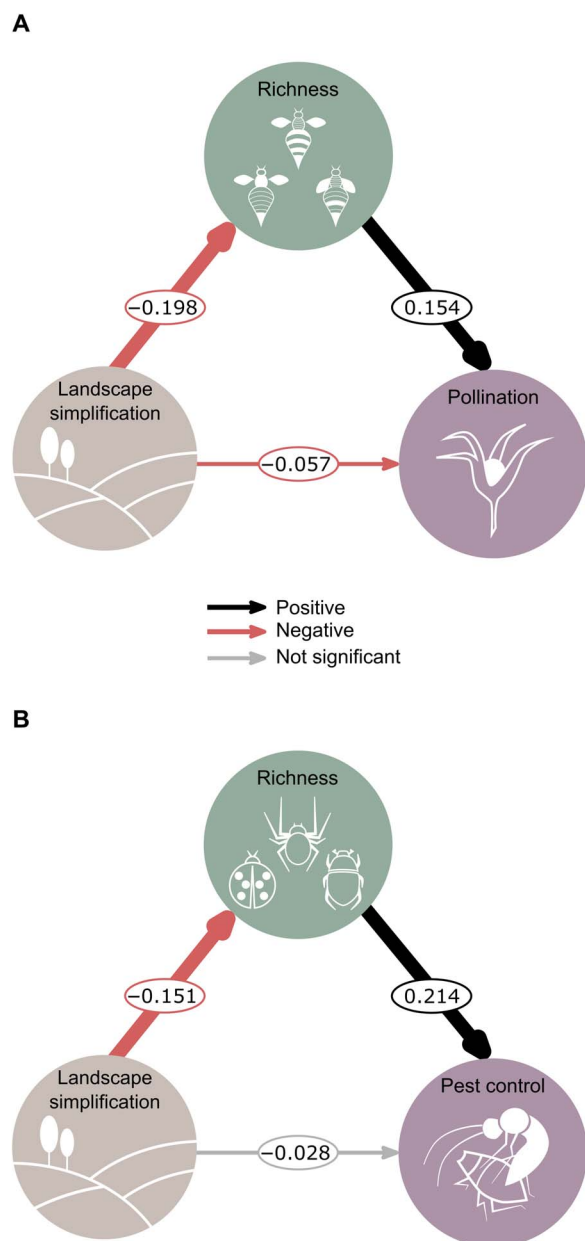


Fig. 3. Direct and indirect effects of landscape simplification on richness of service-providing organisms and associated ecosystem services. (A) Path model of landscape simplification as a predictor of pollination, mediated by pollinator richness ($n = 821$ fields of 52 studies). (B) Path model of landscape simplification as a predictor of pest control, mediated by natural enemy richness ($n = 654$ fields of 37 studies). Path coefficients are effect sizes estimated from the median of the posterior distribution of the model. Black and red arrows represent positive and negative effects, respectively. Arrow widths are proportional to HDIs. Gray arrows represent nonsignificant effects (HDIs overlapped zero).

pectation, landscape simplification led to higher pollinator evenness. It is likely that landscape simplification has stronger effects on more specialized, rare species and shifts assemblages toward more evenly abundant, mobile, generalist species with a higher ability to use scattered resources in the landscape (34, 35). These findings, combined with the larger contribution of richness when tested in combination with evenness, suggest that not only dominant species but also relatively rare spe-

cies contributed to pollination services (32, 36). The effect of landscape simplification on ecosystem services was minimized when not considering the mediated effect of richness, especially for pest control. We did not find a direct landscape simplification effect on pest control [all highest density intervals (HDIs) overlapped zero; Fig. 3B and table S5]. Together, these results demonstrate strong negative indirect effects of landscape simplification on biodiversity-driven ecosystem services in agroecosystems and the importance of the richness and abundance of service-providing organisms in mediating these effects.

Last, for a subset of the data that had crop production information (674 fields of 42 studies), we found that the cascading effects of landscape simplification, mediated through richness and associated ecosystem services, led to lower crop production. This was detected for both pollination (Fig. 4A and table S8) and pest control (Fig. 4B and table S8). Specifically, landscape simplification reduced both pollinator and natural enemy richness, which had indirect consequences for pollination and pest control and, in turn, decreased crop production. For pest control, a positive link with crop production was detected in fields where the study area was not sprayed with insecticides during the course of the experiment (Fig. 4B) but not when considering all sites combined (with and without insecticide use; fig. S4). In sprayed areas, we did not find a pest control effect (all HDIs overlapped zero), probably because effects were masked by insecticide use (37, 38), indicating that insecticide use undermines the full potential of natural pest control. A positive link with crop production was detected, although measures used to estimate pest control (natural enemy and pest activity) were not direct components of crop production, as was the case of pollination measures (fruit or seed set). We also found an indirect influence of dominance (Fig. 4 and table S8) but not of abundance (fig. S4 and table S9) on crop production. However, the indirect dominance pathway was weaker than that via richness. An abundance effect may not have been detectable because of the lower sample size in these submodels with crop production. Although only available from a subset of the data, this result supports the hypothesis that the effects of landscape simplification can cascade up to reducing the final provisioning service of crop production.

Our findings suggest that some previously inconsistent responses of natural enemy abundance and activity to surrounding landscape composition (39) can be reconciled by considering richness in addition to abundance. Although richness and abundance are often correlated, their response to environmental variation can differ. This was evident in the path analysis showing a strong effect of landscape simplification on richness but only a marginal effect on abundance (fig. S3B). Moreover, effect sizes for natural enemy abundances in individual studies (fig. S5) showed similarly inconsistent responses to the previous synthesis (39). Likewise, results for natural enemy activity showed inconsistent responses to surrounding landscape composition in both syntheses [Fig. 3B and (39)].

Using an integrative model to assess key ecological theory, we demonstrate that the negative effects of landscape simplification on service supply and final crop production are primarily mediated by loss of species. We found strong evidence for positive biodiversity–ecosystem service relationships, highlighting that managing landscapes to enhance the richness of service-providing organisms (40, 41) is a promising pathway toward a more sustainable food production globally. In an era of rapid environmental changes, preserving biodiversity-driven services will consistently confer greater resilience to agroecosystems, such that we could expect improved crop production under a broader range of potential future conditions.

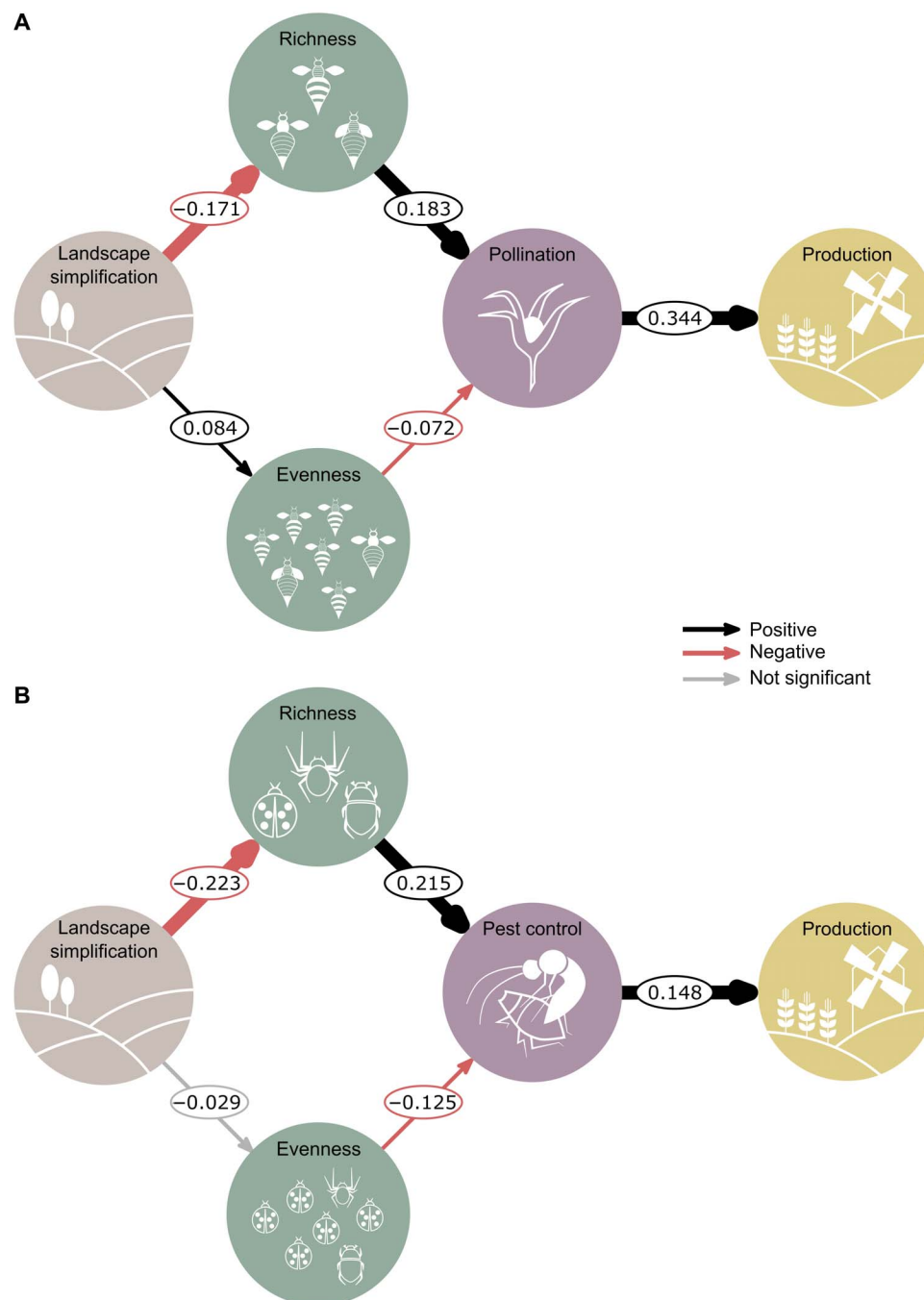


Fig. 4. Direct and cascading effects of landscape simplification on final crop production via changes in richness, evenness, and ecosystem services. (A) Path model representing direct and indirect effects of landscape simplification on final crop production through changes in pollinator richness, evenness, and pollination ($n = 438$ fields of 27 studies). (B) Path model representing direct and indirect effects of landscape simplification on final crop production through changes in natural enemy richness, evenness, and pest control [only insecticide-free areas were considered in the model ($n = 185$ fields of 14 studies)]. Path coefficients are effect sizes estimated from the median of the posterior distribution of the model. Black and red arrows represent positive and negative effects, respectively. Arrow widths are proportional to HDIs. Gray arrows represent nonsignificant effects (HDIs overlapped zero).

MATERIALS AND METHODS

Database compilation

We compiled data from crop studies where measures of richness and abundance of service-providing organisms (pollinators or natural enemies) and associated ecosystem services (pollination and biological

pest control) were available for the same sites. If available, we also included information on yield. Studies were identified by first searching the reference lists of recent meta-analyses (7, 21, 39, 42, 43) and then directly contacting researchers. For pest control, data were mostly provided from a recent pest control database (39). Of 191 researchers

initially contacted, 86 provided data that met our criteria. As similar studies were frequently performed in the same area, occasionally in the same year, and studies with multiple years usually used different sites each year, we did not nest year within study. Likewise, some studies collected data in different crops. Thus, we considered each year of multi-year studies (that is, 10 studies) and each crop of multicrop studies (that is, 5 studies) to be an independent dataset and used study-crop-year combinations as the highest hierarchical unit. Overall, we analyzed data from 89 studies and 1475 fields in 27 countries around the world (table S1). Twenty-nine crops were considered, including a wide array of annual and perennial fruit, seed, nut, stimulant, pulse, cereal, and oilseed crops. Studies represented the spectrum of management practices, including conventional, low-input conventional, integrated pest management, and organic farming. Few studies compared different management practices in similar landscapes, crop types, or sampled year. In 76% of fields, pest control experiments were performed in insecticide-free areas. In some fields, this information was not available (7%) or insecticides were applied (17%).

Pollinator and pest natural enemy richness and abundance

Studies used a broad range of methods, which we categorized as active or passive (44), to sample pollinators or natural enemies. Active sampling methods included netting pollinators seen on crop flowers, hand-collecting individuals on plants, observational counting, sweep-netting, and vacuum sampling. Passive sampling methods were malaise traps, pan traps, pitfall traps, and sticky cards. Active sampling was performed in 85% of pollinator sampling fields and in 50% of natural enemy sampling fields.

Pollinators included representatives from the orders Hymenoptera, Diptera, Lepidoptera, and Coleoptera. Bees (Hymenoptera: Apoidea) were the most commonly observed pollinators and included *Apis* bees (Apidae: *Apis mellifera*, *Apis cerana*, *Apis dorsata*, and *Apis florea*), stingless bees (Apidae: Meliponini), bumblebees (Apidae: *Bombus* spp.), carpenter bees (Apidae: Xylocopini), small carpenter bees (Apidae: Ceratinini), sweat bees (Halictidae), long-horned bees (Apidae: Eucerini), plasterer bees (Colletidae), mining bees (Andrenidae), and mason bees (Megachilidae). Non-bee taxa included syrphid flies (Diptera: Syrphidae), other flies (Diptera: Calliphoridae, Tachinidae, and Muscidae), butterflies and moths (Lepidoptera), various beetle families (Coleoptera), and hymenopterans including ants (Formicidae) and the paraphyletic group of non-bee aculeate wasps. Natural enemies included ground beetles (Coleoptera), flies (Diptera), spiders (Aranea), hymenopterans including ants (Formicidae) and wasps, bugs (Hemiptera), thrips (Thysanoptera), net-winged insects (Neuroptera), bats, and birds.

We calculated pollinator and natural enemy richness as the number of unique taxa sampled per study, method, and field. A taxon was defined as a single biological type (that is, species, morphospecies, genus, and family) determined at the finest taxonomic resolution to which each organism was identified. In almost 70% of cases, taxonomic resolution was to species level (averaged proportion among all studies), but sometimes, it was based on morphospecies (15%), genus (8%), or family levels (7%). Taxon richness per field varied between 1 and 49 for pollinators and between 1 and 40 for natural enemies. Abundance reflected the sum of individuals sampled per study, method, and field. Pollinator richness was calculated either including or excluding honey bees (*A. mellifera*). *A. mellifera* was considered as the only species within the honey bee group for consistency across all datasets (43). Other *Apis* bees (that is, *A. cerana*, *A. dorsata*, and *A. florea*) were not pooled into the honey bee category as the large majority of observed individ-

uals were derived from feral populations. Feral and managed honey bees were analyzed as a single group because they cannot be distinguished during field observations. Feral honey bees were uncommon in most studies except for those in Africa and South America (*A. mellifera* is native in Africa, while it was introduced to the Americas). In studies with subsamples within a field (that is, plots within fields or multiple sampling rounds within fields), we calculated the total number of individuals and unique taxa across these subsamples.

We also calculated evenness using the E_{var} index (27). This index is based on the variance of log abundances (centered on the mean of log abundances) and then appropriately scaled to cover 0 to 1 (0, maximally uneven and 1, perfectly even).

Pollination and pest control services

As different methods were used to quantify pollination or pest control services across studies, standardization was necessary to put all the indices on equivalent terms. Therefore, we transformed each index y in each field i in each study j using z scores. We preferred the use of z scores over other transformations (for example, division by the maximum) because z scores do not constrain the variability found in the raw data, as do other indices that are bounded between 0 and 1. We used the proportion of flowers that set fruit (that is, fruit set), the average number of seeds per fruits (that is, seed set), or the estimated measures of pollinator contribution to plant reproduction (that is, differences in fruit weight between plants with and without insect pollination, hereafter Δ fruit weight) as measures of pollination services. We then converted these measures into the pollination index. The pest control index was calculated using measures of natural enemy activity or pest activity. Natural enemy activity was measured by sentinel pest experiments where pests were placed in crop fields and predation or parasitism rates were monitored or by field enclosure experiments where cages were used to exclude natural enemies to quantify differences in pest abundance or crop damage between plants with and without natural enemies. Pest activity was measured as the fraction or amount of each crop consumed, infested, or damaged. We inverted standardized values of pest activity by multiplying by -1 , as low values indicate positive contributions to the ecosystem service.

Crop production

Depending on the crop type, marketable crop yield is valued by farmers not only in terms of area-based yield but also in terms of fruit or seed weight [for example, in coffee, sunflower, or strawberry fields; (45, 46)] or seed production per plant [for example, in seed production fields; (36)]. Moreover, area-based yield and within-plant yield are often correlated (35, 36). Thus, we used both area-based yield and within-plant yield as measures of final crop production. Within-plant yield was measured by the total number (or mass) of seeds or fruits per plant or by fruit or seed weight. In addition, in this case, we standardized variables (z scores) to put all the indices on equivalent terms.

Landscape simplification

Landscapes were characterized by calculating the percentage of cropland (annual and perennial) within a radius of 1 km around the center of each crop field. This landscape metric has been used as a relevant proxy for characterizing landscape simplification (28, 29) and is often correlated with other indicators of landscape complexity (30, 31). Moreover, we used this metric because cropland data are readily accessible from publicly available land cover data and are more accurate than other land use types such as forests and grasslands (47), especially when

detailed maps are not available. The 1-km spatial extent was chosen to reflect the typical flight and foraging distances of many insects including pollinators (48, 49) and natural enemies (50, 51). For studies where this information was not supplied by the authors, land uses were digitized using GlobeLand30 (52), a high-resolution map of Earth's land cover. The derived land cover maps were verified and, if necessary, corrected using a visual inspection of satellite images (Google Earth). We then calculated the percentage of cropland within the radius using Quantum GIS 2.18 (Open Source Geospatial Foundation Project; <http://qgis.osgeo.org>). The average percentage of cropland was 67.5% for pollination studies and 41.5% for natural enemy studies.

Data analysis

Data standardization

Before performing the analyses, we standardized the predictors (abundance, richness, and landscape simplification) using z scores within each study. This standardization was necessary to allow comparisons between studies with differences in methodology and landscape ranges (53). Moreover, this allows the separation of within-study effects from between-study effects. This separation is important because it prevents the risk of misinterpreting the results based on studies differing in methodology and landscape gradients, by erroneously extrapolating from between-study effects to within-study effects or vice versa (54, 55).

Relationship between richness and ecosystem services

The relationship between richness of service-providing organisms and related ecosystem services (Fig. 1, B and C) was estimated from a Bayesian multilevel (partial pooling) model that allowed the intercept and the slope to vary among studies (also commonly referred to as random intercepts and slopes), following the equation

$$ES_i \sim N(\alpha_{j[i]} + \beta_{j[i]} RIC_i, \sigma_j)$$

$$\alpha_j \sim N(\mu_\alpha, \sigma_\alpha)$$

$$\beta_j \sim N(\mu_\beta, \sigma_\beta)$$

where ES_i is the ecosystem service index (pollination or pest control depending on the model), RIC_i is richness of service-providing organisms (pollinator or natural enemy richness depending on the model), and $j_{[i]}$ represents observation i of study j . This partial-pooling model estimated both study-level responses [yielding an estimate for each study (β_j)] and the distribution from which the study-level estimates were drawn, yielding a higher-level estimate of the overall response across crop systems (μ_β). In addition, it accounted for variation in variance and sample size across observations (for example, studies). The intercepts α_j and slopes β_j varied between studies according to a normal distribution with mean μ and SD σ . Independent within-study errors also followed a normal distribution $\varepsilon_i \sim N(0, \sigma)$. We used weakly informative priors: normal (0, 10) for the population-level parameters (α, β) and half-Student t (3, 0, 5) for the group-level SD and residual SD.

Direct and indirect effects of richness, abundance, and evenness on service provisioning

As natural communities vary not only in number of species but also in relative abundance of each species (evenness) and the total number of individuals (abundance), it is important to incorporate these attributes when assessing or modeling biodiversity effects (12, 56). In a Bayesian multivariate response model, a form of path analysis, we partitioned

the relative importance of richness and total and relative abundance in driving biodiversity-ecosystem relationships. We hypothesized that richness drives both abundance and evenness according to a revised version of the “more individuals hypothesis” (57) that proposes that more species can exploit more diverse resources and may therefore maintain more individuals than species-poor communities. Specifically, we tested (i) whether richness *per se* directly influences ecosystem services or is instead mediated by abundance (Eq. 1, 3; Fig. 2, A and B) and (ii) whether richness *per se* directly influences ecosystem services or is instead mediated by evenness (Eq. 2, 4; Fig. 2, C and D). Before analysis, we also checked for data collinearity among abundance, evenness, and richness by calculating the variance inflation factor (VIF). No signal of collinearity was detected in either model (VIFs were below 1.8). In a preliminary analysis, we also tested a possible interaction between richness and evenness. No significant interaction was found in both pollination and pest control models. To illustrate, we first show the univariate multilevel (partial pooling) models following these equations

$$RIC_i \sim N(\alpha_{j[i]} + \beta_{j[i]} ABU_i, \sigma_j) \quad (1)$$

$$RIC_i \sim N(\alpha_{j[i]} + \beta_{j[i]} EVE_i, \sigma_j) \quad (2)$$

$$ES_i \sim N(\alpha_{j[i]} + \beta_{j[i]} RIC_i + \beta_{j[i]} ABU_i, \sigma_j) \quad (3)$$

$$ES_i \sim N(\alpha_{j[i]} + \beta_{j[i]} RIC_i + \beta_{j[i]} EVE_i, \sigma_j) \quad (4)$$

where RIC_i is richness of service-providing organisms, ABU_i is abundance, EVE_i is evenness, ES_i is the ecosystem service index, and the index $j_{[i]}$ represents observation i of study j . We then specified both multivariate multilevel models in a matrix-vector notation (53), as follows

$$Y_i \sim N(X_i B_{r[i]}, \Sigma_j)$$

$$B_r \sim N(M_B, \Sigma_B)$$

where Y_i is the matrix of response variables with observations i as rows and variables r as columns, X_i is the matrix of all predictors for response r , B_r are the regression parameters (α and β) for response r , M_B represents the mean of the distribution of the regression parameters, and Σ_B is the covariance matrix representing the variation of the regression parameters in the population groups. We used weakly informative priors: normal (0, 10) for the population-level parameters (α, β) and half-Student t (3, 0, 5) for the group-level SD and residual SD. In building the model, we ensured that no residual correlation between ES_i and RIC_i , between ES_i and EVE_i , or between ES_i and ABU_i was estimated [see the “set_rescor” function in the package *brms*; (58)].

Direct and indirect effects of landscape simplification on ecosystem services

To estimate the direct and indirect effects of landscape simplification on richness and associated ecosystem services, we used two models. In a Bayesian multivariate response model with causal mediation effects (hereafter, mediation model), we tested whether landscape simplification directly influences ecosystem services or is mediated by richness. Mediation analysis is a statistical procedure to test whether the effect of an independent variable X on a dependent variable Y ($X \rightarrow Y$) is at least partly explained via the inclusion of a third hypothetical

variable, the mediator variable M ($X \rightarrow M \rightarrow Y$) (59). The three causal paths a , b , and c' correspond to X 's effect on M , M 's effect on Y , and X 's effect on Y accounting for M , respectively. The three causal paths correspond to parameters from two regression models, one in which M is the outcome and X is the predictor and one in which Y is the outcome and X and M are the simultaneous predictors (fig. S6). From these parameters, we can compute the mediation effect (the product ab , also known as the indirect effect) and the total effect of X on Y

$$c = c' + ab$$

Thus, the total causal effect of X , which is captured by the parameter c , can be decomposed precisely into two components, a direct effect c' and an indirect (mediation) effect ab (the product of paths a and b). The model included the ecosystem service index as response, landscape simplification as predictor, and richness as mediator (Fig. 3). The separate regression models that made up the Bayesian multivariate multilevel model followed these equations

$$\text{RIC}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{LAND}_i, \sigma_j)$$

$$\text{ES}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{RIC}_i + \beta_{j[i]} \text{LAND}_i, \sigma_j)$$

We then compiled a multilevel path analysis testing the direct and indirect effects of landscape simplification on ecosystem services via changes in both richness and abundance (Eq. 5, 6, 8; fig. S4, A and B) or richness and evenness (Eq. 5, 7, 9; fig. S4, C and D). The separate regression models that made up the model followed these equations

$$\text{RIC}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{LAND}_i, \sigma_j) \quad (5)$$

$$\text{ABU}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{LAND}_i, \sigma_j) \quad (6)$$

$$\text{EVE}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{LAND}_i, \sigma_j) \quad (7)$$

$$\text{ES}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{RIC}_i + \beta_{j[i]} \text{ABU}_i + \beta_{j[i]} \text{LAND}_i, \sigma_j) \quad (8)$$

$$\text{ES}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{RIC}_i + \beta_{j[i]} \text{EVE}_i + \beta_{j[i]} \text{LAND}_i, \sigma_j) \quad (9)$$

where RIC_i is richness of service-providing organisms, LAND_i is landscape simplification measured as the percentage of arable land surrounding each study site, ABU_i is abundance, EVE_i is evenness, ES_i is the ecosystem service index, and the index $j_{[i]}$ represents observation i of study j . We then specified multivariate multilevel models in a matrix-vector notion, as explained above. The mediation analysis was implemented using the R package `sjstats` [v. 0.15.0; (60)].

Cascading effects of landscape simplification on final crop production

For 42 studies and 675 fields (pollination model, $n = 438$ fields of 27 studies; pest control model, $n = 236$ fields of 15 studies; table S1), the data allowed us to use a multilevel path analysis to examine cascading effects of landscape simplification on final crop produc-

tion via changes in richness, abundance, evenness, and ecosystem services. We expected that (i) landscape simplification would have a direct effect on richness, abundance, and dominance of service-providing organisms and (ii) richness and abundance dominance would relate positively to intermediate services, which, in turn, (iii) would increase final crop production (Fig. 4 and fig. S4). The indirect effects of richness and abundance (Eq. 10, 11, 13, 15; Fig. 4) or richness and evenness (Eq. 10, 12, 14, 15; fig. S4) were tested separately

$$\text{RIC}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{LAND}_i, \sigma_j) \quad (10)$$

$$\text{ABU}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{LAND}_i, \sigma_j) \quad (11)$$

$$\text{EVE}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{LAND}_i, \sigma_j) \quad (12)$$

$$\text{ES}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{RIC}_i + \beta_{j[i]} \text{ABU}_i, \sigma_j) \quad (13)$$

$$\text{ES}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{RIC}_i + \beta_{j[i]} \text{EVE}_i, \sigma_j) \quad (14)$$

$$\text{PROD}_i \sim N(\alpha_{j[i]} + \beta_{j[i]} \text{ES}_i, \sigma_j) \quad (15)$$

where RIC_i is richness of service-providing organisms, LAND_i is landscape simplification measured as the percentage of arable land surrounding each study site, ABU_i is abundance, EVE_i is evenness, ES_i is the ecosystem service index, PROD_i is crop production, and the index $j_{[i]}$ represents observation i of study j . We specified a multivariate multilevel model in a matrix-vector notion, as explained above.

Parameter estimation

All analyses were conducted in the programming language Stan through R (v. 3.4.3) using the package `brms` [v. 2.2.0; (58)]. Stan implements Hamiltonian Monte Carlo and its extension, the No-U-Turn Sampler. These algorithms converge much more quickly than other Markov chain Monte Carlo algorithms especially for high-dimensional models (61). Each model was run with four independent Markov chains of 5000 iterations, discarding the first 2500 iterations per chain as warm-up and resulting in 10,000 posterior samples overall. Convergence of the four chains and sufficient sampling of posterior distributions were confirmed by (i) the visual inspection of parameter traces, (ii) ensuring a scale reduction factor (\hat{R}) below 1.01, and (iii) an effective size (n_{eff}) of at least 10% of the number of iterations. For each model, posterior samples were summarized on the basis of the Bayesian point estimate (median), SE (median absolute deviation), and posterior uncertainty intervals by HDIs, a type of credible interval that contains the required mass such that all points within the interval have a higher probability density than points outside the interval (62). The advantage of the Bayesian approach is the possibility to estimate not only expected values for each parameter but also the uncertainty associated with these estimates (63). Thus, we calculated 80, 90, and 95% HDIs for parameter estimates.

Sensitivity analyses

Given that different methods were used in different studies to quantify richness, ecosystem services, and final crop production, we measured the sensitivity of our results to methodological differences.

(1) We verified whether treating each annual dataset from multiyear studies separately could incorrectly account for the dependence of the

data. We refitted the model testing the relationship between richness and ecosystem services including year nested within study (that is, study defined as study-crop combination). Then, we compared models (year-independent model versus year-nested model) using leave-one-out (LOO) cross-validation, a fully Bayesian model selection procedure for estimating pointwise out-of-sample prediction accuracy (64). We calculated the expected log pointwise predictive density ($\widehat{\text{elpd}}_{\text{loo}}$), using the log-likelihood evaluated at the posterior simulations of the parameter values. Model comparison was implemented using R package loo [v. 2.0.0; (65)]. We found that the year-nested model had a lower average predictive accuracy than the year-independent model for both pollination ($\Delta\widehat{\text{elpd}}_{\text{loo}} = -1.79$) and pest control ($\Delta\widehat{\text{elpd}}_{\text{loo}} = -1.09$) and therefore retained the year-independent model in our analysis.

(2) We verified whether taxonomic resolution influenced the interpretation of results. We recalculated richness considering only organisms classified at the fine taxonomy level (species or morphospecies levels) and refitted the model testing the effect of richness on ecosystem services. We found no evidence that taxonomic resolution influenced our results. With a fine taxonomic resolution, the effects of richness on ecosystem services ($\beta_{\text{pollinators}} = 0.1535$, 90% HDIs = 0.0967 to 0.2141; $\beta_{\text{enemies}} = 0.2264$, 90% HDIs = 0.1475 to 0.3065; table S2) were nearly identical to the estimates presented in the main text ($\beta_{\text{pollinators}} = 0.1532$, 90% HDIs = 0.0892 to 0.2058; $\beta_{\text{enemies}} = 0.2093$, 90% HDIs = 0.1451 to 0.2779; table S2).

(3) We verified whether the sampling methods used to collect pollinators (active versus passive sampling techniques) influenced the relationship between pollinator richness and pollination using Bayesian hypothesis testing (58). Passive methods do not directly capture flower visitors and may introduce some bias (for example, they may underestimate flower visitors). However, our estimate was not influenced by sampling method (the one-sided 90% credibility interval overlapped zero; table S11). In accordance with this finding, the evidence ratio showed that the hypothesis tested (that is, estimates of studies with active sampling > estimates of studies with passive sampling) was only 0.78 times more likely than the alternative hypothesis.

(4) We verified whether methodological differences in measuring pollination and pest control services influenced the relationship between richness and ecosystem services. Using Bayesian hypothesis testing, we tested whether the estimates differed among methods. The two-sided 95% credibility interval overlapped zero in all comparisons (estimates did not differ significantly; table S12), indicating that our estimate was not influenced by methodological differences in measuring ecosystem services. Furthermore, we tested effects including only inverted pest activity as a reflection of pest control. We found positive effects of natural enemy richness on inverted pest activity ($\beta = 0.1307$, 90% HDIs = 0.0102 to 0.2456), indicating that results were robust to the type of pest control measure considered.

(5) As honey bees are the most important and abundant flower visitors in some locations, we verified the potential influence of honey bees on our results by refitting the path models testing direct and indirect effects of richness, abundance, and evenness on pollination services with honey bees. A positive direct contribution of richness to pollination was confirmed even after including honey bees (fig. S7). In these models, both abundance and evenness showed a larger effect compared to models without honey bees (Fig. 2).

(6) Insecticide application during the course of the experiment could mask the effect of pest control on crop production (37, 38). We verified the potential influence of insecticide application on our results by refit-

ting the model considering only fields where the study area was not sprayed with insecticide during the course of the experiment ($n = 85$ fields of 14 studies). We found a pest control effect that was masked when considering all sites combined (with and without insecticide; fig. S4). We therefore show the insecticide-free model in the main text (Fig. 4).

(7) We verified the consistency of our results considering only studies that measured area-based yield (submodel). Only the cascading effects of landscape simplification on final crop production via changes in richness were tested in a simplified model. We found no evident differences between the submodel (fig. S8) and the full model presented in the main text (Fig. 4).

SUPPLEMENTARY MATERIALS

Supplementary material for this article is available at <http://advances.sciencemag.org/cgi/content/full/5/10/eaax0121/DC1>

Supplementary Text

Fig. S1. Forest plot of the effect of pollinator richness on pollination for individual studies.

Fig. S2. Forest plot of the effect of natural enemy richness on pest control for individual studies.

Fig. S3. Direct and indirect landscape simplification effects on ecosystem services via changes in richness and abundance or richness and evenness.

Fig. S4. Direct and cascading landscape simplification effects on final crop production via changes in natural enemy richness, abundance evenness, and pest control (all sites together, with and without insecticide application).

Fig. S5. Forest plot of the effect of landscape simplification on natural enemy abundance for individual studies.

Fig. S6. Mediation model.

Fig. S7. Direct and indirect effects of pollinator richness, abundance, and evenness (with honey bees) on pollination.

Fig. S8. Direct and cascading landscape simplification effects on area-based yield via changes in richness, and ecosystem services.

Table S1. List of 89 studies considered in our analyses.

Table S2. Model output for richness-ecosystem service relationships.

Table S3. Model output for path models testing direct and indirect effects (mediated by changes in abundance) of richness on ecosystem services.

Table S4. Model output for path models testing direct and indirect effects (mediated by changes in evenness) of richness on ecosystem services.

Table S5. Model output for path models testing direct and indirect effects (mediated by changes in richness) of landscape simplification on ecosystem services.

Table S6. Model output for path models testing the direct and cascading landscape simplification effects on ecosystem services via changes in richness and abundance.

Table S7. Model output for path models testing the direct and cascading landscape simplification effects on ecosystem services via changes in richness and evenness.

Table S8. Model output for path models testing the direct and cascading landscape simplification effects on final crop production via changes in richness, evenness, and ecosystem services.

Table S9. Model output for path models testing the direct and cascading landscape simplification effects on final crop production via changes in richness, abundance, and ecosystem services.

Table S10. Model output for path models testing direct and indirect effects of pollinator richness, abundance, and evenness (with honey bees) on pollination.

Table S11. Results of pairwise comparison of richness-ecosystem service relationships according to the methods used to sample pollinators and natural enemies.

Table S12. Results of pairwise comparison of richness-ecosystem service relationships according to the methods used to quantify pollination and pest control services.

Database S1. Data on pollinator and natural enemy diversity and associated ecosystem services compiled from 89 studies and 1475 locations around the world.

References (66–111)

[View/request a protocol for this paper from Bio-protocol.](#)

REFERENCES AND NOTES

1. S. Díaz, U. Pascual, M. Stenseke, B. Martín-López, R. T. Watson, Z. Molnár, R. Hill, K. M. A. Chan, I. A. Baste, K. A. Brauman, S. Polasky, A. Church, M. Lonsdale, A. Larigauderie, P. W. Leadley, A. P. E. van Oudenhoven, F. van der Plaats, M. Schröter, S. Lavorel, Y. Aumeeruddy-Thomas, E. Bukvareva, K. Davies, S. Demissew, G. Erp, P. Failler, C. A. Guerra, C. L. Hewitt, H. Keune, S. Lindley, Y. Shirayama, Assessing nature's contributions to people. *Science* **359**, 270–272 (2018).

2. S. Díaz, J. Settele, E. Brondizio, Summary for policymakers of the global assessment report on biodiversity and ecosystem services of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services (United Nations Paris, Fr., 2019), 1–39.
3. S. Naeem, J. E. Duffy, E. Zavaleta, The functions of biological diversity in an age of extinction. *Science* **336**, 1401–1406 (2012).
4. D. Tilman, F. Isbell, J. M. Cowles, Biodiversity and ecosystem functioning. *Annu. Rev. Ecol. Syst.* **45**, 471–493 (2014).
5. B. J. Cardinale, J. E. Duffy, A. Gonzalez, D. U. Hooper, C. Perrings, P. Venail, A. Narwani, G. M. Mace, D. Tilman, D. A. Wardle, A. P. Kinzig, G. C. Daily, M. Loreau, J. B. Grace, A. Larigauderie, D. S. Srivastava, S. Naeem, Biodiversity loss and its impact on humanity. *Nature* **486**, 59–67 (2012).
6. R. Winfree, J. W. Fox, N. M. Williams, J. R. Reilly, D. P. Cariveau, Abundance of common species, not species richness, drives delivery of a real-world ecosystem service. *Ecol. Lett.* **18**, 626–635 (2015).
7. D. Kleijn, R. Winfree, I. Bartomeus, L. G. Carvalheiro, M. Henry, R. Isaacs, A.-M. Klein, C. Kremen, L. K. M'Gonigle, R. Rader, T. H. Ricketts, N. M. Williams, N. Lee Adamson, J. S. Ascher, A. Báldi, P. Batáry, F. Benjamin, J. C. Biesmeijer, E. J. Blitzer, R. Bommarco, M. R. Brand, V. Bretagnolle, L. Button, D. P. Cariveau, R. Chifflet, J. F. Colville, B. N. Danforth, E. Elle, M. P. D. Garratt, F. Herzog, A. Holzschuh, B. G. Howlett, F. Jauker, S. Jha, E. Knop, K. M. Krewenka, V. le Féon, Y. Mandelik, E. A. May, M. G. Park, G. Pisanty, M. Reemer, V. Riedinger, O. Rollin, M. Rundlöf, H. S. Sardiñas, J. Scheper, A. R. Sciligo, H. G. Smith, I. Steffan-Dewenter, R. Thorp, T. Tscharntke, J. Verhulst, B. F. Viana, B. E. Vaissière, R. Veldtman, K. L. Ward, C. Westphal, S. G. Potts, Delivery of crop pollination services is an insufficient argument for wild pollinator conservation. *Nat. Commun.* **6**, 7414 (2015).
8. M. Loreau, A. Hector, Partitioning selection and complementarity in biodiversity experiments. *Nature* **412**, 72–76 (2001).
9. D. U. Hooper, F. S. Chapin III, J. J. Ewel, A. Hector, P. Inchausti, S. Lavorel, J. H. Lawton, D. M. Lodge, M. Loreau, S. Naeem, B. Schmid, H. Setälä, A. J. Symstad, J. Vandermeer, D. A. Wardle, Effects of biodiversity on ecosystem functioning: A consensus of current knowledge. *Ecol. Monogr.* **75**, 3–35 (2005).
10. M. Schleuning, J. Fründ, D. García, Predicting ecosystem functions from biodiversity and mutualistic networks: An extension of trait-based concepts to plant-animal interactions. *Ecography* **38**, 380–392 (2015).
11. T. H. Larsen, N. M. Williams, C. Kremen, Extinction order and altered community structure rapidly disrupt ecosystem functioning. *Ecol. Lett.* **8**, 538–547 (2005).
12. J. Reiss, J. R. Bridle, J. M. Montoya, G. Woodward, Emerging horizons in biodiversity and ecosystem functioning research. *Trends Ecol. Evol.* **24**, 505–514 (2009).
13. K. H. Bannar-Martin, C. T. Kremer, S. K. M. Ernest, M. A. Leibold, H. Auge, J. Chase, S. A. J. Declerck, N. Eisenhauer, S. Harpole, H. Hillebrand, F. Isbell, T. Koffel, S. Larsen, A. Narwani, J. S. Petermann, C. Roscher, J. S. Cabral, S. R. Supp, Integrating community assembly and biodiversity to better understand ecosystem function: The community assembly and the functioning of ecosystems (CAFE) approach. *Ecol. Lett.* **21**, 167–180 (2018).
14. P. Balvanera, I. Siddique, L. Dee, A. Paquette, F. Isbell, A. Gonzalez, J. Byrnes, M. I. O'Connor, B. A. Hungate, J. N. Griffin, Linking biodiversity and ecosystem services: Current uncertainties and the necessary next steps. *Bioscience* **64**, 49–57 (2014).
15. F. Isbell, A. Gonzalez, M. Loreau, J. Cowles, S. Díaz, A. Hector, G. M. Mace, D. A. Wardle, M. I. O'Connor, J. E. Duffy, L. A. Turnbull, P. L. Thompson, A. Larigauderie, Linking the influence and dependence of people on biodiversity across scales. *Nature* **546**, 65–72 (2017).
16. R. Chaplin-Kramer, M. E. O'Rourke, E. J. Blitzer, C. Kremen, A meta-analysis of crop pest and natural enemy response to landscape complexity. *Ecol. Lett.* **14**, 922–932 (2011).
17. C. M. Kennedy, E. Lonsdorf, M. C. Neel, N. M. Williams, T. H. Ricketts, R. Winfree, R. Bommarco, C. Brittain, A. L. Burley, D. Cariveau, L. G. Carvalheiro, N. P. Chacoff, S. A. Cunningham, B. N. Danforth, J.-H. Dudenhöffer, E. Elle, H. R. Gaines, L. A. Garibaldi, C. Gratton, A. Holzschuh, R. Isaacs, S. K. Javorek, S. Jha, A. M. Klein, K. Krewenka, Y. Mandelik, M. M. Mayfield, L. Morandin, L. A. Neame, M. Otieno, M. Park, S. G. Potts, M. Rundlöf, A. Saez, I. Steffan-Dewenter, H. Taki, B. F. Viana, C. Westphal, J. K. Wilson, S. A. Greenleaf, C. Kremen, A global quantitative synthesis of local and landscape effects on wild bee pollinators in agroecosystems. *Ecol. Lett.* **16**, 584–599 (2013).
18. J. A. Foley, R. DeFries, G. P. Asner, C. Barford, G. Bonan, S. R. Carpenter, F. S. Chapin, M. T. Coe, G. C. Daily, H. K. Gibbs, J. H. Helkowski, T. Holloway, E. A. Howard, C. J. Kucharik, C. Monfreda, J. A. Patz, I. C. Prentice, N. Ramankutty, P. K. Snyder, Global consequences of land use. *Science* **309**, 570–574 (2005).
19. T. H. Ricketts, J. Regetz, I. Steffan-Dewenter, S. A. Cunningham, C. Kremen, A. Bogdanski, B. Gemmill-Herren, S. S. Greenleaf, A. M. Klein, M. M. Mayfield, L. A. Morandin, A. Ochieng', S. G. Potts, B. F. Viana, Landscape effects on crop pollination services: Are there general patterns? *Ecol. Lett.* **11**, 499–515 (2008).
20. F. J. J. A. Bianchi, C. J. H. Booij, T. Tscharntke, Sustainable pest regulation in agricultural landscapes: A review on landscape composition, biodiversity and natural pest control. *Proc. Biol. Sci.* **273**, 1715–1727 (2006).
21. L. A. Garibaldi, I. Steffan-Dewenter, R. Winfree, M. A. Aizen, R. Bommarco, S. A. Cunningham, C. Kremen, L. G. Carvalheiro, L. D. Harder, O. Afik, I. Bartomeus, F. Benjamin, V. Boreux, D. Cariveau, N. P. Chacoff, J. H. Dudenhöffer, B. M. Freitas, J. Ghazoul, S. Greenleaf, J. Hipolito, A. Holzschuh, B. Howlett, R. Isaacs, S. K. Javorek, C. M. Kennedy, K. M. Krewenka, S. Krishnan, Y. Mandelik, M. M. Mayfield, I. Motzke, T. Munyuli, B. A. Nault, M. Otieno, J. Petersen, G. Pisanty, S. G. Potts, R. Rader, T. H. Ricketts, M. Rundlöf, C. L. Seymour, C. Schuepp, H. Szentgyorgyi, H. Taki, T. Tscharntke, C. H. Vergara, B. F. Viana, T. C. Wanger, C. Westphal, N. Williams, A. M. Klein, Wild pollinators enhance fruit set of crops regardless of honey bee abundance. *Science* **339**, 1608–1611 (2013).
22. D. K. Ray, N. Ramankutty, N. D. Mueller, P. C. West, J. A. Foley, Recent patterns of crop yield growth and stagnation. *Nat. Commun.* **3**, 1293 (2012).
23. J. A. Foley, N. Ramankutty, K. A. Brauman, E. S. Cassidy, J. S. Gerber, M. Johnston, N. D. Mueller, C. O'Connell, D. K. Ray, P. C. West, C. Balzer, E. M. Bennett, S. R. Carpenter, J. Hill, C. Monfreda, S. Polasky, J. Rockström, J. Sheehan, S. Siebert, D. Tilman, D. P. M. Zaks, Solutions for a cultivated planet. *Nature* **478**, 337–342 (2011).
24. R. Bommarco, D. Kleijn, S. G. Potts, Ecological intensification: Harnessing ecosystem services for food security. *Trends Ecol. Evol.* **28**, 230–238 (2013).
25. J. Pretty, Intensification for redesigned and sustainable agricultural systems. *Science* **362**, eaav0294 (2018).
26. Millennium Ecosystem Assessment, *Ecosystems and Human Well-being: Biodiversity Synthesis* (World Resources Institute, 2005).
27. B. Smith, J. B. Wilson, A consumer's guide to evenness indices. *Oikos* **76**, 70–82 (1996).
28. T. D. Meehan, B. P. Werling, D. A. Landis, C. Gratton, Agricultural landscape simplification and insecticide use in the Midwestern United States. *Proc. Natl. Acad. Sci. U. S. A.* **108**, 11500–11505 (2011).
29. M. Dainese, N. J. B. Isaac, G. D. Powney, R. Bommarco, E. Öckinger, M. Kuussaari, J. Pöyry, T. G. Benton, D. Gabriel, J. A. Hodgson, W. E. Kunin, R. Lindborg, S. M. Sait, L. Marini, Landscape simplification weakens the association between terrestrial producer and consumer diversity in Europe. *Glob. Chang. Biol.* **23**, 3040–3051 (2017).
30. T. Tscharntke, A. M. Klein, A. Kruess, I. Steffan-Dewenter, C. Thies, Landscape perspectives on agricultural intensification and biodiversity—Ecosystem service management. *Ecol. Lett.* **8**, 857–874 (2005).
31. D. A. Landis, Designing agricultural landscapes for biodiversity-based ecosystem services. *Basic Appl. Ecol.* **18**, 1–12 (2017).
32. R. Winfree, J. R. Reilly, I. Bartomeus, D. P. Cariveau, N. M. Williams, J. Gibbs, Species turnover promotes the importance of bee diversity for crop pollination at regional scales. *Science* **359**, 791–793 (2018).
33. D. K. Letourneau, J. A. Jedlicka, S. G. Bothwell, C. R. Moreno, Effects of natural enemy biodiversity on the suppression of arthropod herbivores in terrestrial ecosystems. *Annu. Rev. Ecol. Syst.* **40**, 573–592 (2009).
34. L. Marini, E. Öckinger, K.-O. Bergman, B. Jauker, J. Krauss, M. Kuussaari, J. Pöyry, H. G. Smith, I. Steffan-Dewenter, R. Bommarco, Contrasting effects of habitat area and connectivity on evenness of pollinator communities. *Ecography* **37**, 544–551 (2014).
35. S. Gámez-Virués, D. J. Perović, M. M. Gossner, C. Börschig, N. Blüthgen, H. de Jong, N. K. Simons, A.-M. Klein, J. Krauss, G. Maier, C. Scherber, J. Steckel, C. Rothenwöhrer, I. Steffan-Dewenter, C. N. Weiner, W. Weisser, M. Werner, T. Tscharntke, C. Westphal, Landscape simplification filters species traits and drives biotic homogenization. *Nat. Commun.* **6**, 8568 (2015).
36. T. P. M. Fijen, J. A. Scheper, T. M. Boom, N. Janssen, I. Raemakers, D. Kleijn, Insect pollination is at least as important for marketable crop yield as plant quality in a seed crop. *Ecol. Lett.* **21**, 1704–1713 (2018).
37. H. Liere, T. N. Kim, B. P. Werling, T. D. Meehan, D. A. Landis, C. Gratton, Trophic cascades in agricultural landscapes: Indirect effects of landscape composition on crop yield. *Ecol. Appl.* **25**, 652–661 (2015).
38. V. Gagic, D. Kleijn, A. Báldi, G. Boros, H. B. Jørgensen, Z. Elek, M. P. D. Garratt, G. A. de Groot, K. Hedlund, A. Kovács-Hostyánszki, L. Marini, E. Martin, I. Peveré, S. G. Potts, S. Redlich, D. Senapathi, I. Steffan-Dewenter, S. Świtek, H. G. Smith, V. Takács, P. Tryjanowski, W. H. van der Putten, S. van Gils, R. Bommarco, Combined effects of agrochemicals and ecosystem services on crop yield across Europe. *Ecol. Lett.* **20**, 1427–1436 (2017).
39. D. S. Karp, R. Chaplin-Kramer, T. D. Meehan, E. A. Martin, F. DeClerck, H. Grab, C. Gratton, L. Hunt, A. E. Larsen, A. Martínez-Salinas, M. E. O'Rourke, A. Rusch, K. Poveda, M. Jonsson, J. A. Rosenheim, N. A. Schellhorn, T. Tscharntke, S. D. Wratten, W. Zhang, A. L. Iverson, L. S. Adler, M. Albrecht, A. Alignier, G. M. Angelella, M. Z. Anjum, J. Avelino, P. Batáry, J. M. Baveco, F. J. J. A. Bianchi, K. Birkhofer, E. W. Bohnenblust, R. Bommarco, M. J. Brewer, B. Caballero-López, Y. Carrière, L. G. Carvalheiro, L. Cayuela, M. Centrella, A. Četković, D. C. Henri, A. Chabert, A. C. Costamagna, A. De la Mora, J. de Kraker, N. Desneux, E. Diehl, T. Diekötter, C. F. Dormann, J. O. Eckberg, M. H. Entling, D. Fiedler, P. Franck, F. J. F. van Veen,

- T. Frank, V. Gagic, M. P. D. Garratt, A. Getachew, D. J. Gonthier, P. B. Goodell, I. Graziosi, R. L. Groves, G. M. Gurr, Z. Hajian-Forooshani, G. E. Heimpel, J. D. Hermann, A. S. Hueseth, D. J. Inčlán, A. J. Ingrao, P. Iv, K. Jacot, G. A. Johnson, L. Jones, M. Kaiser, J. M. Kaser, T. Keasar, T. N. Kim, M. Kishinevsky, D. A. Landis, B. Lavandero, C. Lavigne, A. Le Ralec, D. Lemessa, D. K. Letourneau, H. Lieke, Y. Lu, Y. Lubin, T. Luttermoser, B. Maas, K. Mace, F. Madeira, V. Mader, A. M. Cortesero, L. Marini, E. Martinez, H. M. Martinson, P. Menozzi, M. G. E. Mitchell, T. Miyashita, G. A. R. Molina, M. A. Molina-Montenegro, M. E. O'Neal, I. Opatovsky, S. Ortiz-Martinez, M. Nash, Ö. Östman, A. Ouin, D. Pak, D. Paredes, S. Parsa, H. Parry, R. Perez-Alvarez, D. J. Perović, J. A. Peterson, S. Petit, S. M. Philpott, M. Plantegenest, M. Plečáš, T. Pluess, X. Pons, S. G. Potts, R. F. Pywell, D. W. Ragsdale, T. A. Rand, L. Raymond, B. Ricci, C. Sargent, J. P. Sarthou, J. Saulais, J. Schäckermann, N. P. Schmidt, G. Schneider, C. Schüepp, F. S. Sivakoff, H. G. Smith, K. S. Whitney, S. Stutz, Z. Szendrei, M. B. Takada, H. Taki, G. Tamburini, L. J. Thomson, Y. Tricault, N. Tsafak, M. Tschumi, M. Valentin-Morison, M. Van Trinh, W. van der Werf, K. T. Vierling, B. J. Werling, J. B. Wickens, V. J. Wickens, B. A. Woodcock, K. Woychievsky, H. Xiao, M. Yasuda, A. Yoshioka, Y. Zou, Crop pests and predators exhibit inconsistent responses to surrounding landscape composition. *Proc. Natl. Acad. Sci. U. S. A.* **15**, E7863–E7870 (2018).
40. C. Kremen, A. M. Merenlender, Landscapes that work for biodiversity and people. *Science* **362**, eaau6020 (2018).
41. E. A. Martin, M. Dainese, Y. Clough, A. Baldi, R. Bommarco, V. Gagic, M. P. D. Garratt, A. Holzschuh, D. Kleijn, A. Kovács-Hostyánszki, L. Marini, S. G. Potts, H. G. Smith, D. Al Hassan, M. Albrecht, G. K. S. Andersson, J. D. Asís, S. Aviron, M. B. Balzan, L. Baños-Picón, I. Bartomeus, P. Batáry, F. Burel, B. Caballero-López, E. D. Concepción, V. Coudrain, J. Dänhardt, M. Diaz, T. Diekötter, C. F. Dormann, R. Duflo, M. H. Entling, N. Farwig, C. Fischer, T. Frank, L. A. Garibaldi, J. Hermann, F. Herzog, D. Inčlán, K. Jacot, F. Jauber, P. Jeanneret, M. Kaiser, J. Krauss, V. Le Féon, J. Marshall, A.-C. Moonen, G. Moreno, V. Riedinger, M. Rundlöf, A. Rusch, J. Scheper, G. Schneider, C. Schüepp, S. Stutz, L. Sutter, G. Tamburini, C. Thies, J. Tormos, T. Tschamtké, M. Tschumi, D. Uzman, C. Wagner, M. Zubair-Anjum, I. Steffan-Dewenter, The interplay of landscape composition and configuration: New pathways to manage functional biodiversity and agroecosystem services across Europe. *Ecol. Lett.* **22**, 1083–1094 (2019).
42. L. A. Garibaldi, L. G. Carvalheiro, B. E. Vaissiere, B. Gemmill-Herren, J. Hipolito, B. M. Freitas, H. T. Ngo, N. Azzu, A. Saez, J. Astrom, J. An, B. Blochtein, D. Buchori, F. J. C. Garcia, F. Oliveira da Silva, K. Devkota, M. de Fátima Ribeiro, L. Freitas, M. C. Gaglianone, M. Goss, M. Irshad, M. Kasina, A. J. S. P. Filho, L. H. P. Kiill, P. Kwapong, G. N. Parra, C. Pires, V. Pires, R. S. Rawal, R. Rizali, A. M. Saraiva, R. Veldtman, B. F. Viana, S. Witter, H. Zhang, Mutually beneficial pollinator diversity and crop yield outcomes in small and large farms. *Science* **351**, 388–391 (2016).
43. R. Rader, I. Bartomeus, L. A. Garibaldi, M. P. D. Garratt, B. G. Howlett, R. Winfree, S. A. Cunningham, M. M. Mayfield, A. D. Arthur, G. K. S. Andersson, R. Bommarco, C. Brittain, L. G. Carvalheiro, N. P. Chacoff, M. H. Entling, B. Fouly, B. M. Freitas, B. Gemmill-Herren, J. Ghazoul, S. R. Griffin, C. L. Gross, L. Herbertsson, F. Herzog, J. Hipólito, S. Jaggar, F. Jauber, A. M. Klein, D. Kleijn, S. Krishnan, C. Q. Lemos, S. A. M. Lindström, Y. Mandelik, V. M. Monteiro, W. Nelson, L. Nilsson, D. E. Pattemore, N. de O. Pereira, G. Pisanty, S. G. Potts, M. Reemer, M. Rundlöf, C. S. Sheffield, J. Scheper, C. Schüepp, H. G. Smith, D. A. Stanley, J. C. Stout, H. Szentgyörgyi, H. Taki, C. H. Vergara, B. F. Viana, M. Woyciechowski, Non-bee insects are important contributors to global crop pollination. *Proc. Natl. Acad. Sci. U. S. A.* **113**, 146–151 (2016).
44. E. M. Lichtenberg, C. M. Kennedy, C. Kremen, P. Batáry, F. Berendse, R. Bommarco, N. A. Bosque-Pérez, L. G. Carvalheiro, W. E. Snyder, N. M. Williams, R. Winfree, B. K. Klatt, S. Åström, F. Benjamin, C. Brittain, R. Chaplin-Kramer, Y. Clough, B. Danforth, T. Diekötter, S. D. Eigenbrode, J. Ekroos, E. Elle, B. M. Freitas, Y. Fukuda, H. R. Gaines-Day, H. Grab, C. Gratton, A. Holzschuh, R. Isaacs, M. Isaia, S. Jha, D. Jonason, V. P. Jones, A. M. Klein, J. Krauss, D. K. Letourneau, S. Macfadyen, R. E. Mallinger, E. A. Martin, E. Martinez, J. Memmott, L. Morandin, L. Neame, M. Otieno, M. G. Park, L. Pfiffner, M. J. O. Pocock, C. Ponce, S. G. Potts, K. Poveda, M. Ramos, J. A. Rosenheim, M. Rundlöf, H. Sardiñas, M. E. Saunders, N. L. Schon, A. R. Scilligo, C. S. Sidhu, I. Steffan-Dewenter, T. Tschamtké, M. Vesely, W. W. Weisser, J. K. Wilson, D. W. Crowder, A global synthesis of the effects of diversified farming systems on arthropod diversity within fields and across agricultural landscapes. *Glob. Chang. Biol.* **23**, 4946–4957 (2017).
45. L. G. Carvalheiro, R. Veldtman, A. G. Shenkute, G. B. Tesfay, C. W. W. Pirk, J. S. Donaldson, S. W. Nicolson, Natural and within-farmland biodiversity enhances crop productivity. *Ecol. Lett.* **14**, 251–259 (2011).
46. V. Boreux, C. G. Kusalappa, P. Vaast, J. Ghazoul, Interactive effects among ecosystem services and management practices on crop production: Pollination in coffee agroforestry systems. *Proc. Natl. Acad. Sci. U.S.A.* **110**, 8387–8392 (2013).
47. R. Tropek, O. Sedláček, J. Beck, P. Keil, Z. Musilova, I. Šimova, D. Storch, Comment on “High-resolution global maps of 21st-century forest cover change”. *Science* **344**, 981 (2014).
48. M. Rundlöf, J. Bengtsson, H. G. Smith, Local and landscape effects of organic farming on butterfly species richness and abundance. *J. Appl. Ecol.* **45**, 813–820 (2008).
49. A. Holzschuh, M. Dainese, J. P. González-Varo, S. Mudri-Stojnić, V. Riedinger, M. Rundlöf, J. Scheper, J. B. Wickens, V. J. Wickens, R. Bommarco, D. Kleijn, S. G. Potts, S. P. M. Roberts, H. G. Smith, M. Vilà, A. Vujić, I. Steffan-Dewenter, Mass-flowering crops dilute pollinator abundance in agricultural landscapes across Europe. *Ecol. Lett.* **19**, 1228–1236 (2016).
50. C. Thies, T. Tschamtké, Landscape structure and biological control in agroecosystems. *Science* **285**, 893–895 (1999).
51. A. Rusch, R. Chaplin-Kramer, M. M. Gardiner, V. Hawro, J. Holland, D. Landis, C. Thies, T. Tschamtké, W. W. Weisser, C. Winqvist, M. Woltz, R. Bommarco, Agricultural landscape simplification reduces natural pest control: A quantitative synthesis. *Agric. Ecosyst. Environ.* **221**, 198–204 (2016).
52. J. Chen, Y. Ban, S. Li, China: Open access to Earth land-cover map. *Nature* **514**, 434 (2014).
53. A. Gelman, J. Hill, *Data Analysis Using Regression and Multilevel Hierarchical Models* (Cambridge Univ. Press, 2007).
54. M. van de Pol, J. Wright, A simple method for distinguishing within- versus between-subject effects using mixed models. *Anim. Behav.* **77**, 753–758 (2009).
55. H. Schielzeth, Simple means to improve the interpretability of regression coefficients. *Methods Ecol. Evol.* **1**, 103–113 (2010).
56. B. J. Cardinale, D. S. Srivastava, J. Emmett Duffy, J. P. Wright, A. L. Downing, M. Sankaran, C. Jouseau, Effects of biodiversity on the functioning of trophic groups and ecosystems. *Nature* **443**, 989–992 (2006).
57. P.-C. Bürkner, brms: An R package for Bayesian multilevel models using Stan. *J. Stat. Softw.* **80**, 1–27 (2017).
58. D. P. MacKinnon, A. J. Fairchild, M. S. Fritz, Mediation analysis. *Annu. Rev. Psychol.* **58**, 593–614 (2007).
59. D. Lüdtke, sjstats: Statistical functions for regression models. 10.5281/ZENODO.1308979 (2018).
60. B. Carpenter, A. Gelman, M. D. Hoffman, D. Lee, B. Goodrich, M. Betancourt, M. Brubaker, J. Guo, P. Li, A. Riddell, Stan: A probabilistic programming language. *J. Stat. Softw.* **76**, 1–37 (2017).
61. J. Kruschke, *Doing Bayesian Data Analysis: A Tutorial with R, JAGS, and Stan* (Academic Press, ed. 2, 2015).
62. R. McElreath, *Statistical Rethinking: A Bayesian Course with Examples in R and Stan* (CRC Press, 2016).
63. A. Vehtari, A. Gelman, J. Gabry, Practical Bayesian model evaluation using leave-one-out cross-validation and WAIC. *Stat. Comput.* **27**, 1413–1432 (2017).
64. A. Vehtari, A. Gelman, J. Gabry, Y. Yao, A. Gelman, loo: Efficient leave-one-out cross-validation and WAIC for Bayesian models. R package version, 2.0.0 2018; <https://cran.r-project.org/package=loo>.
65. U. Samnegård, G. Alins, V. Boreux, J. Bosch, D. García, A. K. Happe, A. M. Klein, M. Miñarro, K. Mody, M. Porcel, A. Rodrigo, L. Roquer-Bení, M. Tasin, P. A. Hambäck, Management trade-offs on ecosystem services in apple orchards across Europe: Direct and indirect effects of organic production. *J. Appl. Ecol.* **56**, 802–811 (2019).
66. M. P. D. Garratt, C. L. Truslove, D. J. Coston, R. L. Evans, E. D. Moss, C. Dodson, N. Jenner, J. C. Biesmeijer, S. G. Potts, Pollination deficits in UK apple orchards. *J. Pollinat. Ecol.* **12**, 9–14 (2014).
67. G. A. de Groot, R. van Kats, M. Reemer, D. van der Sterren, J. C. Biesmeijer, D. Kleijn, “De bijdrage van (wilde) bestuivers aan de opbrengst van appels en blauwe bessen: Kwantificering van ecosystemdiensten in Nederland,” Alterra, Wageningen-UR, 2015; <http://edepot.wur.nl/353774>.
68. R. E. Mallinger, C. Gratton, Species richness of wild bees, but not the use of managed honeybees, increases fruit set of a pollinator-dependent crop. *J. Appl. Ecol.* **52**, 323–330 (2015).
69. M. P. D. Garratt, D. J. Coston, C. L. Truslove, M. G. Lappage, C. Polce, R. Dean, J. C. Biesmeijer, S. G. Potts, The identity of crop pollinators helps target conservation for improved ecosystem services. *Biol. Conserv.* **169**, 128–135 (2014).
70. D. d. L. Ramos, M. M. C. Bustamante, F. D. D. S. E. Silva, L. G. Carvalheiro, Crop fertilization affects pollination service provision—Common bean as a case study. *PLOS ONE* **13**, e0204460 (2018).
71. H. Taki, K. Okabe, S. Makino, Y. Yamaura, M. Sueyoshi, Contribution of small insects to pollination of common buckwheat, a distylous crop. *Ann. Appl. Biol.* **155**, 121–129 (2009).
72. H. Taki, K. Okabe, Y. Yamaura, T. Matsuura, M. Sueyoshi, S. Makino, K. Maeto, Effects of landscape metrics on *Apis* and non-*Apis* pollinators and seed set in common buckwheat. *Basic Appl. Ecol.* **11**, 594–602 (2010).
73. A. Holzschuh, J.-H. Dudenhöffer, T. Tschamtké, Landscapes with wild bee habitats enhance pollination, fruit set and yield of sweet cherry. *Biol. Conserv.* **153**, 101–107 (2012).
74. V. Boreux, S. Krishnan, K. G. Cheppudira, J. Ghazoul, Impact of forest fragments on bee visits and fruit set in rain-fed and irrigated coffee agro-forests. *Agric. Ecosyst. Environ.* **172**, 42–48 (2013).
75. V. Boreux, P. Vaast, L. P. Madappa, K. G. Cheppudira, C. Garcia, J. Ghazoul, Agroforestry coffee production increased by native shade trees, irrigation, and liming. *Agron. Sustain. Dev.* **36**, 10.1007/s13593-016-0377-7, (2016).

76. A. Classen, M. K. Peters, S. W. Ferger, M. Helbig-Bonitz, J. M. Schmack, G. Maassen, M. Schleuning, E. K. V. Kalko, K. Böhning-Gaese, I. Steffan-Dewenter, Complementary ecosystem services provided by pest predators and pollinators increase quantity and quality of coffee yields. *Proc. Biol. Sci.* **281**, 20133148 (2014).
77. J. Hipólito, D. Boscolo, B. F. Viana, Landscape and crop management strategies to conserve pollination services and increase yields in tropical coffee farms. *Agric. Ecosyst. Environ.* **256**, 218–225 (2018).
78. S. Krishnan, C. G. Kushalappa, R. U. Shaanker, J. Ghazoul, Status of pollinators and their efficiency in coffee fruit set in a fragmented landscape mosaic in South India. *Basic Appl. Ecol.* **13**, 277–285 (2012).
79. S. Krishnan, K. G. Cheppudira, J. Ghazoul, Pollinator services in coffee agroforests of the western Ghats, in *Agroforestry*, J. Dagar, V. Tewari, Eds. (Springer Singapore, 2017), pp. 771–795.
80. M. Nesper, C. Kueffer, S. Krishnan, C. G. Kushalappa, J. Ghazoul, Shade tree diversity enhances coffee production and quality in agroforestry systems in the Western Ghats. *Agric. Ecosyst. Environ.* **247**, 172–181 (2017).
81. S. Cusser, J. L. Neff, S. Jha, Natural land cover drives pollinator abundance and richness, leading to reductions in pollen limitation in cotton agroecosystems. *Agric. Ecosyst. Environ.* **226**, 33–42 (2016).
82. N. P. Chacoff, M. A. Aizen, Edge effects on flower-visiting insects in grapefruit plantations bordering premontane subtropical forest. *J. Appl. Ecol.* **43**, 18–27 (2006).
83. N. P. Chacoff, M. A. Aizen, V. Aschero, Proximity to forest edge does not affect crop production despite pollen limitation. *Proc. R. Soc. B Biol. Sci.* **275**, 907–913 (2008).
84. L. G. Carvalheiro, C. L. Seymour, R. Veldtman, S. W. Nicolson, Pollination services decline with distance from natural habitat even in biodiversity-rich areas. *J. Appl. Ecol.* **47**, 810–820 (2010).
85. L. G. Carvalheiro, C. L. Seymour, S. W. Nicolson, Creating patches of native flowers facilitates crop pollination in large agricultural fields: Mango as a case study. *J. Appl. Ecol.* **49**, 1373–1383 (2012).
86. I. Bartomeus, V. Gagic, R. Bommarco, Pollinators, pests and soil properties interactively shape oilseed rape yield. *Basic Appl. Ecol.* **16**, 737–745 (2015).
87. D. A. Stanley, D. Gunning, J. C. Stout, Pollinators and pollination of oilseed rape crops (*Brassica napus* L.) in Ireland: Ecological and economic incentives for pollinator conservation. *J. Insect Conserv.* **17**, 1181–1189 (2013).
88. D. A. Stanley, J. C. Stout, Pollinator sharing between mass-flowering oilseed rape and co-flowering wild plants: Implications for wild plant pollination. *Plant Ecol.* **215**, 315–325 (2014).
89. Y. Zou, F. J. A. Bianchi, F. Jauker, S. Luo, W. van der Werf, Wild pollinators enhance oilseed rape yield in small-holder farming systems in China. *BMC Ecol.* **17**, 6 (2017).
90. H. Connelly, K. Poveda, G. Loeb, Landscape simplification decreases wild bee pollination services to strawberry. *Agric. Ecosyst. Environ.* **211**, 51–56 (2015).
91. H. Grab, K. Poveda, B. Danforth, G. Loeb, Landscape context shifts the balance of costs and benefits from wildflower borders on multiple ecosystem services. *Proc. R. Soc. B Biol. Sci.* **285**, 20181102 (2018).
92. R. I. A. Stewart, G. K. S. Andersson, C. Brönmark, B. K. Klatt, L.-A. Hansson, V. Zülsdorff, H. G. Smith, Ecosystem services across the aquatic-terrestrial boundary: Linking ponds to pollination. *Basic Appl. Ecol.* **18**, 13–20 (2016).
93. B. Ricci, P. Franck, J.-F. Toubon, J.-C. Bouvier, B. Sauphanor, C. Lavigne, The influence of landscape on insect pest dynamics: A case study in southeastern France. *Landsc. Ecol.* **24**, 337–349 (2009).
94. M. Maaloulou, P. Franck, J.-C. Bouvier, J.-F. Toubon, C. Lavigne, Codling moth parasitism is affected by semi-natural habitats and agricultural practices at orchard and landscape levels. *Agric. Ecosyst. Environ.* **169**, 33–42 (2013).
95. B. Caballero-López, R. Bommarco, J. M. Blanco-Moreno, F. X. Sans, J. Pujade-Villar, M. Rundlöf, H. G. Smith, Aphids and their natural enemies are differently affected by habitat features at local and landscape scales. *Biol. Control.* **63**, 222–229 (2012).
96. G. Tamburini, S. De Simone, M. Sigura, F. Boscutti, L. Marini, Conservation tillage mitigates the negative effect of landscape simplification on biological control. *J. Appl. Ecol.* **53**, 233–241 (2016).
97. G. Tamburini, S. De Simone, M. Sigura, F. Boscutti, L. Marini, Soil management shapes ecosystem service provision and trade-offs in agricultural landscapes. *Proc. R. Soc. B Biol. Sci.* **283**, 20161369 (2016).
98. G. Tamburini, I. Pevero, N. Fornasini, S. de Simone, M. Sigura, F. Boscutti, L. Marini, Conservation tillage reduces the negative impact of urbanisation on carabid communities. *Insect Conserv. Divers.* **9**, 438–445 (2016).
99. H. Taki, K. Maeto, K. Okabe, N. Haruyama, Influences of the seminatural and natural matrix surrounding crop fields on aphid presence and aphid predator abundance within a complex landscape. *Agric. Ecosyst. Environ.* **179**, 87–93 (2013).
100. D. K. Letourneau, S. G. Bothwell Allen, J. O. Stireman III, Perennial habitat fragments, parasitoid diversity and parasitism in ephemeral crops. *J. Appl. Ecol.* **49**, 1405–1416 (2012).
101. B. Maas, Y. Clough, T. Tschamtkke, Bats and birds increase crop yield in tropical agroforestry landscapes. *Ecol. Lett.* **16**, 1480–1487 (2013).
102. M. E. O'Rourke, K. Rienzo-Stack, A. G. Power, A multi-scale, landscape approach to predicting insect populations in agroecosystems. *Ecol. Appl.* **21**, 1782–1791 (2011).
103. M. Jonsson, H. L. Buckley, B. S. Case, S. D. Wratten, R. J. Hale, R. K. Didham, Agricultural intensification drives landscape-context effects on host-parasitoid interactions in agroecosystems. *J. Appl. Ecol.* **49**, 706–714 (2012).
104. M. Kishinevsky, T. Keasar, A. R. Harari, E. Chiel, A comparison of naturally growing vegetation vs. border-planted companion plants for sustaining parasitoids in pomegranate orchards. *Agric. Ecosyst. Environ.* **246**, 117–123 (2017).
105. E. A. Martin, B. Seo, C.-R. Park, B. Reineking, I. Steffan-Dewenter, Scale-dependent effects of landscape composition and configuration on natural enemy diversity, crop herbivory, and yields. *Ecol. Appl.* **26**, 448–462 (2016).
106. K. Poveda, E. Martínez, M. F. Kersch-Becker, M. A. Bonilla, T. Tschamtkke, Landscape simplification and altitude affect biodiversity, herbivory and Andean potato yield. *J. Appl. Ecol.* **49**, 513–522 (2012).
107. M. B. Takada, A. Yoshioka, S. Takagi, S. Iwabuchi, I. Washitani, Multiple spatial scale factors affecting mirid bug abundance and damage level in organic rice paddies. *Biol. Control.* **60**, 169–174 (2012).
108. M. G. E. Mitchell, E. M. Bennett, A. Gonzalez, Agricultural landscape structure affects arthropod diversity and arthropod-derived ecosystem services. *Agric. Ecosyst. Environ.* **192**, 144–151 (2014).
109. F. Geiger, J. Bengtsson, F. Berendse, W. W. Weisser, M. Emmerson, M. B. Morales, P. Ceryngier, J. Liira, T. Tschamtkke, C. Winqvist, S. Eggers, R. Bommarco, T. Pärt, V. Bretagnolle, M. Plantegenest, L. W. Clement, C. Dennis, C. Palmer, J. J. Oñate, I. Guerrero, V. Hawro, T. Aavik, C. Thies, A. Flohre, S. Hänke, C. Fischer, P. W. Goedhart, P. Inchausti, Persistent negative effects of pesticides on biodiversity and biological control potential on European farmland. *Basic Appl. Ecol.* **11**, 97–105 (2010).
110. M. Plečaš, V. Gagić, M. Janković, O. Petrović-Obradović, N. G. Kavallieratos, Ž. Tomanović, C. Thies, T. Tschamtkke, A. Četković, Landscape composition and configuration influence cereal aphid-parasitoid-hyperparasitoid interactions and biological control differentially across years. *Agric. Ecosyst. Environ.* **183**, 1–10 (2014).
111. M. Tschumi, M. Albrecht, M. H. Entling, K. Jacot, High effectiveness of tailored flower strips in reducing pests and crop plant damage. *Proc. R. Soc. Biol. Sci.* **282**, 20151369 (2015).

Acknowledgments: We thank R. Carloni for producing icons in Figs. 1 to 4. We also thank the Department of Innovation, Research and University of the Autonomous Province of Bozen/Bolzano for covering the Open Access publication costs. For all further acknowledgements, see the Supplementary Materials. **Funding:** This work was funded by EU-FP7 LIBERATION (311781) and Biodiversa-FACCE ECODEAL (PCIN-2014-048). **Author contributions:** M.D., E.A.M., and I.S.-D. conceived the study. M.D. performed statistical analyses and wrote the manuscript draft. The authors named from E.A.M. to T.T. and I.S.-D. discussed and revised earlier versions of the manuscript. The authors named from G.K.S.A. to Y.Z. are listed alphabetically, as they contributed equally in gathering field data and providing several important corrections to subsequent manuscript drafts. **Competing interests:** The authors declare that they have no competing interests. **Data and materials availability:** All data needed to evaluate the conclusions in the paper are present in the paper and/or the Supplementary Materials. Additional data related to this paper may be requested from the authors.

Submitted 14 February 2019

Accepted 22 September 2019

Published 16 October 2019

10.1126/sciadv.aax0121

Citation: M. Dainese, E. A. Martin, M. A. Aizen, M. Albrecht, I. Bartomeus, R. Bommarco, L. G. Carvalheiro, R. Chaplin-Kramer, V. Gagic, L. A. Garibaldi, J. Ghazoul, H. Grab, M. Jonsson, D. S. Karp, C. M. Kennedy, D. Kleijn, C. Kremen, D. A. Landis, D. K. Letourneau, L. Marini, K. Poveda, R. Rader, H. G. Smith, T. Tschamtkke, G. K. S. Andersson, I. Badenhausser, S. Baensch, A. D. M. Bezerra, F. J. A. Bianchi, V. Boreux, V. Bretagnolle, B. Caballero-Lopez, P. Cavigliasso, A. Četković, N. P. Chacoff, A. Classen, S. Cusser, F. D. da Silva e Silva, G. A. deGroot, J. H. Dudenhöffer, J. Ekroos, T. Fijen, P. Franck, B. M. Freitas, M. P. D. Garratt, C. Gratton, J. Hipólito, A. Holzschuh, L. Hunt, A. L. Iverson, S. Jha, T. Keasar, T. N. Kim, M. Kishinevsky, B. K. Klatt, A.-M. Klein, K. M. Krewenka, S. Krishnan, A. E. Larsen, C. Lavigne, H. Liere, B. Maas, R. E. Mallinger, E. Martinez Pachon, A. Martinez-Salinas, T. D. Meehan, M. G. E. Mitchell, G. A. R. Molina, M. Nesper, L. Nilsson, M. E. O'Rourke, M. K. Peters, M. Plečaš, S. G. Potts, D. d. L. Ramos, J. A. Rosenheim, M. Rundlöf, A. Rusch, A. Sáez, J. Scheper, M. Schleuning, J. M. Schmack, A. R. Sciligo, C. Seymour, D. A. Stanley, R. Stewart, J. C. Stout, L. Sutter, M. B. Takada, H. Taki, G. Tamburini, M. Tschumi, B. F. Viana, C. Westphal, B. K. Willcox, S. D. Wratten, A. Yoshioka, C. Zaragoza-Trello, W. Zhang, Y. Zou, I. Steffan-Dewenter, A global synthesis reveals biodiversity-mediated benefits for crop production. *Sci. Adv.* **5**, eaax0121 (2019).

A global synthesis reveals biodiversity-mediated benefits for crop production

Matteo Dainese, Emily A. Martin, Marcelo A. Aizen, Matthias Albrecht, Ignasi Bartomeus, Riccardo Bommarco, Luisa G. Carvalheiro, Rebecca Chaplin-Kramer, Vesna Gagic, Lucas A. Garibaldi, Jaboury Ghazoul, Heather Grab, Mattias Jonsson, Daniel S. Karp, Christina M. Kennedy, David Kleijn, Claire Kremen, Douglas A. Landis, Deborah K. Letourneau, Lorenzo Marini, Katja Poveda, Romina Rader, Henrik G. Smith, Teja Tscharntke, Georg K. S. Andersson, Isabelle Badenhausser, Svenja Baensch, Antonio Diego M. Bezerra, Felix J. J. A. Bianchi, Virginie Boreux, Vincent Bretagnolle, Berta Caballero-Lopez, Pablo Cavigliasso, Aleksandar Cetkovic, Natacha P. Chacoff, Alice Classen, Sarah Cusser, Felipe D. da Silva e Silva, G. Arjen de Groot, Jan H. Dudenhöffer, Johan Ekroos, Thijs Fijen, Pierre Franck, Breno M. Freitas, Michael P. D. Garratt, Claudio Gratton, Juliana Hipólito, Andrea Holzschuh, Lauren Hunt, Aaron L. Iverson, Shalene Jha, Tamar Keasar, Tania N. Kim, Miriam Kishinevsky, Björn K. Klatt, Alexandra-Maria Klein, Kristin M. Krewenka, Smitha Krishnan, Ashley E. Larsen, Claire Lavigne, Heidi Liere, Bea Maas, Rachel E. Mallinger, Eliana Martinez Pachon, Alejandra Martínez-Salinas, Timothy D. Meehan, Matthew G. E. Mitchell, Gonzalo A. R. Molina, Maike Nesper, Lovisa Nilsson, Megan E. O'Rourke, Marcell K. Peters, Milan Plecas, Simon G. Potts, Davi de L. Ramos, Jay A. Rosenheim, Maj Rundlöf, Adrien Rusch, Agustín Sáez, Jeroen Scheper, Matthias Schleuning, Julia M. Schmack, Amber R. Sciligo, Colleen Seymour, Dara A. Stanley, Rebecca Stewart, Jane C. Stout, Louis Sutter, Mayura B. Takada, Hisatomo Taki, Giovanni Tamburini, Matthias Tschumi, Blandina F. Viana, Catrin Westphal, Bryony K. Willcox, Stephen D. Wratten, Akira Yoshioka, Carlos Zaragoza-Trello, Wei Zhang, Yi Zou and Ingolf Steffan-Dewenter

Sci Adv 5 (10), eaax0121.
DOI: 10.1126/sciadv.aax0121

ARTICLE TOOLS

<http://advances.sciencemag.org/content/5/10/eaax0121>

SUPPLEMENTARY MATERIALS

<http://advances.sciencemag.org/content/suppl/2019/10/10/5.10.eaax0121.DC1>

REFERENCES

This article cites 101 articles, 13 of which you can access for free
<http://advances.sciencemag.org/content/5/10/eaax0121#BIBL>

PERMISSIONS

<http://www.sciencemag.org/help/reprints-and-permissions>

Use of this article is subject to the [Terms of Service](#)

Science Advances (ISSN 2375-2548) is published by the American Association for the Advancement of Science, 1200 New York Avenue NW, Washington, DC 20005. The title *Science Advances* is a registered trademark of AAAS.

Copyright © 2019 The Authors, some rights reserved; exclusive licensee American Association for the Advancement of Science. No claim to original U.S. Government Works. Distributed under a Creative Commons Attribution NonCommercial License 4.0 (CC BY-NC).

Supplementary Materials for

A global synthesis reveals biodiversity-mediated benefits for crop production

Matteo Dainese*, Emily A. Martin, Marcelo A. Aizen, Matthias Albrecht, Ignasi Bartomeus, Riccardo Bommarco, Luisa G. Carvalheiro, Rebecca Chaplin-Kramer, Vesna Gagic, Lucas A. Garibaldi, Jaboury Ghazoul, Heather Grab, Mattias Jonsson, Daniel S. Karp, Christina M. Kennedy, David Kleijn, Claire Kremen, Douglas A. Landis, Deborah K. Letourneau, Lorenzo Marini, Katja Poveda, Romina Rader, Henrik G. Smith, Teja Tscharntke, Georg K. S. Andersson, Isabelle Badenhauer, Svenja Baensch, Antonio Diego M. Bezerra, Felix J. J. A. Bianchi, Virginie Boreux, Vincent Bretagnolle, Berta Caballero-Lopez, Pablo Cavigliasso, Aleksandar Četković, Natacha P. Chacoff, Alice Classen, Sarah Cusser, Felipe D. da Silva e Silva, G. Arjen de Groot, Jan H. Dudenhöffer, Johan Ekroos, Thijs Fijen, Pierre Franck, Breno M. Freitas, Michael P. D. Garratt, Claudio Gratton, Juliana Hipólito, Andrea Holzschuh, Lauren Hunt, Aaron L. Iverson, Shalene Jha, Tamar Keasar, Tania N. Kim, Miriam Kishinevsky, Björn K. Klatt, Alexandra-Maria Klein, Kristin M. Krewenka, Smitha Krishnan, Ashley E. Larsen, Claire Lavigne, Heidi Liere, Bea Maas, Rachel E. Mallinger, Eliana Martinez Pachon, Alejandra Martínez-Salinas, Timothy D. Meehan, Matthew G. E. Mitchell, Gonzalo A. R. Molina, Maike Nesper, Lovisa Nilsson, Megan E. O'Rourke, Marcell K. Peters, Milan Plečaš, Simon G. Potts, Davi de L. Ramos, Jay A. Rosenheim, Maj Rundlöf, Adrien Rusch, Agustín Sáez, Jeroen Scheper, Matthias Schleuning, Julia M. Schmack, Amber R. Sciligo, Colleen Seymour, Dara A. Stanley, Rebecca Stewart, Jane C. Stout, Louis Sutter, Mayura B. Takada, Hisatomo Taki, Giovanni Tamburini, Matthias Tschumi, Blandina F. Viana, Catrin Westphal, Bryony K. Willcox, Stephen D. Wratten, Akira Yoshioka, Carlos Zaragoza-Trello, Wei Zhang, Yi Zou, Ingolf Steffan-Dewenter

*Corresponding author. Email: matteo.dainese@eurac.edu

Published 16 October 2019, *Sci. Adv.* **5**, eaax0121 (2018)
DOI: 10.1126/sciadv.aax0121

The PDF file includes:

Supplementary Text

Fig. S1. Forest plot of the effect of pollinator richness on pollination for individual studies.

Fig. S2. Forest plot of the effect of natural enemy richness on pest control for individual studies.

Fig. S3. Direct and indirect landscape simplification effects on ecosystem services via changes in richness and abundance or richness and evenness.

Fig. S4. Direct and cascading landscape simplification effects on final crop production via changes in natural enemy richness, abundance evenness, and pest control (all sites together, with and without insecticide application).

Fig. S5. Forest plot of the effect of landscape simplification on natural enemy abundance for individual studies.

Fig. S6. Mediation model.

Fig. S7. Direct and indirect effects of pollinator richness, abundance, and evenness (with honey bees) on pollination.

Fig. S8. Direct and cascading landscape simplification effects on area-based yield via changes in richness and ecosystem services.

Table S1. List of 89 studies considered in our analyses.

Table S2. Model output for richness–ecosystem service relationships.

Table S3. Model output for path models testing direct and indirect effects (mediated by changes in abundance) of richness on ecosystem services.

Table S4. Model output for path models testing direct and indirect effects (mediated by changes in evenness) of richness on ecosystem services.

Table S5. Model output for path models testing direct and indirect effects (mediated by changes in richness) of landscape simplification on ecosystem services.

Table S6. Model output for path models testing the direct and cascading landscape simplification effects on ecosystem services via changes in richness and abundance.

Table S7. Model output for path models testing the direct and cascading landscape simplification effects on ecosystem services via changes in richness and evenness.

Table S8. Model output for path models testing the direct and cascading landscape simplification effects on final crop production via changes in richness, evenness, and ecosystem services.

Table S9. Model output for path models testing the direct and cascading landscape simplification effects on final crop production via changes in richness, abundance, and ecosystem services.

Table S10. Model output for path models testing direct and indirect effects of pollinator richness, abundance, and evenness (with honey bees) on pollination.

Table S11. Results of pairwise comparison of richness–ecosystem service relationships according to the methods used to sample pollinators and natural enemies.

Table S12. Results of pairwise comparison of richness–ecosystem service relationships according to the methods used to quantify pollination and pest control services.

References (66–111)

Other Supplementary Material for this manuscript includes the following:

(available at advances.sciencemag.org/cgi/content/full/5/10/eaax0121/DC1)

Database S1 (Microsoft Excel format). Data on pollinator and natural enemy diversity and associated ecosystem services compiled from 89 studies and 1475 locations around the world.

Supplementary Text

Detailed Acknowledgements

M.D., E.A.M and I.S.D. were supported by EU-FP7 LIBERATION (311781).

M.D., E.A.M, A.H., J.S and I.S.D were supported by Biodiversa-FACCE ECODEAL (PCIN-2014–048).

C.M.K. was supported by an USDA-NIFA Organic Agriculture Research & Extension Award (2015-51300-24155).

D.S.K. was supported by the "National Socio-Environmental Synthesis Center (SESYNC)—National Science Foundation Award DBI-1052875".

D.A.S. was supported by the Environmental Protection Agency, Ireland.

D.K.L. was supported by the United States Dept. Agriculture-NRI grant 2005-55302-16345.

D.A.L. was supported by the US DOE Office of Science (DE-FCO2-07ER64494) and Office of Energy Efficiency and Renewable Energy (DE-ACO5-76RL01830) to the DOE Great Lakes Bioenergy Research Center, and by the NSF Long-term Ecological Research Program (DEB 1027253) at the Kellogg Biological Station and by Michigan State University AgBioResearch.

H.G.S. was supported by The Swedish Research Council Formas & the Strategic Research Area BECC.

J.G. was supported by the Swiss National Science Foundation, the Mercator Foundation, and ETH Zurich grants. L.M. was supported by EU-FP7 LIBERATION (311781).

H.G. was supported by an USDA Northeast Sustainable Agriculture Research and Extension Award #GNE12-037.

M.J. was supported by the Centre for Biological Control, SLU.

T.T. was supported by the DFG FOR 2432 (India), the project "Diversity Turn" (VW Foundation), the DFG SFB 990 EFForTS and the GIZ-Bioversity project on Peruvian cocoa.

I.B. was supported by Biodiversa-FACCE ECODEAL (PCIN-2014–048).

S.B. acknowledges her Ph.D scholarship by the Deutsche Bundesstiftung Umwelt (German Federal Environmental Foundation).

A.D.M.B. thanks for a Ph.D scholarship financed by The Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Finance Code 001. P.C. was supported by Programa Nacional Apícola (PROAPI).

A.C. research conducted within the Research Unit FOR1246 (KILI project) funded by the German Research Foundation (DFG).

VB was supported by the Project “Pollinisateurs” from MTES and Biodiversa-FACCE ECODEAL (PCIN-2014–048).

B.C.L. was supported by a PhD scholarship of the Catalan Council for Universities, Research and Society (2005-FI AGAUR, and 2006 BE-2 00229) and a grant from the Margit and Folke Pehrzon Foundation (2007). Research was funded from two grants from the Swedish Research

Council for Environment, Agricultural Sciences and Spatial planning (FORMAS) (to HGS and RB).

F.D.S.S. was supported by the “Fundação de Apoio à Pesquisa do Distrito Federal”/FAPDF, Brazil (Foundation for Research Support of the Distrito Federal), nº 9852.56.31658.07042016. This study was also financed in part by the Coordenação de Aperfeiçoamento de Pessoal de Nível Superior - Brasil (CAPES) - Finance Code 001.

G.A.G. was funded by the Dutch Ministry of Agriculture, Nature and Food Quality (BO-11-0.11.01-0.51).

J.E. was supported by The Swedish Research Council Formas & the Strategic Research Area BECC.

T.F. was jointly funded by the Netherlands Organization for Scientific Research (research programme NWO-Green) and BASF - Vegetable Seeds under project number 870.15.030. P.F. and C. L. were supported by ANR grant Peerless “Predictive Ecological Engineering for Landscape Ecosystem Services and Sustainability” (ANR-12-AGRO-0006).

B.M.F thanks the Project "Conservation and Management of Pollinators for Sustainable Agriculture, through an Ecosystem Approach", which is supported by the Global Environmental Facility Bank (GEF), coordinated by the Food and Agriculture Organization of the United Nations (FAO) with implementation support from the United Nations Environment Programme (UNEP) and supported in Brazil by the Ministry of Environment (MMA) and Brazilian Biodiversity Fund (Funbio). Also to the National Council for Scientific and Technological Development - CNPq, Brasília-Brazil for financial support to the Brazilian Network of Cashew Pollinators (project # 556042/2009-3) and a Productivity Research Grant (#302934/2010-3).

M.P.D.G. was funded jointly by a grant from BBSRC, Defra, NERC, the Scottish Government and the Wellcome Trust, under the UK Insect Pollinators Initiative.

C.G. was funded in part by the DOE Great Lakes Bioenergy Research Center (DOE Office of Science BER DE-FC02-07ER64494) and by a USDA Agriculture and Food Research Initiative Competitive Grant 2011-67009-30022.

J.H. was supported by CNPq – INCT-IN-TREE (465767/2014-1), CNPq-PVE (407152/2013-0) and Capes for Ph.D scholarship.

A.L.I. was supported by the University of Michigan - Department of Ecology and Evolutionary Biology and the Rackham Graduate School, University of Michigan.

B.K.K. was supported by the German Research Foundation (DFG).

A.M.K. data are part of the EcoFruit project funded through the 2013-2014 BiodivERsA/FACCE-JPI joint call (agreement# BiodivERsA-FACCE2014-74), with the national funder of the German Federal Ministry of Education and Research (PT-DLR/BMBF) (grant# 01LC1403).

T.K. was supported by the Israeli Ministry of Agriculture and Rural Development (grant number 131-1793-14).

T.N.K. was funded in part by the DOE Great Lakes Bioenergy Research Center (DOE Office of Science BER DE-FC02-07ER64494) and by a USDA Agriculture and Food Research Initiative Competitive Grant 2011-67009-30022.

B.M. was partly funded by a scholarship of the German National Academic Foundation (Studienstiftung des deutschen Volkes) and benefited from financial support from the German Science Foundation (DFG Grant CL-474/1-1, ELUC).

R.E.M. was funded by a Specialty Crop Block Grant from the Wisconsin Department of Agriculture, Trade, and Consumer Protection.

M.G.E.M. was supported by an NSERC PGS-D Scholarship and an NSERC Strategic Projects Grant.

S.G.P. was funded jointly by a grant from BBSRC, Defra, NERC, the Scottish Government and the Wellcome Trust, under the UK Insect Pollinators Initiative.

J.A.R. was supported by an USDA grant CA-D-ENM-5671-RR.

A.M.S. was supported by The University of Idaho, the USFWS (NMBCA grant F11AP01025), the Perennial Crop Platform (PCP) and CGIAR - Water Land and Ecosystems (WLE).

M.S. was supported by DFG grant FOR 1246 “Kilimanjaro ecosystems under global change: Linking biodiversity, biotic interactions and biogeochemical ecosystem processes”.

A.R.S. research was funded by the CS Fund and the United States Army Research Office (ARO).

M.T. was supported by the Hauser and Sur-La-Croix foundations.

H.T. was supported by the Environment Research and Technology Development Fund (S-15-2) of the Ministry of the Environment, and JSPS KAKENHI Grant Number 18H02220.

B.F.V. was supported by CNPq – INCT-IN-TREE (#465767/2014-1), CNPq-PVE (407152/2013-0) and a Productivity Research Grant CNPq (#305470/2013-2).

C.W. was supported by the Deutsche Forschungsgemeinschaft (DFG) (Projekt number 405945293).

B.K.W. was supported by a PhD scholarship from the University of New England and funded by RnD4Profit-14-01-008 “Multi-scale monitoring tools for managing Australian Tree Crops: Industry meets innovation”.

C.Z.T. was supported by a Severo-Ochoa predoctoral fellowship (SVP-2014-068580) and by the Biodiversa-FACCE project ECODEAL.

W.Z. was supported by CGIAR research program on Water, land and ecosystems (WLE).

Y.Z. was funded by the Division for Earth and Life Sciences of the Netherlands Organization for Scientific Research (grant 833.13.004).

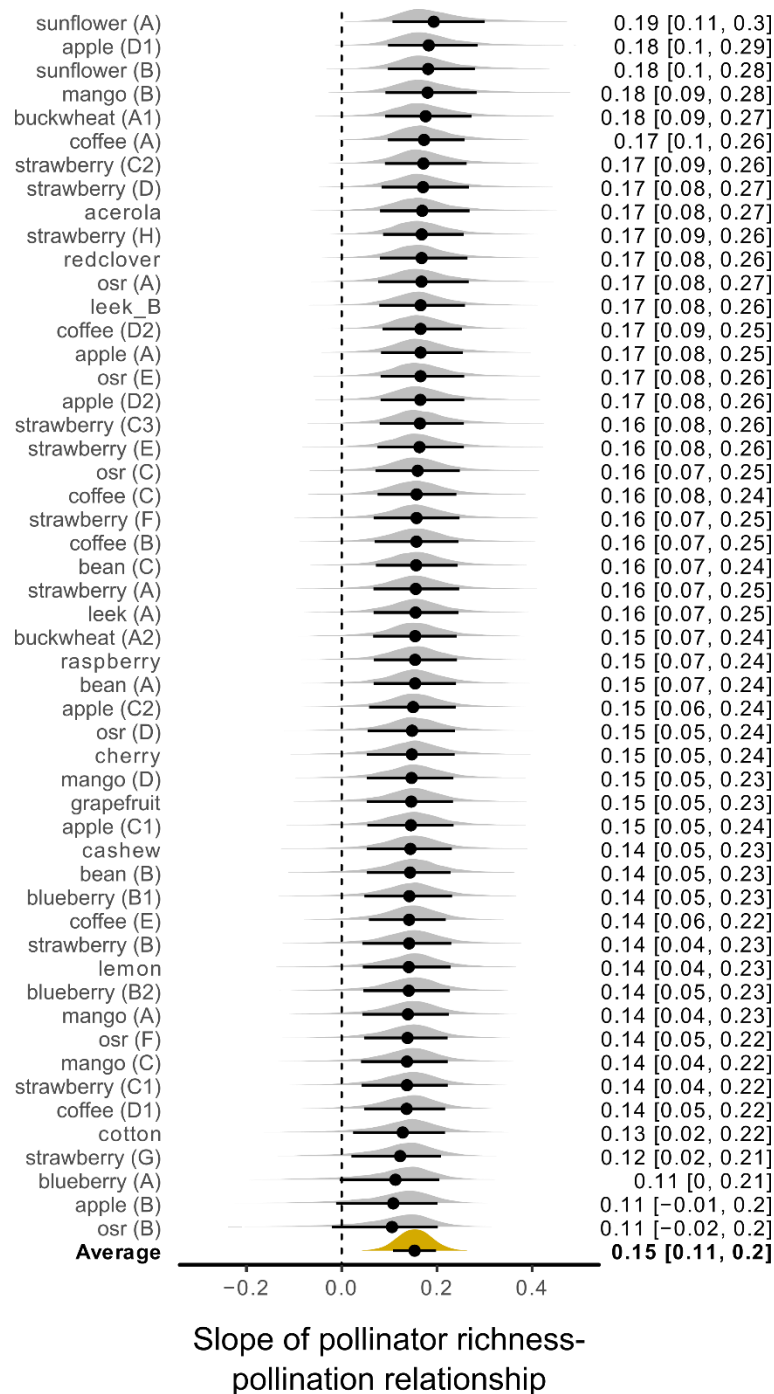


Fig. S1. Forest plot of the effect of pollinator richness on pollination for individual studies. Each posterior distribution represents medians (symbol centres) and 90% density intervals (black lines).

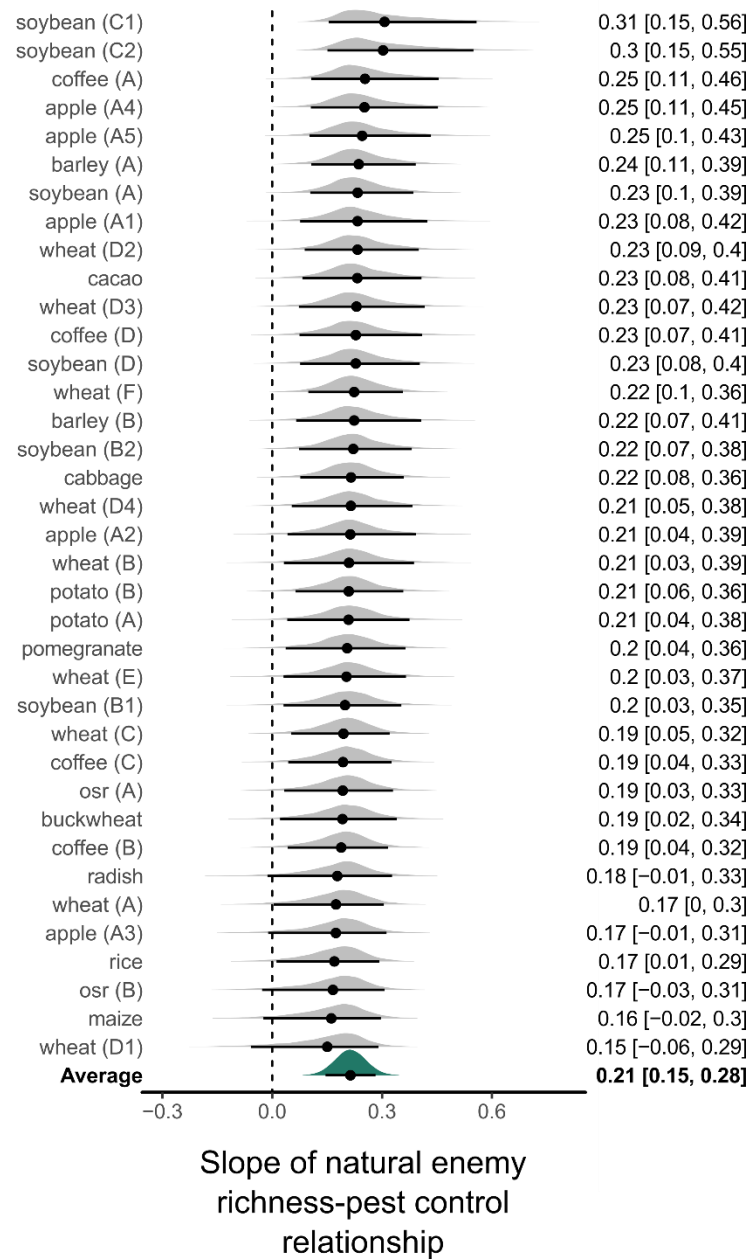


Fig. S2. Forest plot of the effect of natural enemy richness on pest control for individual studies. Each posterior distribution represents medians (symbol centres) and 90% density intervals (black lines).

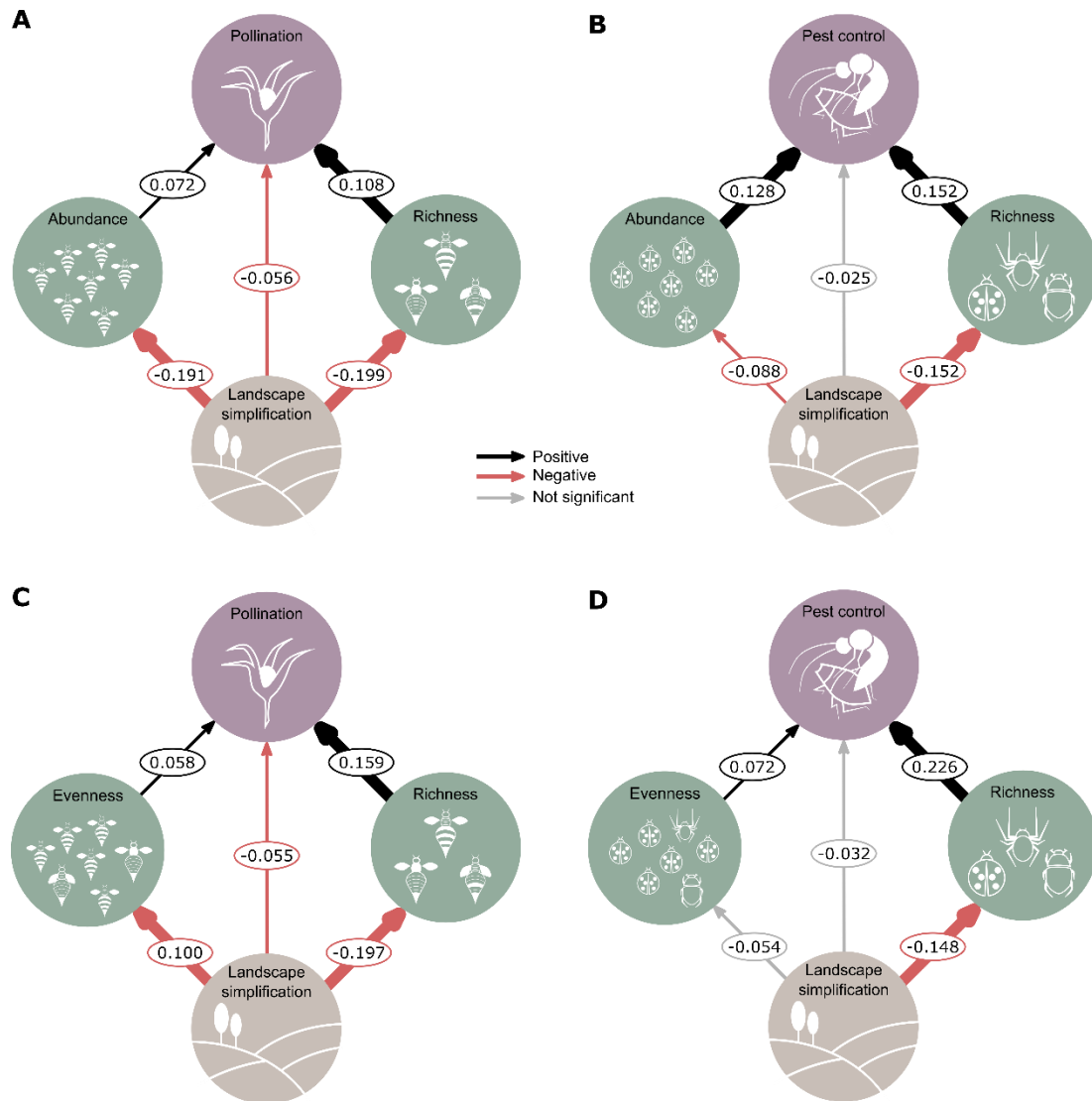


Fig. S3. Direct and indirect landscape simplification effects on ecosystem services via changes in richness and abundance or richness and evenness. (A) Path model representing direct and indirect effects of landscape simplification on pollination through changes in pollinator richness and abundance. (B) Path model representing direct and indirect effects of landscape simplification on pest control services through changes in natural enemy richness and abundance. (C) Path model representing direct and indirect effects of landscape simplification on pollination through changes in pollinator richness and evenness. (D) Path model representing direct and indirect effects of landscape simplification on pest control services through changes in natural enemy richness and evenness. Pollination models: $n = 821$ fields of 52 studies. Pest control models: $n = 654$ fields of 37 studies. Path coefficients are effect sizes estimated from the median of the posterior distribution of the model. Black and red arrows represent positive or negative effects, respectively. Arrow widths are proportional to highest density intervals (HDIs). Grey arrows represent non-evident effects (HDIs overlapped zero).

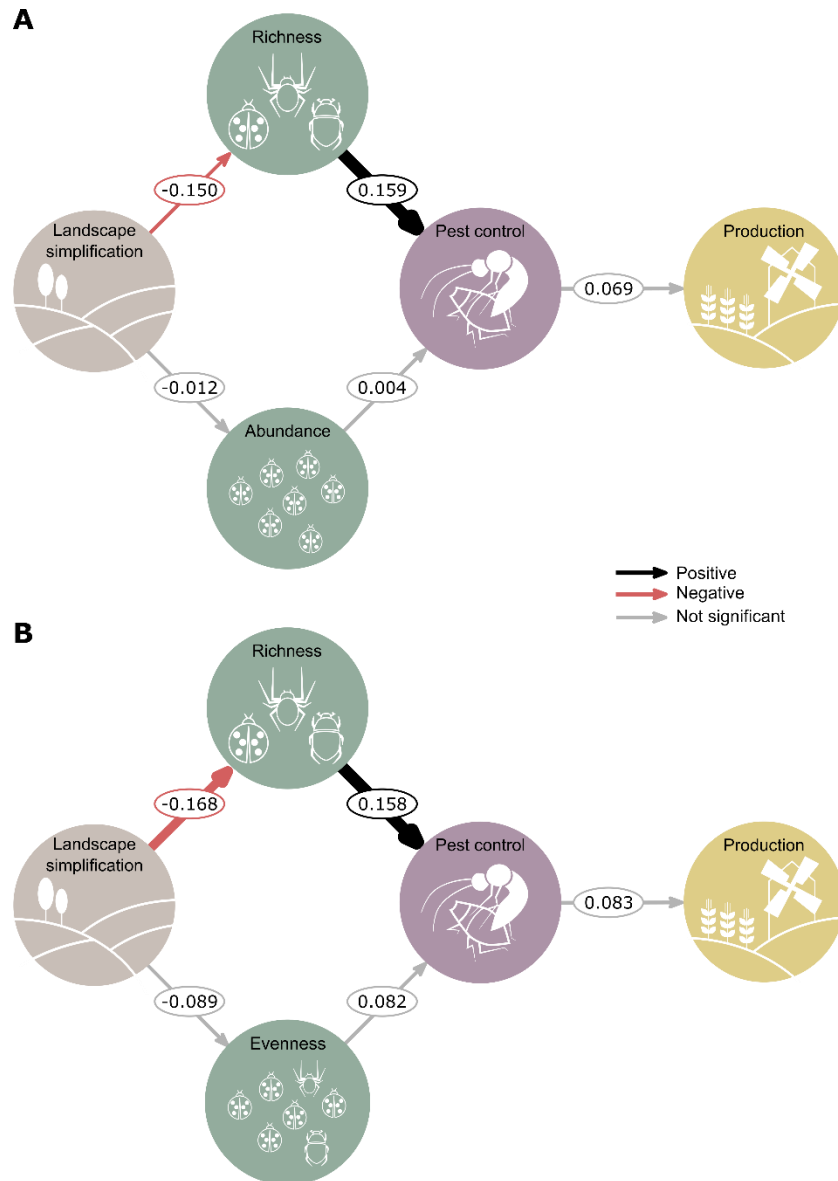


Fig. S4. Direct and cascading landscape simplification effects on final crop production via changes in natural enemy richness, abundance evenness, and pest control (all sites together, with and without insecticide application). (A) Path model representing direct and indirect effects of landscape simplification on final crop production through changes in natural enemy richness, abundance and pest control. (B) Path model representing direct and indirect effects of landscape simplification on final crop production through changes in natural enemy richness, evenness and pest control. Path coefficients are effect sizes estimated from the median of the posterior distribution of the model ($n = 236$ fields of 15 studies). Black and red arrows represent positive or negative effects, respectively. Arrow widths are proportional to highest density intervals (HDIs). Grey arrows represent non-significant effects (HDIs overlapped zero).

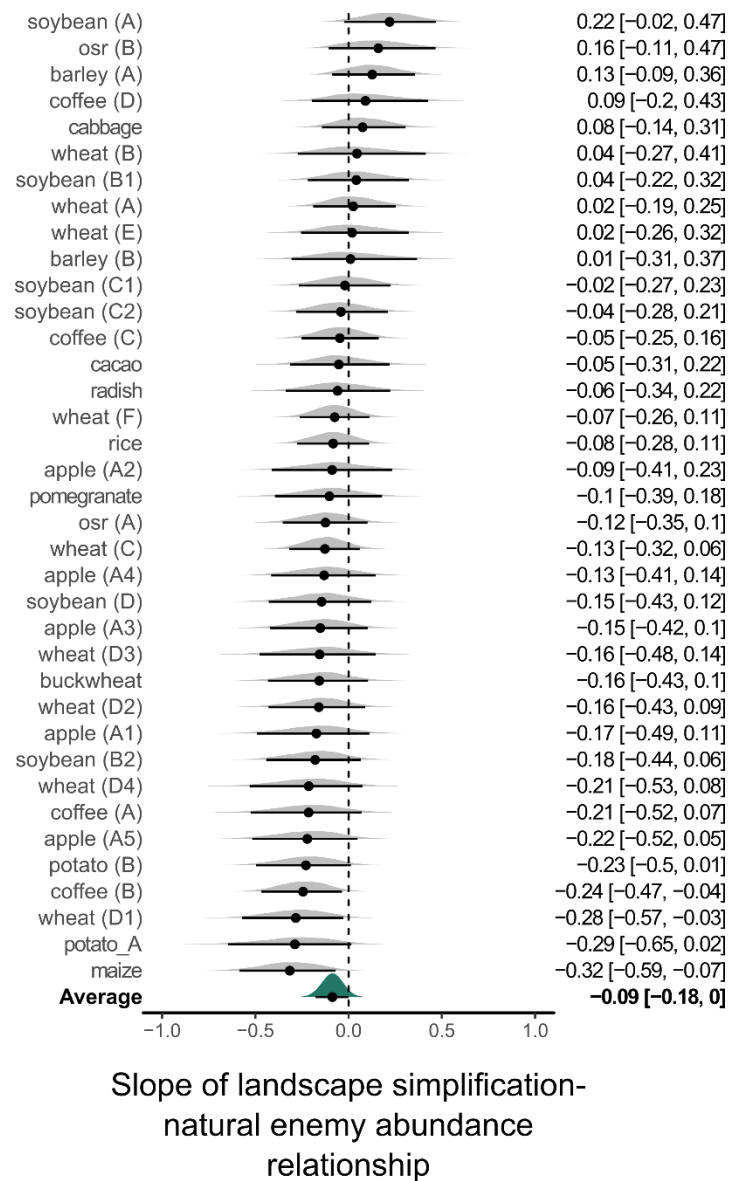


Fig. S5. Forest plot of the effect of landscape simplification on natural enemy abundance for individual studies. Each posterior distribution represents medians (symbol centres) and 90% density intervals (black lines).

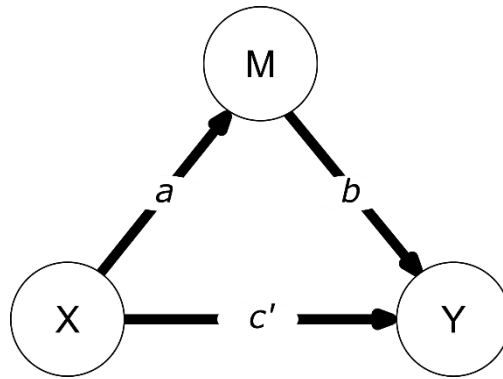


Fig. S6. Mediation model. Mediation analysis is a statistical procedure to test whether the effect of an independent variable X on a dependent variable Y ($X \rightarrow Y$) is at least partly explained via the inclusion of a third hypothetical variable, the mediator variable M ($X \rightarrow M \rightarrow Y$). The three causal paths a , b , and c' represent X 's effect on M , M 's effect on Y , and X 's effect on Y while accounting for M , respectively. The three causal paths correspond to parameters from two regression models, one in which M is the outcome and X the predictor, and one in which Y is the outcome and X and M the simultaneous predictors.

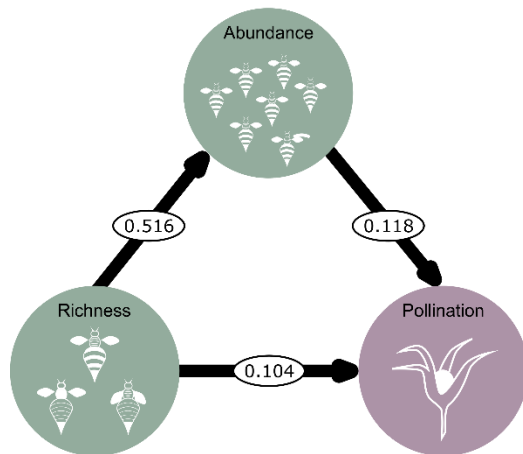
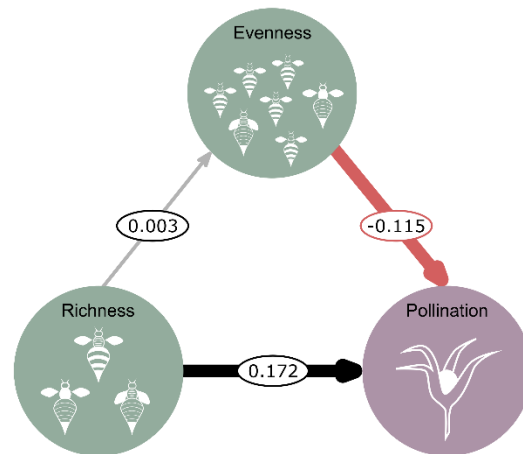
A**B**

Fig. S7. Direct and indirect effects of pollinator richness, abundance, and evenness (with honey bees) on pollination. (A) Path model of pollinator richness as a predictor of pollination, mediated by pollinator abundance. (B) Path model of pollinator richness as a predictor of pollination, mediated by pollinator evenness. $n = 821$ fields of 52 studies. Coefficients of the three causal paths (a , b , c) correspond to the median of the posterior distribution of the model. The proportion mediated is the mediated effect ($a \times b$) divided by the total effect (c).

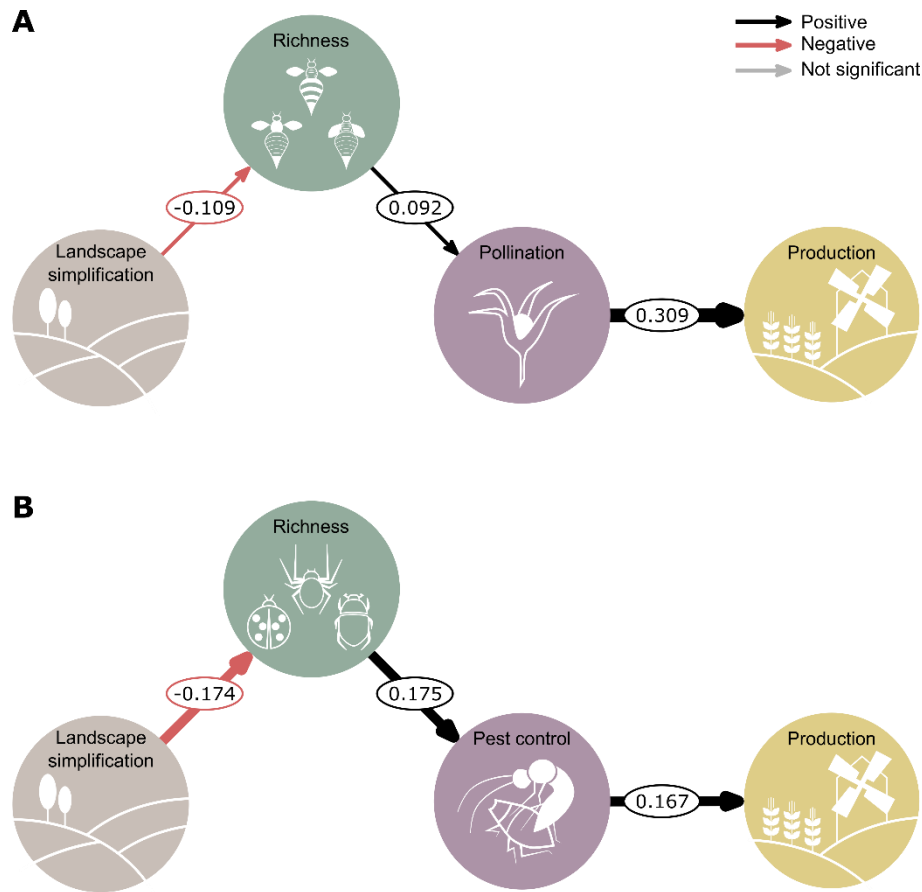


Fig. S8. Direct and cascading landscape simplification effects on area-based yield via changes in richness and ecosystem services. (A) Path model representing direct and indirect effects of landscape simplification on final area-based yield through changes in pollinator richness and pollination ($n = 203$ fields of 13 studies). (B) Path model representing direct and indirect effects of landscape simplification on final area-based yield through changes in natural enemy richness and pest control ($n = 93$ fields of 7 studies). Path coefficients are effect sizes estimated from the median of the posterior distribution of the model. Black and red arrows represent positive or negative effects, respectively. Arrow widths are proportional to highest density intervals (HDIs). Grey arrows represent non-significant effects (HDIs overlapped zero).

Table S1. List of 89 studies considered in our analyses.

Study code	Reference and (or) data holder contact	Crop species	Country, region	Study year	Sites (with yield)	Sampling methods	Taxa	Functions	Production
Pollination studies									
acerola	(43) Freitas, freitas@ufc.br	<i>Malpighia emarginata</i>	Brazil, Ceará	2011	8	active	bees	fruit set	-
apple (A)	(65) Boreux, virginie.boreux@nature.uni-freiburg.de	<i>Malus domestica</i>	Germany, Lake Constance	2015	25	active	bees	fruit set	-
apple (B)	(66) Garratt, m.p.garratt@reading.ac.uk	<i>Malus domestica</i>	UK, Kent	2011	8	active, passive	bees	fruit set	-
apple (C1)	(67) de Groot, g.a.degroot@wur.nl	<i>Malus domestica</i>	Netherlands, Betuwe	2013	8 (4)	active	bees, hoverflies	fruit set	crop yield
apple (C2)	(67) de Groot, g.a.degroot@wur.nl	<i>Malus domestica</i>	Netherlands, Betuwe	2014	10 (9)	active	bees, hoverflies	fruit set	crop yield
apple (D1)	(68) Mallinger, rachel.mallinger@ars.usda.gov	<i>Malus domestica</i>	USA, Wisconsin	2012	17	passive	bees	fruit set	-
apple (D2)	(68) Mallinger, rachel.mallinger@ars.usda.gov	<i>Malus domestica</i>	USA, Wisconsin	2013	19	passive	bees	fruit set	-
bean (A)	Ekroos, johan.ekroos@cec.lu.se	<i>Vicia faba</i>	Sweden, Scania	2016	16 (16)	active	bees	seed set	plant yield
bean (B)	(69) Garratt, m.p.garratt@reading.ac.uk	<i>Vicia faba</i>	UK, Berkshire	2011	8	active, passive	bees	seed set	-
bean (C)	(70) Ramos, Silva davilramos91@gmail.com felipe.silva@bag.ifmt.edu.br	<i>Phaseolus vulgaris</i>	Brazil, Goiás/DF	2015/2016	22 (22)	active	bees	seed set	crop yield
blueberry (A)	Cavigliasso, pablo.cavigliasso@gmail.com	<i>Vaccinium corymbosum</i>	Argentina, Espinal-Nandubay	2016	13	active	bees, wasps, hoverflies	fruit set	-
blueberry (B1)	(67) de Groot, g.a.degroot@wur.nl	<i>Vaccinium corymbosum</i>	Netherlands, Limburg/Overijssel	2013	10 (9)	active	bees	fruit set	crop yield
blueberry (B2)	(67) de Groot, g.a.degroot@wur.nl	<i>Vaccinium corymbosum</i>	Netherlands, Limburg/Overijssel	2014	15 (13)	active	bees	fruit set	crop yield
buckwheat (A1)	(71, 72) Taki, htaki@affrc.go.jp	<i>Fagopyrum esculentum</i>	Japan, Ibaraki	2007	15	active	bees, butterflies, flies, wasps	seed set	-
buckwheat (A2)	(71, 72) Taki, htaki@affrc.go.jp	<i>Fagopyrum esculentum</i>	Japan, Ibaraki	2008	17	active	bees, butterflies, flies, wasps	seed set	-
cashew	(21) Freitas, freitas@ufc.br	<i>Anacardium occidentale</i>	Brazil, Ceará	2012	10 (10)	active	bees	fruit set	crop yield
cherry	(73) Holzschuh, andrea.holzschuh@uni-wuerzburg.de	<i>Prunus avium</i>	Germany, Hesse	2008	7	active	bees	fruit set	-
coffee (A)	(46, 74, 75) Boreux,	<i>Coffea canephora</i>	India, Kodagu	2008	53 (51)	active	bees	fruit set	plant yield

	virginie.boreux@nature.uni-freiburg.de								
coffee (B)	(76) Classen, alice.classen@uni-wuerzburg.de	<i>Coffea arabica</i>	Tanzania, Kilimanjaro	2011/2012	11 (6)	active, passive	bees	fruit set	plant yield
coffee (C)	(77) Hipólito, jhdsousa@yahoo.com	<i>Coffea arabica</i>	Brazil, Chapada Diamantina	2013	30 (28)	active	bee, flies, butterflies , beetles, wasps	fruit set	crop yield
coffee (D1)	(74, 78, 79) Krishnan, smithakrishnan@gmail.com	<i>Coffea canephora</i>	India, Kodagu	2007	35	active	bees	fruit set	-
coffee (D2)	(74, 78, 79) Krishnan, smithakrishnan@gmail.com	<i>Coffea canephora</i>	India, Kodagu	2008	37	active	bees	fruit set	
coffee (E)	(80) Krishnan, Nesper, smithakrishnan@gmail.com maike.nesper@gmail.com	<i>Coffea canephora</i>	India, Kodagu	2014	49 (49)	active	bees	fruit set	crop yield
cotton	(81) Cusser, sarah.cusser@gmail.com	<i>Gossypium hirsutum</i>	USA, Gulf Coast Texas	2014	11	active	bee, hoverflies, butterflies , beetles	fruit set	-
grapefruit	(82, 83) Chacoff, nchacoff@gmail.com	<i>Citrus paradisi</i>	Argentina, Yungas	2000	6	active	bee, flies, butterflies , wasps	fruit set	
leek (A)	(36) Fijen, thijs.fijen@wur.nl	<i>Allium porrum</i>	France, Loire	2016	18 (18)	active	bees, wasps, hoverflies	seed set	plant yield
leek (B)	(36) Fijen, thijs.fijen@wur.nl	<i>Allium porrum</i>	Italy, South Italy	2016	18 (18)	active	bees, wasps, hoverflies	seed set	plant yield
lemon	Chacoff, nchacoff@gmail.com	<i>Citrus limon</i>	Argentina, Yungas	2015	9	active	bee, flies, butterflies , wasps	fruit set	-
mango (A)	(84) Carvalheiro, lgcarvalheiro@gmail.com	<i>Mangifera indica</i>	South Africa, Limpopo	2008	8	active	bee, flies, butterflies , beetles, wasps	fruit set	-
mango (B)	(85) Carvalheiro, lgcarvalheiro@gmail.com	<i>Mangifera indica</i>	South Africa, Limpopo	2009	14 (10)	active	bee, flies, butterflies , beetles, wasps	fruit set	plant yield
mango (C)	Rader, rrader@une.edu.au	<i>Mangifera indica</i>	Australia, Queensland	2014	10	active	bees, flies, hoverflies, beetles, moths, butterflies	fruit set	-
mango (D)	Willcox, bwillcox@myune.edu.au	<i>Mangifera indica</i>	Australia, Queensland	2016	7	active	bees, flies, hoverflies, beetles, moths, butterflies	fruit set	-
osr (A)	Andersson,	<i>Brassica napus</i>	Sweden, Scania	2010	6	active	bees,	seed set	-

	gandersson@unrn.edu.ar						hoverflies		
osr (B)	(86) Bartomeus, Gagic, nacho.bartomeus@gmail.com vesna.gagic@bio.bg.ac.rs	<i>Brassica napus</i>	Sweden, Västergötland	2013	12 (9)	active	bees, butterflies	seed set	crop yield
osr (C)	(69) Garratt, m.p.garratt@reading.ac.uk	<i>Brassica napus</i>	UK, Yorkshire	2012	8	active, passive	bees	seed set	-
osr (D)	(87, 88) Stanley, dara.stanley@ucd.ie	<i>Brassica napus</i>	Ireland, South-East	2010	3	active	bees, hoverflies	seed set	-
osr (E)	Sutter, louis.sutter@agroscope.admin.ch	<i>Brassica napus</i>	Switzerland, Zurich	2014	18 (18)	active	bees, hoverflies	seed set	crop yield
osr (F)	(89) Zou Yi, yi.zou.1@hotmail.com	<i>Brassica napus</i>	China, Jiangxi	2015	18	passive	bees, hoverflies, butterflies	fruit set	-
raspberry	(42) Saez, agustinsaez@live.com.ar	<i>Rubus idaeus</i>	Argentina, Comarca Andina	2014	16 (16)	active	bees	fruit set	crop yield
red clover	Rundlöf, maj.rundlof@biol.lu.se	<i>Trifolium pratense</i>	Sweden, Scania	2013	6 (6)	active	bees	seed set	crop yield
strawberry (A)	Andersson, gandersson@unrn.edu.ar	<i>Fragaria × ananassa</i>	Sweden, Scania	2009	11	passive	bees, hoverflies	fruit set	-
strawberry (B)	Baensch, Tschamtk, Westphal, svenja.baensch@agr.uni-goettingen.de cwestph@gwdg.de ttschar@gwdg.de	<i>Fragaria × ananassa</i>	Germany, Lower Saxony,	2015	8 (8)	active	bees	Δ fruit weight	plant yield
strawberry (C1)	(90) Grab, hlc66@cornell.edu	<i>Fragaria × ananassa</i>	USA, New York	2012	11 (10)	active, passive	bees	Δ fruit weight	plant yield
strawberry (C2)	Grab, hlc66@cornell.edu	<i>Fragaria × ananassa</i>	USA, New York	2014	27 (27)	active	bees	seed set	plant yield
strawberry (C3)	(91) Grab, hlc66@cornell.edu	<i>Fragaria × ananassa</i>	USA, New York	2015	14 (14)	active	bees	seed set	plant yield
strawberry (D)	Garratt, m.p.garratt@reading.ac.uk	<i>Fragaria × ananassa</i>	UK, Yorkshire	2011	7 (7)	active, passive	bees	Δ fruit weight	plant yield
strawberry (E)	Klatt, klattbk@googlemail.com	<i>Fragaria × ananassa</i>	Germany, Lower Saxony	2010	8 (8)	active	bees	fruit set	plant yield
strawberry (F)	Krewenka, kristin.marie.krewenka@uni-hamburg.de	<i>Fragaria × ananassa</i>	Germany, Lower Saxony	2005	10 (10)	active	bees	fruit set	crop yield
strawberry (G)	Sciligo, amber.sciligo@berkeley.edu	<i>Fragaria × ananassa</i>	USA, California	2012	15 (15)	active, passive	bees	Δ fruit weight	plant yield
strawberry (H)	(92) Stewart, rebecca.stewart@cec.lu.se	<i>Fragaria × ananassa</i>	Sweden, Scania	2014	27 (27)	active	hoverflies	fruit set	plant yield
sunflower (A)	(45) Carvalheiro, lgcarvalheiro@gmail.com	<i>Helianthus annuus</i>	South Africa, Limpopo	2009	28	active	bee, flies, butterflies, beetles, wasps	seed set	-
sunflower (B)	Scheper, jeroen.scheper@wur.nl	<i>Helianthus annuus</i>	France, Poitou-Charentes	2015	24	active	bees, hoverflies	seed set	-
Pest control studies									
apple (A1)	(93, 94) Lavigne, claire.lavigne@inra.fr	<i>Malus domestica</i>	France, Provence-Alpes-Côte d'Azur	2006	9	active	parasitoids	sentinel exp. (enemy	-

								activity)	
apple (A2)	(93, 94) Lavigne, claire.lavigne@inra.fr	<i>Malus domestica</i>	France, Provence- Alpes-Côte d'Azur	2007	6	active	parasitoid s	sentinel exp. (enemy activity)	-
apple (A3)	(93, 94) Lavigne, claire.lavigne@inra.fr	<i>Malus domestica</i>	France, Provence- Alpes-Côte d'Azur	2008	17	active	parasitoid s	sentinel exp. (enemy activity)	-
apple (A4)	(93, 94) Lavigne, claire.lavigne@inra.fr	<i>Malus domestica</i>	France, Provence- Alpes-Côte d'Azur	2009	12	active	parasitoid s	sentinel exp. (enemy activity)	-
apple (A5)	(93, 94) Lavigne, claire.lavigne@inra.fr	<i>Malus domestica</i>	France, Provence- Alpes-Côte d'Azur	2010	14	active	parasitoid s	sentinel exp. (enemy activity)	-
barley (A)	(95) Caballero-Lopez, bcaballero@bcn.cat	<i>Hordeum vulgare</i>	Sweden, Scania	2007	20	active, passive	carabids, ladybugs, parasitoid s	sentinel exp. (enemy activity)	-
barley (B)	(96–98) Tamburini, giovanni.tamburini@slu.se	<i>Hordeum vulgare</i>	Italy, Friuli Venezia- Giulia	2014	5 (5)	passive	carabids	cage exp. (infestation)	crop yield
buckwheat	(99) Taki, htaki@affrc.go.jp	<i>Fagopyrum esculentum</i>	Japan, Ibaraki	2008	15	passive	ladybugs, lacewings	sentinel exp. (enemy activity)	-
cabbage	(100) Letourneau, dletour@ucsc.edu	<i>Brassica oleracea</i>	USA, Monterey Bay Area	2006	33	passive	parasitoid s	sentinel exp. (enemy activity)	-
cacao	(101) Maas, beamaas@gmx.at	<i>Theobroma cacao</i>	Indonesia, Sulawesi	2010	15 (15)	active	spiders	cage exp. (crop damage)	plant yield
coffee (A)	Schleuning, Schmack, Matthias.Schleuning@senckenberg.de juliaschmack@gmx.de	<i>Coffea arabica</i>	Tanzania, Kilimanjaro	2011/2012	11 (6)	passive	bats, birds	cage exp. (crop damage)	plant yield
coffee (B)	Iverson, iverson@umich.edu	<i>Coffea arabica</i>	Mexico, Soconusco	2012	37 (35)	passive	parasitoid s	sentinel exp. (enemy activity)	crop yield
coffee (C)	Iverson, iverson@umich.edu	<i>Coffea arabica</i>	Puerto Rico, Utuado	2013	36	passive	parasitoid s, wasps	cage exp. (crop damage)	-
coffee (D)	Martinez-Salinas, amartinez@catie.ac.cr	<i>Coffea arabica</i>	Costa Rica, Turrialba	2013	10	passive	birds	cage exp. (crop damage)	-
maize	(102) O'Rourke, megorust@vt.edu	<i>Zea mays</i>	USA, New York	2006	26	passive	ladybugs	pest damage	-
osr (A)	(103) Jonsson, mattias.jonsson@slu.se	<i>Brassica napus</i>	New Zealand, Canterbury	2007	26	active	hoverflies, ladybugs, lacewings	pest damage	-
osr (B)	Sutter, louis.sutter@agroscope.admin.ch	<i>Brassica napus</i>	Switzerland, Zurich	2014	18 (18)	passive	carabids	sentinel exp. (enemy activity)	crop yield
pomegranate	(104) Keasar, tkeasar@research.haifa.ac.il	<i>Punica granatum</i>	Israel, Hefer Valley	2014	10	active	spiders, parasitoid s	pest damage	-
potato (A)	(105) Martin,	<i>Solanum tuberosum</i>	South Korea, Haeon	2009	6 (2)	active,	birds,	pest damage	plant yield

	emily.martin@uni-wuerzburg.de					passive	carabids, hoverflies, parasitoids, rove beetles, wasps		
potato (B)	(106) Poveda, kap235@cornell.edu	<i>Solanum tuberosum</i>	Colombia, Cundinamarca	2007	11 (11)	active, passive	carabids, hoverflies, ladybugs, lacewings, parasitoids	pest damage	crop yield
radish	(105) Martin, emily.martin@uni-wuerzburg.de	<i>Raphanus raphanistrum</i> subsp. <i>sativus</i>	South Korea, Haean	2009	8 (5)	active, passive	birds, carabids, hoverflies, parasitoids, rove beetles, wasps	pest damage	plant yield
rice	(107) Takada, mayura@isas.a.u-tokyo.ac.jp	<i>Oryza sativa</i>	Japan, Miyagi	2008	44	active	spiders	pest damage	-
soybean (A)	(37) Kim, tkim@glbrc.wisc.edu	<i>Glycine max</i>	USA, Upper Midwest	2012	35 (33)	passive	flower bugs, ladybugs	cage exp. (infestation)	plant yield
soybean (B1)	(108) Mitchell, matthew.mitchell@ubc.ca	<i>Glycine max</i>	Canada, Montérégie	2010	15 (15)	active	hoverflies, ladybugs, lacewings, true bugs	pest damage	crop yield
soybean (B2)	(108) Mitchell, matthew.mitchell@ubc.ca	<i>Glycine max</i>	Canada, Montérégie	2011	19 (19)	active	hoverflies, ladybugs, lacewings, true bugs	pest damage	crop yield
soybean (C1)	Molina, gonzalormolina@agro.uba.ar	<i>Glycine max</i>	Argentina, North Buenos Aires	2011	20	active	parasitoids	sentinel exp. (enemy activity)	-
soybean (C2)	Molina, gonzalormolina@agro.uba.ar	<i>Glycine max</i>	Argentina, North Buenos Aires	2012	20	active	parasitoids	sentinel exp. (enemy activity)	-
soybean (D)	(105) Martin, emily.martin@uni-wuerzburg.de	<i>Glycine max</i>	South Korea, Haean	2009	8 (6)	active, passive	birds, carabids, hoverflies, parasitoids, rove beetles, wasps	pest damage	plant yield
wheat (A)	(109) Bommarco, riccardo.bommarco@slu.se	<i>Triticum aestivum</i>	Sweden, Scania	2007	31 (31)	passive	carabids	sentinel exp. (enemy activity)	crop yield
wheat (B)	(95) Caballero-Lopez, bcaballero@bcn.cat	<i>Triticum aestivum</i>	Sweden, Scania	2007	4	active, passive	carabids, ladybugs,	sentinel exp. (enemy	-

							parasitoids	activity)	
wheat (C)	Kim, tkim@glbrc.wisc.edu	<i>Triticum aestivum</i>	USA, Upper Midwest	2012	24 (24)	active, passive	flower bugs, ladybugs	cage exp. (infestation)	plant yield
wheat (D1)	(110) Plečas, mplecas@bio.bg.ac.rs	<i>Triticum aestivum</i>	Serbia, Pacevacki Rit	2008	18	active	parasitoids	sentinel exp. (enemy activity)	-
wheat (D2)	(110) Plečas, mplecas@bio.bg.ac.rs	<i>Triticum aestivum</i>	Serbia, Pacevacki Rit	2009	17	active	parasitoids	sentinel exp. (enemy activity)	-
wheat (D3)	(110) Plečas, mplecas@bio.bg.ac.rs	<i>Triticum aestivum</i>	Serbia, Pacevacki Rit	2010	8	active	parasitoids	sentinel exp. (enemy activity)	-
wheat (D4)	(110) Plečas, mplecas@bio.bg.ac.rs	<i>Triticum aestivum</i>	Serbia, Pacevacki Rit	2011	10	active	parasitoids	sentinel exp. (enemy activity)	-
wheat (E)	(96–98) Tamburini, giovanni.tamburini@slu.se	<i>Triticum aestivum</i>	Italy, Friuli Venezia- Giulia	2014	11 (11)	passive	carabids	cage exp. (infestation)	crop yield
wheat (F)	(111) Tschumi, matthias.tschumi@vogelwarte.ch	<i>Triticum aestivum</i>	Switzerland, Central Plateau	2012	25	active, passive	carabids, ladybugs, true bugs	pest damage	-

Table S2. Model output for richness–ecosystem service relationships. (A) Richness was calculated as the number of unique taxa sampled per study. (B) Richness was calculated considering only organisms classified at the fine taxonomy level (i.e. species- or morphospecies-levels). Posterior samples were summarized based on the Bayesian point estimate (median), standard error (median absolute deviation), and 80%, 90% and 95% highest density intervals (HDI). HDIs that do not include zero are reported in bold.

(A)

Parameter	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Pollination</i>					
Intercept	0.0004	0.0311	[-0.0419, 0.0411]	[-0.0531, 0.0544]	[-0.0670, 0.0622]
Pollinator richness	0.1532	0.0353	[0.1062, 0.1962]	[0.0951, 0.2110]	[0.0865, 0.2266]
<i>Pest control</i>					
Intercept	-0.0003	0.0353	[-0.0434, 0.0485]	[-0.0579, 0.0589]	[-0.0724, 0.0657]
Natural enemy richness	0.2093	0.0417	[0.1551, 0.2618]	[0.1415, 0.2779]	[0.1283, 0.2932]

(B)

Parameter	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Pollination</i>					
Intercept	0.0010	0.0333	[-0.0409, 0.0421]	[-0.0536, 0.0537]	[-0.0662, 0.0617]
Pollinator richness	0.1535	0.0356	[0.1096, 0.2006]	[0.0967, 0.2141]	[0.0848, 0.2256]
<i>Pest control</i>					
Intercept	0.0001	0.0401	[-0.0536, 0.0514]	[-0.0712, 0.0646]	[-0.0834, 0.0775]
Natural enemy richness	0.2264	0.0484	[0.1638, 0.2861]	[0.1475, 0.3065]	[0.1254, 0.3199]

Table S3. Model output for path models testing direct and indirect effects (mediated by changes in abundance) of richness on ecosystem services. Posterior samples were summarized based on the Bayesian point estimate (median), standard error (median absolute deviation), and 80%, 90% and 95% highest density intervals (HDIs). HDIs that do not include zero are reported in bold.

Effect	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Pollination</i>					
Richness → Pollination	0.1058	0.0428	[0.0511, 0.1635]	[0.0326, 0.1779]	[0.0199, 0.1933]
Richness → Abundance	0.5701	0.0379	[0.5222, 0.6212]	[0.5044, 0.6319]	[0.4900, 0.6449]
Abundance → Pollination	0.0804	0.0460	[0.0232, 0.1401]	[0.0057, 0.1564]	[-0.0140, 0.1665]
<i>Pest control</i>					
Richness → Pest control	0.1413	0.0434	[0.0832, 0.1951]	[0.0684, 0.2105]	[0.0564, 0.2275]
Richness → Abundance	0.4447	0.0494	[0.3782, 0.5070]	[0.3646, 0.5315]	[0.3467, 0.5452]
Abundance → Pest control	0.1481	0.0553	[0.0772, 0.2170]	[0.0612, 0.2406]	[0.0467, 0.2619]

Table S4. Model output for path models testing direct and indirect effects (mediated by changes in evenness) of richness on ecosystem services. Posterior samples were summarized based on the Bayesian point estimate (median), standard error (median absolute deviation), and 80%, 90% and 95% highest density intervals (HDIs). HDIs that do not include zero are reported in bold.

Effect	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Pollination</i>					
Richness → Pollination	0.1580	0.0361	[0.1105, 0.2033]	[0.0978, 0.2175]	[0.0863, 0.2287]
Richness → Evenness	0.0900	0.0578	[0.0171, 0.1653]	[-0.0095, 0.1804]	[-0.0269, 0.1991]
Evenness → Pollination	-0.0719	0.0390	[-0.1238, -0.0240]	[-0.1419, -0.0127]	[-0.1525, 0.0006]
<i>Pest control</i>					
Richness → Pest control	0.2298	0.0415	[0.1748, 0.2832]	[0.1619, 0.3041]	[0.1444, 0.3153]
Richness → Evenness	0.2358	0.0767	[0.1345, 0.3313]	[0.1011, 0.3560]	[0.0804, 0.3854]
Evenness → Pest control	-0.0844	0.0430	[-0.1393, -0.0302]	[-0.1587, -0.0165]	[-0.1683, 0.0027]

Table S5. Model output for path models testing direct and indirect effects (mediated by changes in richness) of landscape simplification on ecosystem services. Posterior samples were summarized based on the Bayesian point estimate (median), standard error (median absolute deviation), and 80%, 90% and 95% highest density intervals (HDIs). HDIs that do not include zero are reported in bold.

Effect	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Pollination</i>					
Landscape → Pollination	-0.0573	0.0409	[-0.1083, -0.0041]	[-0.1203, 0.0147]	[-0.1374, 0.0229]
Landscape → Richness	-0.1984	0.0453	[-0.2593, -0.1430]	[-0.2750, -0.1263]	[-0.2909, -0.1119]
Richness → Pollination	0.1543	0.0362	[0.1060, 0.1992]	[0.0937, 0.2148]	[0.0815, 0.2278]
<i>Causal mediation analysis</i>					
Direct effect	-0.0573		[-0.1083, -0.0041]	[-0.1203, 0.0147]	[-0.1374, 0.0229]
Indirect effect	-0.0293		[-0.0425, -0.0168]	[-0.0465, -0.0136]	[-0.0515, -0.0117]
Total effect	-0.0859		[-0.1391, -0.0361]	[-0.1560, -0.0239]	[-0.1642, -0.0074]
Proportion mediated	34.0%				-
<i>Pest control</i>					
Landscape → Pest control	-0.0285	0.0442	[-0.0864, -0.0289]	[-0.1043, 0.0461]	[-0.1248, 0.0570]
Landscape → Richness	-0.1510	0.0479	[-0.2123, -0.0886]	[-0.2299, -0.0706]	[-0.2491, -0.0581]
Richness → Pest control	0.2114	0.0418	[0.1609, 0.2682]	[0.1429, 0.2810]	[0.1315, 0.2962]
<i>Causal mediation analysis</i>					
Direct effect	-0.0285		[-0.0864, -0.0289]	[-0.1043, 0.0461]	[-0.1248, 0.0570]
Indirect effect	-0.0311		[-0.0460, -0.0149]	[-0.0523, -0.0118]	[-0.0578, -0.0083]
Total effect	-0.0610		[-0.1214, -0.0060]	[-0.1378, 0.0120]	[-0.1511, 0.0301]
Proportion mediated	50.9%				

Table S6. Model output for path models testing the direct and cascading landscape simplification effects on ecosystem services via changes in richness and abundance.

Posterior samples were summarized based on the Bayesian point estimate (median), standard error (median absolute deviation), and 80%, 90% and 95% highest density intervals (HDIs).

HDIs that do not include zero are reported in bold.

Effect	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Pollination</i>					
Landscape → Richness	-0.1991	0.0458	[-0.2593, -0.1431]	[-0.2779, -0.1269]	[-0.2918, -0.1109]
Landscape → Abundance	-0.1914	0.0462	[-0.2503, -0.1302]	[-0.2721, -0.1167]	[-0.2812, -0.0955]
Landscape → Pollination	-0.0559	0.0390	[-0.1043, -0.0035]	[-0.1223, 0.0078]	[-0.1351, 0.0215]
Richness → Pollination	0.1082	0.0430	[0.0517, 0.1629]	[0.0366, 0.1810]	[0.0197, 0.1924]
Abundance → Pollination	0.0721	0.0444	[0.0155, 0.1313]	[-0.0017, 0.1479]	[-0.0229, 0.1558]
<i>Pest control</i>					
Landscape → Richness	-0.1515	0.0471	[-0.2160, -0.0939]	[-0.2322, -0.0730]	[-0.2430, -0.0471]
Landscape → Abundance	-0.0880	0.0511	[-0.1617, -0.0240]	[-0.1727, 0.0044]	[-0.1968, 0.0148]
Landscape → Pest control	-0.0250	0.0436	[-0.0785, 0.0316]	[-0.0971, 0.0451]	[-0.1128, 0.0559]
Richness → Pest control	0.1524	0.0436	[0.0928, 0.2049]	[0.0822, 0.2272]	[0.0642, 0.2385]
Abundance → Pest control	0.1282	0.0540	[0.0597, 0.1967]	[0.0398, 0.2146]	[0.0323, 0.2403]

Table S7. Model output for path models testing the direct and cascading landscape simplification effects on ecosystem services via changes in richness and evenness. Posterior samples were summarized based on the Bayesian point estimate (median), standard error (median absolute deviation), and 80%, 90% and 95% highest density intervals (HDIs). HDIs that do not include zero are reported in bold.

Effect	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Pollination</i>					
Landscape → Richness	-0.1973	0.0461	[-0.2527, -0.1345]	[-0.2739, -0.1220]	[-0.2894, -0.1091]
Landscape → Evenness	0.1006	0.0447	[0.0433, 0.1571]	[0.0268, 0.1723]	[0.0159, 0.1906]
Landscape → Pollination	-0.0554	0.0404	[-0.1074, -0.0024]	[-0.1227, 0.0118]	[-0.1352, 0.0258]
Richness → Pollination	0.1591	0.0373	[0.1117, 0.2061]	[0.0997, 0.2203]	[0.0879, 0.2316]
Evenness → Pollination	-0.0583	0.0388	[-0.1069, -0.0091]	[-0.1198, 0.0073]	[-0.1346, 0.0161]
<i>Pest control</i>					
Landscape → Richness	-0.1480	0.0487	[-0.2056, -0.0819]	[-0.2269, -0.0659]	[-0.2465, -0.0540]
Landscape → Evenness	-0.0538	0.0554	[-0.1221, 0.0204]	[-0.1510, 0.0334]	[-0.1599, 0.0617]
Landscape → Pest control	-0.0319	0.0443	[-0.0869, 0.0273]	[-0.1054, 0.0416]	[-0.1182, 0.0594]
Richness → Pest control	0.2260	0.0431	[0.1704, 0.2803]	[0.1563, 0.2979]	[0.1440, 0.3135]
Evenness → Pest control	-0.0717	0.0433	[-0.1299, -0.0196]	[-0.1419, 0.0009]	[-0.1537, 0.0152]

Table S8. Model output for path models testing the direct and cascading landscape simplification effects on final crop production via changes in richness, evenness, and ecosystem services. Posterior samples were summarized based on the Bayesian point estimate (median), standard error (median absolute deviation), and 80%, 90% and 95% highest density intervals (HDIs). HDIs that do not include zero are reported in bold.

Effect	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Pollination</i>					
Landscape → Richness	-0.1709	0.0589	[-0.2431, -0.892]	[-0.2697, -0.0696]	[-0.2984, -0.0494]
Landscape → Evenness	0.0837	0.0549	[0.0117, 0.1540]	[-0.0163, 0.1710]	[-0.0306, 0.1956]
Richness → Pollination	0.1829	0.0504	[0.1205, 0.2495]	[0.1011, 0.2665]	[0.0818, 0.2799]
Evenness → Pollination	-0.0714	0.0515	[-0.1415, -0.0075]	[-0.1607, 0.0116]	[-0.1744, 0.0326]
Pollination → Production	0.3344	0.0862	[0.2279, 0.4536]	[0.2012, 0.4901]	[0.1707, 0.5178]
<i>Pest control</i>					
Landscape → Richness	-0.2225	0.0881	[-0.3406, -0.1075]	[-0.3771, -0.0697]	[-0.4019, -0.0223]
Landscape → Evenness	-0.0287	0.0992	[-0.1680, 0.0950]	[-0.2045, 0.1477]	[-0.2484, 0.1874]
Richness → Pest control	0.2145	0.0797	[0.1123, 0.3174]	[0.0790, 0.3447]	[0.0614, 0.3798]
Evenness → Pest control	-0.1251	0.0808	[-0.2346, -0.0248]	[-0.258, 0.0134]	[-0.2959, 0.0341]
Pest control → Production	0.1483	0.0823	[0.0377, 0.2488]	[0.0133, 0.2877]	[-0.0094, 0.3213]

Table S9. Model output for path models testing the direct and cascading landscape simplification effects on final crop production via changes in richness, abundance, and ecosystem services. Posterior samples were summarized based on the Bayesian point estimate (median), standard error (median absolute deviation), and 80%, 90% and 95% highest density intervals (HDIs). HDIs that do not include zero are reported in bold.

Effect	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Pollination</i>					
Landscape → Richness	-0.1870	0.0571	[-0.2631, -0.1128]	[-0.2876, -0.0926]	[-0.3067, -0.0722]
Landscape → Abundance	-0.1988	0.0542	[-0.2680, -0.1270]	[-0.2915, -0.1069]	[-0.3103, -0.0884]
Richness → Pollination	0.1477	0.0638	[0.0645, 0.2281]	[0.0485, 0.2568]	[0.0212, 0.2712]
Abundance → Pollination	0.0104	0.0667	[-0.0742, 0.0961]	[-0.0951, 0.1243]	[-0.1250, 0.1373]
Pollination → Production	0.3388	0.0868	[0.2268, 0.4509]	[0.1910, 0.4813]	[0.1549, 0.5070]
<i>Pest control</i>					
Landscape → Richness	-0.2073	0.0840	[-0.3197, -0.0977]	[-0.3502, -0.0554]	[-0.3915, -0.0216]
Landscape → Abundance	-0.0304	0.1060	[-0.1759, 0.1106]	[-0.2242, 0.1587]	[-0.2745, 0.1938]
Richness → Pest control	0.2255	0.0786	[0.1201, 0.3236]	[0.0932, 0.3573]	[0.0730, 0.3905]
Abundance → Pest control	0.0040	0.0793	[-0.1016, 0.1064]	[-0.1331, 0.1413]	[-0.1572, 0.1769]
Pest control → Production	0.1395	0.0786	[0.0404, 0.2451]	[0.0151, 0.2822]	[-0.0257, 0.3011]

Table S10. Model output for path models testing direct and indirect effects of pollinator richness, abundance, and evenness (with honey bees) on pollination. Posterior samples were summarized based on the Bayesian point estimate (median), standard error (median absolute deviation), and 80%, 90% and 95% highest density intervals (HDIs). HDIs that do not include zero are reported in bold.

Effect	Estimate	SE	HDI (80%)	HDI (90%)	HDI (95%)
<i>Model 1</i>					
Richness → Pollination	0.1043	0.0419	[0.0524, 0.1588]	[0.0356, 0.1715]	[0.0250, 0.1878]
Richness → Abundance	0.5160	0.0377	[0.4682, 0.5659]	[0.4511, 0.5784]	[0.4328, 0.5882]
Abundance → Pollination	0.1183	0.0452	[0.0617, 0.1768]	[0.0430, 0.1903]	[0.0278, 0.2038]
<i>Model 2</i>					
Richness → Pollination	0.1722	0.0370	[0.1272, 0.2199]	[0.1110, 0.2294]	[0.1009, 0.2420]
Richness → Evenness	0.0027	0.0551	[-0.0633, 0.0788]	[-0.0941, 0.0886]	[-0.1067, 0.1101]
Evenness → Pollination	-0.1148	0.0365	[-0.1598, -0.0647]	[-0.1759, -0.0544]	[-0.1890, -0.0438]

Table S11. Results of pairwise comparison of richness–ecosystem service relationships according to the methods used to sample pollinators and natural enemies. A Bayesian hypothesis testing was used to assess the relative statistical evidence in favor of the null hypothesis versus the alternative hypothesis.

Hypothesis	Estimate difference	Estimate Error	CI lower	CI upper	Evidence Ratio
Pollination					
Active > Passive	-0.02	0.10	-0.18	Inf	0.78
Pest control					
Active > Passive	0.04	0.08	-0.01	Inf	2.17

Table S12. Results of pairwise comparison of richness–ecosystem service relationships according to the methods used to quantify pollination and pest control services. A Bayesian hypothesis testing was used to assess the relative statistical evidence in favor of the null hypothesis versus the alternative hypothesis.

Hypothesis	Estimate difference	Estimate Error	CI lower	CI upper
Pollination				
Fruit set = Δ Fruit weight	0.11	0.13	-0.15	0.37
Fruit set = Seed set	-0.10	0.08	-0.26	0.06
Δ Fruit weight = Seed set	0.22	0.15	-0.07	0.50
Pest control				
Cage (damage) = Cage (infestation)	0.09	0.18	-0.26	0.44
Cage (damage) = Pest damage	0.17	0.15	-0.12	0.47
Cage (damage) = Sentinel experiments	0.05	0.15	-0.24	0.34
Cage (infestation) = Pest damage	0.08	0.14	-0.19	0.36
Cage (infestation) = Sentinel experiments	-0.04	0.13	-0.30	0.22
Pest damage = Sentinel experiments	-0.12	0.10	-0.32	0.07