

Decoupled genomic elements and the evolution of partner quality in nitrogen-fixing rhizobia

Benjamin R. Gordon¹, Christie R. Klinger¹, Dylan J. Weese², Jennifer A. Lau³, Patricia V. Burke¹, Bryn T. M. Dentinger⁴ & Katy D. Heath¹

¹Department of Plant Biology, University of Illinois Urbana-Champaign, 505 S. Goodwin Ave., Urbana, Illinois 61801

²Department of Biology, St. Ambrose University, 518 West Locust St, Davenport, Iowa 52803

³Kellogg Biological Station and Department of Plant Biology, Michigan State University, 3700 E. Gull Lake Drive, Hickory Corners, Michigan 49060

⁴Jodrell Laboratory, Royal Botanic Gardens, Kew, Richmond, Surrey TW9 3DS, United Kingdom

Keywords

Cheating, mutualism, nitrogen deposition, partner quality, Rhizobium, symbiosis.

Correspondence

Katy D. Heath, Department of Plant Biology, University of Illinois Urbana-Champaign, 505 S. Goodwin Ave., Urbana, IL 61801.
Tel: 1-217-265-5473;
Fax: 1-217-244-7246;
E-mail: kheath@life.illinois.edu

Funding Information

This work was supported by NSF DEB-1257938. Support for this research was also provided by the NSF Long-term Ecological Research Program (DEB 1027253) at the Kellogg Biological Station and by Michigan State University AgBioResearch.

Received: 2 October 2015; Revised: 14 December 2015; Accepted: 15 December 2015

Ecology and Evolution 2016; 6(5): 1317–1327

doi: 10.1002/ece3.1953

Introduction

N (Nitrogen) is a critical nutrient for all plants and animals, and N availability is a major determinant of ecology in terrestrial communities (Sterners and Elser 2002; Craine 2009; Grman et al. 2010). Human activities are drastically altering the N cycle via fossil fuel combustion, the Haber–Bosch process (an industrial process that reduces N₂ to NH₄⁺), and modern agricultural practices – all of which add millions of tons of N to terrestrial environments globally. These anthropogenic activities represent 45% of all N fixed on Earth, and they double the rate of natural terrestrial N fixation, which is primarily driven by microbial processes and supplemented by heat shock

Abstract

Understanding how mutualisms evolve in response to a changing environment will be critical for predicting the long-term impacts of global changes, such as increased N (nitrogen) deposition. Bacterial mutualists in particular might evolve quickly, thanks to short generation times and the potential for independent evolution of plasmids through recombination and/or HGT (horizontal gene transfer). In a previous work using the legume/rhizobia mutualism, we demonstrated that long-term nitrogen fertilization caused the evolution of less-mutualistic rhizobia. Here, we use our 63 previously isolated rhizobium strains in comparative phylogenetic and quantitative genetic analyses to determine the degree to which variation in partner quality is attributable to phylogenetic relationships among strains versus recent genetic changes in response to N fertilization. We find evidence of distinct evolutionary relationships between chromosomal and pSym genes, and broad similarity between pSym genes. We also find that *nifD* has a unique evolutionary history that explains much of the variation in partner quality, and suggest MoFe subunit interaction sites in the evolution of less-mutualistic rhizobia. These results provide insight into the mechanisms behind the evolutionary response of rhizobia to long-term N fertilization, and we discuss the implications of our results for the evolution of the mutualism.

associated with lightning strikes (Canfield et al. 2010). It is estimated that up to 50% of anthropogenic N fertilizer is lost to water, the atmosphere, and uncultivated soils (Vitousek 1997; Vitousek et al. 1997; Smil 2002). The ecological effects of this N deposition are detrimental to natural communities, ranging from decreases in biodiversity (Tilman 1987; Clark and Tilman 2008) to coastal dead zones arising from eutrophication (Diaz and Rosenberg 2008). Furthermore, increases in N availability are predicted to cause the breakdown of a resource mutualism of major ecological and economic importance – that between legumes and symbiotic rhizobia (Sprent and Sutherland 1987; West et al. 2002; Kiers et al. 2006; Oono et al. 2009; Sachs et al. 2011; Frederickson 2013).

Mutualisms, cooperative partnerships between species, are both diverse and widespread, playing key roles in the evolution and ecology of land-plants, marine ecosystems, the biology of herbivores, and more (Saffo 2014). The diverse roles played by mutualisms, such as the maintenance of biodiversity and nutrient cycling (Stachowicz 2001), are widely distributed globally (Brussaard *et al.* 2007), and of recognized economic importance (Pimentel *et al.* 1997; Vitousek *et al.* 1997). Despite the apparent long-term stability of many mutualistic relationships, models predict that mutualisms should be extremely susceptible to the invasion of less-beneficial genotypes, which give less than they take from their symbiotic partners (West *et al.* 2007), leading to mutualism breakdown (Sachs *et al.* 2004; Holland and DeAngelis 2010). Such mutualism breakdown might be exacerbated under global change (Six 2009; Kiers *et al.* 2010). This prediction arises from a large body of work indicating that mutualisms are quite context-dependent, making cost-benefit ratios for partner fitness, and ultimately mutualism stability, highly contingent upon the surrounding environmental conditions (Bronstein 1994; Chamberlain *et al.* 2014). Moreover, because partner species often have very different physiologies, phylogenetic origins, and environmental responses, the frequencies with which partner species interact might also shift in response to a rapidly changing environment and lead to mutualism breakdown (Sachs and Simms 2006).

Leguminous plants obtain N by pairing with soil bacteria called rhizobia, which are housed in root nodules and therein fix atmospheric dinitrogen (N_2) to ammonium (NH_4^+) in exchange for plant photosynthates. Given that N is a key traded resource in this mutualistic symbiosis, theory predicts that excess N will destabilize the legume-rhizobium mutualism and lead to the evolution of less-cooperative rhizobia through several potential selective mechanisms (Akçay and Simms 2011). While host control mechanisms that favor cooperative rhizobium partners have been shown to remain active in high N, at least in the short-term (Kiers *et al.* 2006; Regus *et al.* 2014), long-term studies are needed to understand how the mutualism evolves when exposed to increased N over many generations. Indeed in a recent study, Weese *et al.* (2015) demonstrated that populations of *Rhizobium leguminosarum* bv. *trifolii* (*Rlt*) were less beneficial for their clover host plants after long-term exposure (22 years) to N fertilizer, and moreover that these N-evolved versus control strains were not phylogenetically distinct (i.e., were not separate, monophyletic lineages). Nevertheless, the evolutionary processes leading to this phenotypic differentiation were not investigated.

Rhizobia are excellent models for studying the evolution and breakdown of cooperation in changing

environments, and for microbial adaptation in general. Bacteria are experimentally tractable, have short generation times, and harbor relatively small genomes, simplifying the detection of evolutionary responses (Elena and Lenski 2003). Ecological shifts in taxonomic composition and/or abundance have been shown to occur within natural bacterial communities in response to changing environments (Allison and Martiny 2008; Logares *et al.* 2013; Newton *et al.* 2013; Youngblut *et al.* 2013). Yet despite recent investigations, we still lack fundamental knowledge on how bacterial populations in nature evolve in response to environmental changes. While adaptive variants in bacteria can arise from point mutations (Stallforth *et al.* 2013), bacteria can also evolve via recombination, or by HGT (horizontal gene transfer) (Gogarten *et al.* 2002; Ochman & Moran 2001), “the non-genealogical transmission of genetic material from one organism to another” (Goldenfeld & Woese 2007). HGT can potentially facilitate rapid evolutionary responses to environmental stress (Jain *et al.* 2003; Beaber *et al.* 2004), or to exposure to correlated environments of increased complexity (Mozhayskiy and Tagkopoulos 2012). If recombination is infrequent compared to mutation, then the entire genome should sweep to fixation along with a newly selected locus (i.e., the “ecotype model” (Koeppl *et al.* 2008)); by contrast, frequent recombination in natural bacterial populations could break up the linkage between selected and other loci (i.e., similar to classical models of eukaryotic adaptive evolution). Existing data on recombination within and among genomic elements in rhizobia is mixed (Orozco-Mosqueda *et al.* 2009), and moreover depends on the phylogenetic scale analyzed. Some studies have found congruent phylogenies consistent with a general lack of recombination at the species level in contrast to abundant recombination at the population level (Wernegreen *et al.* 1997; Kumar *et al.* 2015), while others have found a high level of consistency between the chromosome and symbiosis plasmids even at the population scale (Wernegreen and Riley 1999; Bailly *et al.* 2007; Epstein *et al.* 2012). Studies of other microbial species suggest that recombination is abundant (Reno *et al.* 2009; Cadillo-Quiroz *et al.* 2012; Shapiro *et al.* 2012; Held *et al.* 2013), and that the occurrence of genome-wide sweeps is rare (Polz *et al.* 2013).

The *Rhizobium leguminosarum* reference genome (bv. *trifolii* WSM1689) has a 4.85 Mb chromosome and five plasmids varying in size from 259 to 609 Kb (Genbank accession GCA_000517615.1). In rhizobia, key genes necessary for nodulation (*nod*) and N fixation (*nif* and *fix*) have been shown to cluster (Young *et al.* 2006), and often reside either on a symbiosis plasmid (pSym) or as part of a genomic SI (symbiosis island). In *Rlt*, they have previously been shown to localize to the pSym

(López-Guerrero *et al.* 2012; Ormeño-Orrillo and Martínez-Romero 2013). The ability to fix N in rhizobia is derived from nitrogenases, which constitute a class of complex enzymes that catalyze biological nitrogen fixation by reducing dinitrogen to ammonia. *Rlt* possess the commonly studied Mo-dependent nitrogenase which is primarily composed of a heterotetrameric MoFe-core encoded by *nifD* and *nifK* that catalyzes the reduction of nitrogen to ammonia, and a dinitrogenase reductase Fe-subunit encoded by *nifH* that provides electrons to nitrogenase and ATP-derived energy for catalysis (Stacey *et al.* 1992; O'Carroll and Dos Santos 2011). Previous studies have found evidence that *nifH* has a different history of HGT from the *nifDK* enzyme core and that genetic divergence of *nifH* and 16S rRNA genes does not commonly correlate well for microbial species (Gaby and Buckley 2014). Haukka *et al.* (1998) concluded that the phylogeny of *nifH* was generally inconsistent with 16S, but was broadly similar to that of *nod* genes. Indeed, phylogenetic trees based on sequences of *nod* genes are generally not congruent with those based on 16S rDNA sequences, but the *nod* trees show some correlation with host plant range (Wernegreen and Riley 1999).

The extent to which sequence variation accounts for recent phenotypic divergence in partner quality should differ between loci, with strong phylogenetic signal near-causative loci; here, we expect that strains of similar partner quality will have less sequence variation near-causative loci. Moreover, the degree to which genomewide (versus unlinked/site-specific) sweeps contribute to phenotypic divergence should be reflected by the degree to which all loci in the genome display phylogenetic signal; for example, without recombination we would expect the phylogenetic signal near-causative loci to be reflected throughout the genome to some degree. Here, we use the collection of N-evolved and control *R. leguminosarum* strains previously isolated from the long-term N deposition experiment at the Kellogg Biological Station LTER site (Weese *et al.* 2015) in phylogenetically informed generalized linear mixed models and comparative phylogenetic analyses to determine the congruence between strains of *Rlt* based on pSym genes and the chromosomal ITS (internal transcribed spacer) region that lies between the 16S and 23S rRNA genes, and to estimate evolutionary relationships among N-evolved and control strains. We chose *nifD* and *nifH* as they encode key components of the nitrogenase enzyme complex, and the *nodD-A* spacer as a marker of nodulation genes. If *nif* explains the majority of partner quality variation, this would suggest that a related functional change, such as a reduction in N fixation efficiency, has driven changes in partner quality. Alternatively, if *nod* explains the majority of partner quality variation, changes to the flavonoid-inducible

regulatory cascade that coordinates the expression of symbiotic genes with nodule development, or potentially a host range shift may have driven the decrease in partner quality. Finally, the degree to which these loci are congruent is illustrative of the relative importance of recombination and HGT on their evolution.

Methods

Study system and sequencing

Briefly, our study focuses on a collection of *Rhizobium leguminosarum* bv. *trifolii* strains (from Weese *et al.* 2015) previously isolated from a long-term N fertilization experiment at the Kellogg Biological Station LTER site (Huberty *et al.* 1998). In previous work, we performed a single-inoculation experiment in which three species of clover were inoculated with 63 strains isolated from either N fertilized (28 strains) or control plots (35 strains). *Rhizobium* strains from N-fertilized plots were of lower mutualistic partner quality (i.e., they resulted in plants with fewer leaves and biomass and lower chlorophyll content). See Weese *et al.* (2015) for full methods on rhizobium isolations, quantitative trait phenotyping, and sequencing of the ITS region. Each strain was grown in TY (Vincent 1970) liquid media prior to DNA extraction via the Qiagen Genra Puregene Yeast/Bacteria kit (Qiagen, Valencia, CA). Three loci from the pSym-located symbiosis island were amplified via PCR: *nifH* and *nifD*, which encode dinitrogenase reductase and the α -subunit of dinitrogenase (which are the key components of the nitrogenase enzyme complex), respectively, and the *nodD-A* region, which spans the intergenic region of *nodD* and *nodA* genes. PCR was performed using the GoTaq PCR protocol (Promega), and successfully amplified samples were sequenced at the University of Illinois at Urbana-Champaign W. M. Keck Center for Sanger sequencing. All ambiguous base calls were manually confirmed using Sequencher 5.1 (Gene Codes Corporation Ann Arbor, MI). The accession number for NifD used in the protein alignment is YP_009081007 (*Rhizobium leguminosarum* bv. *trifolii*) (Miller *et al.* 2007). All datasets were initially aligned using MUSCLE (Edgar 2004) and then trimmed of uneven ends and examined for potential nonhomologs using SeaView 4.5.3 (Gouy *et al.* 2010). MAFFT v7.221 (Katoh and Standley 2013) was used to realign the 55 ITS sequences from strains for which *nifD* genes were available with the Q-INS-i algorithm, and the 36 *nodD* sequences with the L-INS-i algorithm. For each gene comparison, datasets were pruned of taxa that were missing data from one of the two genes, so that all datasets consisted of identical taxa. Separate ML analyses were run for all

individual datasets using the GTRGAMMA model in RAxML (Stamatakis 2014).

Phylogenetic and mixed model analyses

Unless otherwise noted, tests of congruence and recombination were implemented using R (R Core Team 2013). We first applied the SH (Shimodaira–Hasegawa) test, a widely used nonparametric maximum likelihood test. SH tests the null hypothesis that all tested topologies are equally good explanations of the data by comparing the log likelihood values from two or more trees. Even when the number of trees compared is minimized, the SH test is known to be conservative (Buckley 2002). For each gene dataset, the best ML tree for that gene was compared with the best ML topology for the competing gene using the SH test implemented in PAUP* v4.0a146 (Swofford 1993). The likelihood scores for each tree were calculated by estimating model parameters under a GTR+G model, and the SH test was carried out using RELL bootstrapping (100,000 replicates) in PAUP*v4.0a146 (Swofford 1993). In addition to the character-based SH test, we also applied a topology-based test to calculate the measure of incongruence depicted by tanglegrams. Following Pérez-Escobar *et al.* (2015), we applied a PACo (Procrustean Approach to Cophylogeny) analysis (Balbuena *et al.* 2013) implemented in the *ape* and *vegan* packages (Paradis *et al.* 2004; Oksanen *et al.* 2013) to provide a measure of topological congruence by assessing the degree to which one phylogenetic distance matrix predicts another (i.e., is significantly better than random) using a goodness-of-fit statistic with significance established by randomization of the phylogeny association data.

Briefly, using packages *adephylo*, *ape*, and *phylobase* (Paradis *et al.* 2004; Jombart *et al.* 2010; Bolker *et al.* 2013), we utilized *phylo4d()* (Bolker *et al.* 2010) to combine our phylogenies and dataset to visually assess the distribution of partner quality on the phylogenies. Finally, to visually assess whether incongruence was restricted to a particular clade or region of the tree, we formed tanglegrams using the software Dendroscope 3 (Scornavacca *et al.* 2011; Huson and Scornavacca 2012). The amount of tangle, or the degree to which lines cross, is a rough approximation of topological incongruence, and while tanglegrams ignore support values on the trees, they allow a determination of whether incongruence segregates by region or clade.

We used phylogenetic GLMMs (generalized linear mixed models) to estimate the proportion of rhizobium partner quality variation attributable to phylogenetic variation (i.e., linked to the phylogenetic marker locus) versus the recent N fertilization treatment (i.e., changes elsewhere in the genome in response to N fertilization). This

analysis, implemented using the MCMCglmm package in R (Hadfield and Nakagawa 2010), jointly estimates the effects of a phylogenetic distance matrix and other variables on quantitative trait variation (chlorophyll content, leaf number, root mass, shoot mass, and stolon number), similar to the “animal model” of quantitative genetics (Wilson *et al.* 2010). We ran separate models that included N treatment and either the ITS phylogeny or the pSym phylogenies. We followed the recommendations specified by Hadfield (2010) and Villemereuil *et al.* (2012) for stipulating the MCMCglmm priors by dividing the observed phenotypic variance evenly between the effects (the phylogeny and N treatment), with a shape parameter of 1 (weak, to account for phylogenetic uncertainty), and using 1,000,000 iterations with a sample interval of 50 and a burn-in of 100,000. GLMM trace plots were free of pattern, and autocorrelation among samples calculated between iterations were all <0.1, indicating that the model had performed well (Hadfield 2010).

Results

The SH test rejected the null hypothesis of congruence for all pairwise comparisons of loci in our study (Table 1), consistent with some amount of recombination among all loci; the PACo test, however, rejected the null hypothesis of random association and instead supported significant topological congruence in comparisons among all genes (Table 1). Comparisons of the ITS and pSym genes resulted in less congruence via PACo and a high degree of entanglement in tanglegrams compared to among-pSym comparisons (Table 1; Figs. 1, S1–5). When comparing among just the three pSym loci, *nifH* and *nodD-A* were most congruent (Table 1; Fig. S4), while *nifD* and *nifH* were least congruent (Table 1; Fig. S3). In total, these analyses indicate some amount of phylogenetic conflict among loci, but do suggest that the pSym loci are more genealogically concordant with each other compared to the ITS. While the SH test was significant for all comparisons, it is important to note that this test is well known to be conservative (Buckley 2002). Moreover, both of these analyses do not account for phylogenetic uncertainty, which is apparent in these datasets by the overall low bootstrap supports. The pSym datasets are particularly information-limited due to low levels of phylogenetically informative characters (*nifD*: 16/456; *nifH*: 19/834; *nodD-A*: 42/447) relative to the ITS (114/807) and incomplete taxon sampling due to PCR failure despite repeated attempts (≥ 3 attempts/strain/locus) (*nifD*: 55/63; *nifH*: 54/63; *nodD-A*: 37/63). PCR failure might be due to sequence divergence, gene rearrangement, or gene loss, potentially of the entire plasmid.

We found that partitioning of partner quality variation depended considerably on the locus analyzed. The N fertilization treatment explained the majority of rhizobium partner quality for nearly all models that included the ITS, *nifH*, and *nodD-A* phylogenies (Fig. 2). These findings would suggest that loci elsewhere in the genome (i.e., apart from these three marker loci) have changed in response to N fertilization. Conversely, in our analysis of *nifD*, phylogeny explained the vast majority of partner quality variation (Fig. 2), and two clades of generally high- and low-quality stand out when partner quality is mapped to the tips of the tree (Fig. 3A), potentially implicating genetic changes within *nifD* in partner quality variation. Bullseye plots showing partner quality mapped to *ITS*, *nifH*, and *nodD-A* can be found in Figure S6. In

contrast to the other partner quality traits (leaf number, stolon number, and plant biomass), the ITS phylogeny accounted for a large proportion (23.4%) of the variance in chlorophyll content (Fig. 2); moreover, including N treatment in the model of chlorophyll content did not improve model fit. These results suggest that causative loci underlying variation in partner chlorophyll content might be linked to the ITS region, most likely somewhere on the chromosome.

Using an amino acid alignment of *nifD* with the reference, we determined that the reference along with all of the strains from the high-quality clade have a methionine at residue 244, whereas all of the strains from the low-quality clade possess a tyrosine (Fig. 3B). Residue 244 is a MoFe subunit interaction site, and previous studies have shown that changes to these sites can alter a bacterium's ability to fix N (Brigle et al. 1987). Additionally strain 116, which stands out as a low-quality member of a high-quality subclade, is the only strain in the clade that possesses a mutation at residue 100, another MoFe subunit interaction site.

Table 1. Above the diagonal: Shimodaira–Hasegawa test (SH test: In L diff). Below the diagonal: PACo (Procrustean Approach to Cophylogeny: $m^2 = \text{lack-of-fit}$).

Locus	<i>ITS</i>	<i>nifD</i>	<i>nifH</i>	<i>nodD</i>
<i>ITS</i>		1551***	1531***	793***
<i>nifD</i>	0.824****		1162*	106***
<i>nifH</i>	0.921***	0.161****		66**
<i>nodD</i>	0.541***	0.114****	0.090****	

****< 0.0001; ***< 0.001; **< 0.01; *< 0.05.

Discussion

Mutualisms are thought to be susceptible to anthropogenic change, although the ecological and evolutionary

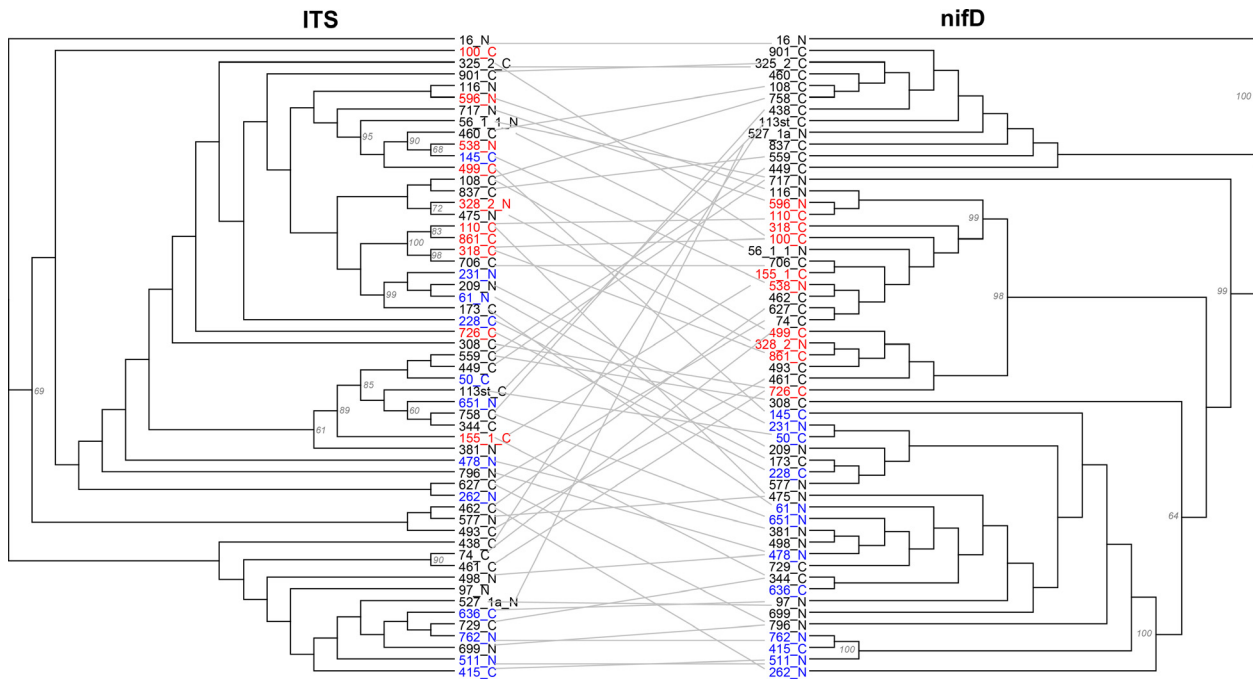


Figure 1. Tanglegram, ITS/*nifD* – A tanglegram highlighting patterns of phylogenetic variation between the ITS and *nifD* phylogenies. All nodes with bootstrap support ≥ 60 are labeled on the phylogenies. Strains in blue represent poor partners and strains in red represent superior partners visually identified from trait mapping (see Fig. 3).

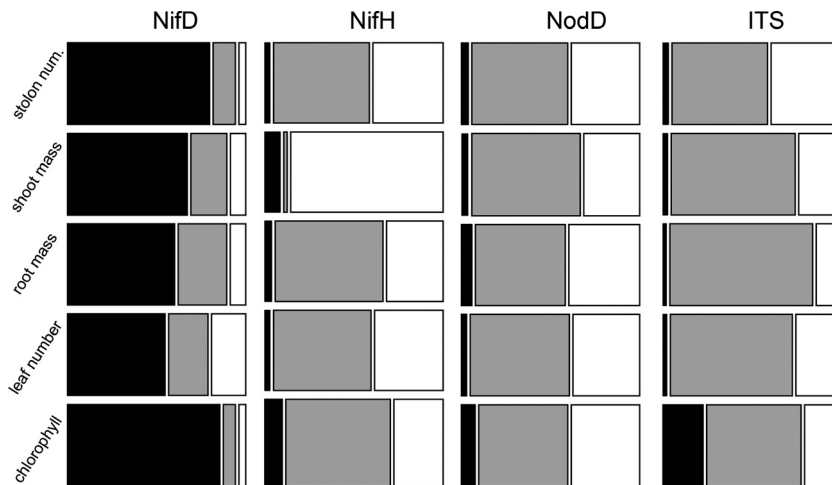
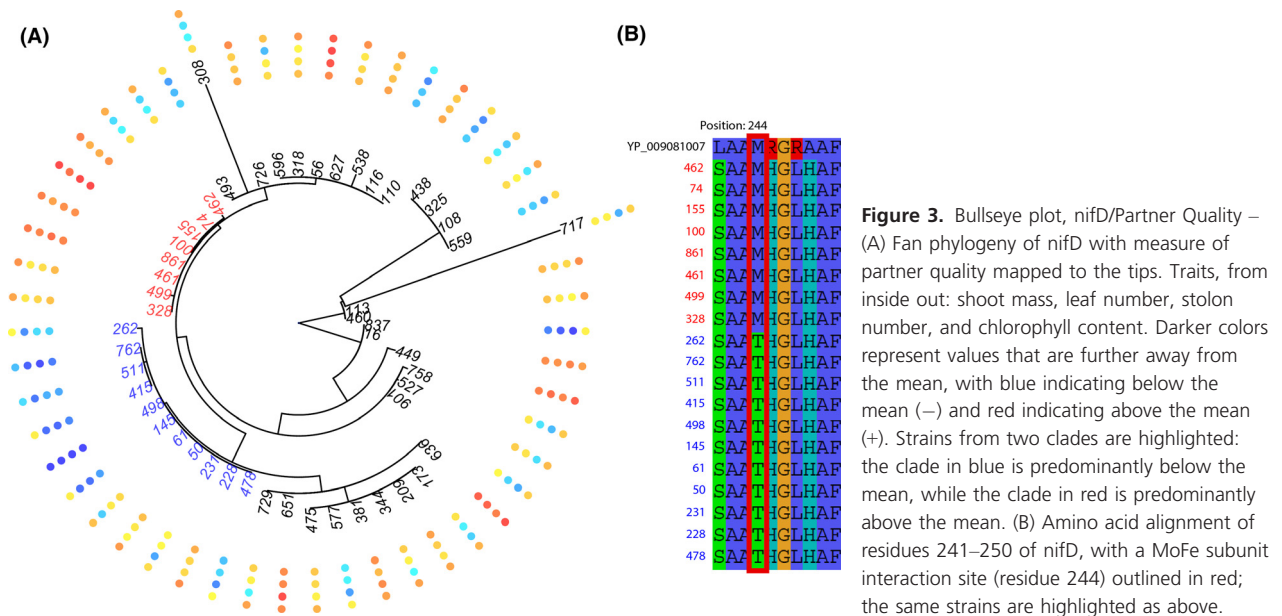


Figure 2. Mosaic plot, GLMM attributable variance – The percent of variance attributable to phylogenetic relationships versus N fertilization treatment. Black: variation attributable to phylogeny; gray: variation attributable to N fertilization; white: residual variance.



processes that drive mutualism decline are not yet understood. Microbes are key players in many important mutualisms, including the N-fixing legume–rhizobium symbiosis; therefore, understanding the evolution of rhizobium partner quality in natural populations can shed light on the evolution of bacteria in nature and the maintenance of mutualisms in general. Building on our previous phenotypic analysis (Weese *et al.* 2015), which indicated that rhizobia from N-fertilized plots were less-beneficial symbionts of clover, we compared evolutionary histories of four gene sequences (one on the chromosome and three pSym genes) with phylogenetic mixed modeling to investigate the evolutionary process underlying the observed reductions in partner quality. Our results indicate that the ITS region and the pSym have different

evolutionary histories and relationships with rhizobium partner quality, but that patterns for chlorophyll content differed from other measures of partner quality. Our results differ from expectations under the ecotype model with zero recombination, but do suggest significant phylogenetic signal in partner quality, particularly at *nifD*, but also the ITS for chlorophyll content. We discuss these main results below.

Our findings indicate both phylogenetic signal and N treatment contribute to among-strain variation in partner quality and thus implicate evolutionary changes in response to experimental N treatments as well as among-lineage variation for mutualism. Given the results of our quantitative trait modeling and various analyses of congruence/coevolution, however, it appears that the

pSym and the chromosome have different evolutionary histories. By contrast, more congruence among the three pSym genes confirms that these genes are in partial linkage and that they are more likely to move as a functional unit (Broughton *et al.* 1984). An alternative explanation is that phylogenetic uncertainty is driving incongruence, as co-diversifying genes that lack in phylogenetic signal may appear incongruent due to random noise (Zaneveld *et al.* 2008). Given previous work showing that rhizobium symbiosis plasmids are self-transmissible (Bittinger *et al.* 2000) and that novel symbiosis islands can confer a strong selective advantage (Parker 2012; Horn *et al.* 2014), it is possible that recombination or HGT of the pSym across ITS backgrounds has led to the differing signals in the various loci.

What do our data suggest about the evolutionary process leading to partner quality decline in high N plots? Because we find evidence for both phylogenetic signal at our marker loci (particularly at *nifD*) and unlinked evolutionary changes in response to N treatment, we can reject a strict one-to-one relationship between phylogeny and partner quality and thus strict clonal evolution. Nevertheless, the correct interpretation of these findings regarding the evolutionary process that has occurred during the last 22 years depends somewhat on the age of these rhizobium lineages, compared to the length of the ecological experiment. On the one hand, if the phylogenetic signal in our dataset predates the initiation of LTER treatments (1988), this would suggest a combination of lineage sorting (changes in abundance of pre-existing lineages) and microevolutionary changes at causative genes in driving the difference in partner quality between N-evolved and control plots. On the other hand, if these lineages have diversified because the N treatments were first established, then our findings would suggest that particular lineages have increased in frequency along with adaptive changes in partner quality. Another interpretation would be that 22 years is not long enough for complete lineage sorting of recent substitutions. Given current best estimates of point mutation rates in bacteria (Wielgoss *et al.* 2011), we would expect approximately four mutations over the course of our 22-year experiment, yet we find 19 informative sites, which implies that the phylogenetic signal predates the LTER treatment. Nevertheless, all three scenarios imply a mix of clonality and recombination consistent with other recent studies of bacteria in nature (Feil *et al.* 2001; Hanage *et al.* 2006; Narra and Ochman 2006; Pérez-Losada *et al.* 2006; Vos and Didelot 2009).

Our results showing that the *nifD* phylogeny accounts for the vast majority of variation in partner quality might suggest a causal relationship between the locus and measures of partner quality, particularly given the lack of variation accounted for by phylogeny at *nifH* or *nodD-A*.

This would be consistent with the known importance of *nifD* in the mutualism (Hennecke 1990; Spaink *et al.* 1998), as well as a previous study showing that mutations at the MoFe subunit interaction sites can lead to changes in the efficiency of N fixation, resulting in a range of effects on growth in N-limiting conditions despite normal accumulation levels of both MoFe subunits (Brigle *et al.* 1987).

While we are unable to rule out the possibility of linked loci elsewhere on the plasmid, it should be noted that there is divergent signal between *nifD* versus the structurally adjacent *nifH/nodD-A* despite evidence of co-diversification, which implies a lack of causally linked loci near our markers. The contribution of chromosomal loci is inconclusive based on sequencing the ITS alone. Understanding which loci determine partner quality and underlie the difference between N-evolved and control rhizobia will require genomewide sequencing, which is currently ongoing. Our current evidence suggests that variation in *nifD* might have played a role in contributing to the reduction in partner quality in N-fertilized strains. The mechanism leading to a change in allele frequency remains unclear; nevertheless, our data are compatible with conjugative transfer of the pSym and subsequent homologous recombination of the *nifD* locus, a hypothesis consistent with previous findings on the architecture and nature of rhizobial plasmids (Mercado-Blanco and Toro 1996; Herrera-Cervera *et al.* 1998; Ding *et al.* 2013).

Interestingly, the ITS phylogeny accounted for considerable variance in chlorophyll content (>23.4%), a key measure of partner quality; this was quite different from other measures of partner quality, where phylogeny never accounted for >3.2% of variation in partner quality. This finding seemingly contrasts with the fact that chlorophyll content and other measures of partner quality are correlated among strain means (chlorophyll content ~ shoot mass, $r^2 = 0.87$, $P < 0.0001$; chlorophyll content ~ leaf number, $r^2 = 0.80$ $P < 0.0001$). Partner quality traits are quantitative, varying more-or-less continuously among strains (Weese *et al.* 2015); therefore, these complex phenotypes are likely to be underlain by multiple loci, some of which might contribute more to some traits than others. Our findings might suggest that the ITS region is in partial linkage with loci that are important in determining plant chlorophyll content, but more work would be needed to test this hypothesis.

Conclusion

Here, we have found that phylogenetic variation at *nifD* explains much of the variation in partner quality, and we suggest MoFe subunit interaction sites as candidates in

the reduction in rhizobium partner quality. We find that the phylogenies of pSym genes are not consistent with the phylogeny of the ITS region, but are broadly similar to each other. These results allow for a number of evolutionary scenarios and further sequencing will be required to determine whether additional changes throughout the genome have driven reductions in partner quality, as well as to elucidate the role of recombination and HGT on whole genomes.

Acknowledgments

This work was supported by NSF DEB-1257938. Support for this research was also provided by the NSF Long-term Ecological Research Program (DEB 1027253) at the Kellogg Biological Station and by Michigan State University AgBioResearch. This is KBS publication #1905. The authors thank Charles Roseman for thoughtful comments on a draft version of the manuscript and Elida Iniguez for help with DNA preparation and sequencing. Sequence data are archived in GenBank (KU517885-KU518029).

Data Archival Location

Genbank (sequence) and TreeBASE (phylogenies).

Conflict of Interest

The authors declare no conflict of interest.

References

- Akçay, E., and E. L. Simms. 2011. Negotiation, sanctions, and context dependency in the legume-Rhizobium mutualism. *Am. Nat.* 178:1–14.
- Allison, S. D., and J. B. H. Martiny. 2008. Resistance, resilience, and redundancy in microbial communities. *Proc. Natl Acad. Sci. USA* 105:11512–11519.
- Bailly, X., I. Olivieri, B. Brunel, J.-C. Cleyet-Marel, and G. Béna. 2007. Horizontal gene transfer and homologous recombination drive the evolution of the nitrogen-fixing symbionts of Medicago species. *J. Bacteriol.* 189:5223–5236.
- Balbuena, J. A., R. Míguez-Lozano, and I. Blasco-Costa. 2013. PACo: a novel Procrustes application to cophylogenetic analysis. *PLoS ONE* 8:e61048.
- Beaber, J. W., B. Hochhut, and M. K. Waldor. 2004. SOS response promotes horizontal dissemination of antibiotic resistance genes. *Nature* 427:72–74.
- Bittinger, M. A., J. A. Gross, J. Widom, J. Clardy, and J. Handelsman. 2000. Rhizobium etli CE3 carries vir gene homologs on a self-transmissible plasmid. *Mol. Plant Microbe Interact.* 13:1019–1021.

- Bolker, B., T. Jombart, S. Kembel, F. Michonneau, and J. Regetz. 2010. phylo4d: Combine a phylogenetic tree with data.
- Bolker, B., M. Butler, P. Cowan, de Vienne D., D. Edelbuettel, M. Holder, et al. 2013. phylobase: Base package for phylogenetic structures and comparative data.
- Brigle, K. E., R. A. Setterquist, D. R. Dean, J. S. Cantwell, M. C. Weiss, and W. E. Newton. 1987. Site-directed mutagenesis of the nitrogenase MoFe protein of *Azotobacter vinelandii*. *Proc. Natl Acad. Sci. USA* 84:7066–7069.
- Bronstein, J. L. 1994. Conditional outcomes in mutualistic interactions. *Trends Ecol. Evol.* 9:214–217.
- Broughton, W. J., N. Heycke, H. Meyer, and C. E. Pankhurst. 1984. Plasmid-linked nif and nod genes in fast-growing rhizobia that nodulate Glycine max, Psophocarpus-tetragonolobus, and Vigna-unguiculata. *Proc. Natl Acad. Sci. USA* 81:3093–3097.
- Brussaard, L., P. C. de Ruiter, and G. G. Brown. 2007. Soil biodiversity for agricultural sustainability. *Agric. Ecosyst. Environ.* 121:233–244.
- Buckley, T. 2002. Model misspecification and probabilistic tests of topology: evidence from empirical data sets. *Syst. Biol.* 51:509–523.
- Cadillo-Quiroz, H., X. Didelot, N. L. Held, A. Herrera, A. Darling, M. L. Reno, et al. 2012. Patterns of gene flow define species of thermophilic Archaea. *PLoS Biol.* 10:e1001265.
- Canfield, D. E., A. N. Glazer, and P. G. Falkowski. 2010. The evolution and future of earth's nitrogen cycle. *Science* 330:192–196.
- Chamberlain, S., D. P. Vázquez, L. Carvalheiro, E. Elle, and J. C. Vamosi. 2014. Phylogenetic tree shape and the structure of mutualistic networks. *J. Ecol.* 102:1234–1243.
- Clark, C., and D. Tilman. 2008. Loss of plant species after chronic low-level nitrogen deposition to prairie grasslands. *Nature* 451:712–715.
- Craine, J. M. 2009. Resource strategies of wild plants. Princeton Univ. Press, Princeton, NJ.
- Diaz, R. J., and R. Rosenberg. 2008. Spreading dead zones and consequences for marine ecosystems. *Science* 321:926–929.
- Ding, H., C. B. Yip, and M. F. Hynes. 2013. Genetic characterization of a novel rhizobial plasmid conjugation system in *Rhizobium leguminosarum* bv. viciae strain VF39SM. *J. Bacteriol.* 195:328–339.
- Edgar, R. C. 2004. MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Res.* 32:1792–1797.
- Elena, S. F., and R. E. Lenski. 2003. Evolution experiments with microorganisms: the dynamics and genetic bases of adaptation. *Nat. Rev. Genet.* 4:457–469.
- Epstein, B., A. Branca, J. Mudge, A. K. Bharti, R. Briskine, A. D. Farmer, et al. 2012. Population genomics of the facultatively mutualistic bacteria *Sinorhizobium meliloti* and *S. medicae*. *PLoS Genet.* 8:e1002868.

- Feil, E. J., E. C. Holmes, D. E. Bessen, M. S. Chan, N. P. Day, M. C. Enright, et al. 2001. Recombination within natural populations of pathogenic bacteria: short-term empirical estimates and long-term phylogenetic consequences. *Proc. Natl Acad. Sci. USA* 98:182–187.
- Frederickson, M. 2013. Rethinking mutualism stability: cheaters and the evolution of sanctions. *Q. Rev. Biol.* 88:269–295.
- Gaby, J. C., and D. H. Buckley. 2014. A comprehensive aligned nifH gene database: a multipurpose tool for studies of nitrogen-fixing bacteria. *Database* 2014:bau001.
- Goldenfeld, N., and C. Woese. 2007. Biology's next revolution. *Nature* 445:369.
- Gouy, M., S. Guindon, and O. Gascuel. 2010. SeaView version 4: a multiplatform graphical user interface for sequence alignment and phylogenetic tree building. *Mol. Biol. Evol.* 27:221–224.
- Gogarten, J. P., W. F. Doolittle, and J. G. Lawrence. 2002. Prokaryotic evolution in light of gene transfer. *Mol. Biol. Evol.* 19:2226–2238.
- Grman, E., J. A. Lau, D. R. Schoolmaster, and K. L. Gross. 2010. Mechanisms contributing to stability in ecosystem function depend on the environmental context. *Ecol. Lett.* 13:1400–1410.
- Hadfield, J. D. 2010. MCMC methods for multi-response generalized linear mixed models: the MCMCglmm R package. *J. Stat. Softw.* 33:1–22.
- Hadfield, J. D., and S. Nakagawa. 2010. General quantitative genetic methods for comparative biology: phylogenies, taxonomies and multi-trait models for continuous and categorical characters. *J. Evol. Biol.* 23:494–508.
- Hanage, W. P., C. Fraser, and B. G. Spratt. 2006. The impact of homologous recombination on the generation of diversity in bacteria. *J. Theor. Biol.* 239:210–219.
- Haukka, K., K. Lindstrom, and J. Young. 1998. Three phylogenetic groups of nodA and nifH genes in Sinorhizobium and Mesorhizobium isolates from leguminous trees growing in Africa and Latin America. *Appl. Environ. Microbiol.* 64:419–426.
- Held, N. L., A. Herrera, and R. J. Whitaker. 2013. Reassortment of CRISPR repeat-spacer loci in *Sulfolobus islandicus*. *Environ. Microbiol.* 15:3065–3076.
- Hennecke, H. 1990. Nitrogen fixation genes involved in the *Bradyrhizobium japonicum*-soybean symbiosis. *FEBS Lett.* 268:422–426.
- Herrera-Cervera, J. A., J. M. Sanjuan-Pinilla, J. Olivares, and J. Sanjuan. 1998. Cloning and identification of conjugative transfer origins in the *Rhizobium meliloti* genome. *J. Bacteriol.* 180:4583–4590.
- Holland, J. N., and D. L. DeAngelis. 2010. A consumer-resource approach to the density-dependent population dynamics of mutualism. *Ecology* 91:1286–1295.
- Horn, K., I. M. Parker, W. Malek, S. Rodríguez-Echeverría, and M. A. Parker. 2014. Disparate origins of Bradyrhizobium symbionts for invasive populations of *Cytisus scoparius* (Leguminosae) in North America. *FEMS Microbiol. Ecol.* 89:89–98.
- Huberty, L., K. Gross, and C. Miller. 1998. Effects of nitrogen addition on successional dynamics and species diversity in Michigan old-fields. *J. Ecol.* 86:794–803.
- Huson, D. H., and C. Scornavacca. 2012. Dendroscope 3: an interactive tool for rooted phylogenetic trees and networks. *Syst. Biol.* 61:1061–1067.
- Jain, R., M. C. Rivera, J. E. Moore, and J. A. Lake. 2003. Horizontal gene transfer accelerates genome innovation and evolution. *Mol. Biol. Evol.* 20:1598–1602.
- Jombart, T., F. Balloux, and S. Dray. 2010. Adephylo: new tools for investigating the phylogenetic signal in biological traits. *Bioinformatics* 26:1907–1909.
- Katoh, K., and D. M. Standley. 2013. MAFFT Multiple sequence alignment software version 7: improvements in performance and usability. *Mol. Biol. Evol.* 30:772–780.
- Kiers, E. T., R. A. Rousseau, and R. F. Denison. 2006. Measured sanctions: legume hosts detect quantitative variation in rhizobium cooperation and punish accordingly. *Evol. Ecol. Res.* 8:1077–1086.
- Kiers, T. E., T. M. Palmer, A. R. Ives, J. F. Bruno, and J. L. Bronstein. 2010. Mutualisms in a changing world: an evolutionary perspective. *Ecol. Lett.* 13:1459–1474.
- Koepfel, A., E. B. Perry, J. Sikorski, D. Krizanc, A. Warner, D. M. Ward, et al. 2008. Identifying the fundamental units of bacterial diversity: a paradigm shift to incorporate ecology into bacterial systematics. *Proc. Natl Acad. Sci. USA* 105:2504–2509.
- Kumar, N., G. Lad, E. Giuntini, M. E. Kaye, P. Udomwong, N. J. Shamsani, et al. 2015. Bacterial genospecies that are not ecologically coherent: population genomics of *Rhizobium leguminosarum*. *Open Biol.* 5:140133.
- Logares, R., E. S. Lindström, S. Langenheder, J. B. Logue, H. Paterson, J. Laybourn-Parry, et al. 2013. Biogeography of bacterial communities exposed to progressive long-term environmental change. *ISME J.* 7:937–948.
- López-Guerrero, M. G., E. Ormeño-Orrillo, J. L. Acosta, A. Mendoza-Vargas, M. A. Rogel, M. A. Ramírez, et al. 2012. Rhizobial extrachromosomal replicon variability, stability and expression in natural niches. *Plasmid* 68:149–158.
- Mercado-Blanco, J., and N. Toro. 1996. Plasmids in rhizobia: The role of nonsymbiotic plasmids. *Mol. Plant-Microbe Interact.* 9:535–545.
- Miller, S. H., R. M. Elliot, J. T. Sullivan, and C. W. Ronson. 2007. Host-specific regulation of symbiotic nitrogen fixation in *Rhizobium leguminosarum* biovar trifolii. *Microbiology* 153:3184–3195.
- Mozhayskiy, V., and I. Tagkopoulos. 2012. Horizontal gene transfer dynamics and distribution of fitness effects during microbial *in silico* evolution. *BMC Bioinformatics* 13(Suppl. 10):S13.

- Narra, H. P., and H. Ochman. 2006. Of what use is sex to bacteria? *Curr. Biol.* 16:705–710.
- Newton, R. J., S. M. Huse, H. G. Morrison, C. S. Peake, M. L. Sogin, and S. L. McLellan. 2013. Shifts in the microbial community composition of Gulf Coast beaches following beach oiling. *PLoS ONE* 8:e74265.
- O'Carroll, I. P., and P. C. Dos Santos. 2011. Genomic analysis of nitrogen fixation. Pp. 49–65 *in* M. W. Ribbe, ed. *Nitrogen fixation: methods and protocols*. Humana Press, New York City.
- Ochman, H., and N. Moran. 2001. Genes lost and genes found: Evolution of bacterial pathogenesis and symbiosis. *Science* 292:1096–1098.
- Oksanen, J., F. Blanchet, R. Kindt, P. Legendre, P. Minchin, R. O'Hara, et al. 2013. *vegan: community ecology package*. R package version 2.0-8.
- Oono, R., R. F. Denison, and E. T. Kiers. 2009. Controlling the reproductive fate of rhizobia: how universal are legume sanctions? *New Phytol.* 183:967–979.
- Ormeño-Orrillo, E., and E. Martínez-Romero. 2013. Phenotypic tests in *Rhizobium* species description: an opinion and (a sympatric speciation) hypothesis. *Syst. Appl. Microbiol.* 36:145–147.
- Orozco-Mosqueda, M. D. C., J. Altamirano-Hernandez, R. Farias-Rodriguez, E. Valencia-Cantero, and G. Santoyo. 2009. Homologous recombination and dynamics of rhizobial genomes. *Res. Microbiol.* 160:733–741.
- Paradis, E., J. Claude, and K. Strimmer. 2004. APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* 20:289–290.
- Parker, M. A. 2012. Legumes select symbiosis island sequence variants in *Bradyrhizobium*. *Mol. Ecol.* 21:1769–1778.
- Pérez-Escobar, O. A., J. A. Balbuena, and M. Gottschling. 2015. Rumbling orchids: how to assess divergent evolution between chloroplast endosymbionts and the nuclear host. *Syst. Biol.* 65:51–65.
- Pérez-Losada, M., E. B. Browne, A. Madsen, T. Wirth, R. P. Viscidi, and K. A. Crandall. 2006. Population genetics of microbial pathogens estimated from multilocus sequence typing (MLST) data. *Infect. Genet. Evol.* 6:97–112.
- Pimentel, D., C. Wilson, C. McCullum, R. Huang, P. Dwen, J. Flack, et al. 1997. Economic and environmental benefits of biodiversity. *Bioscience* 47:747–757.
- Polz, M., E. Alm, and W. Hanage. 2013. Horizontal gene transfer and the evolution of bacterial and archaeal population structure. *Trends Genet.* 29:170–175.
- R Core Team. 2013. *R: a language and environment for statistical computing*. R Foundation for Statistical Computing, Vienna, Austria.
- Regus, J. U., K. A. Gano, A. C. Hollowell, and J. L. Sachs. 2014. Efficiency of partner choice and sanctions in *Lotus* is not altered by nitrogen fertilization. *Proc. R. Soc. B* 281:20132587.
- Reno, M. L., N. L. Held, C. J. Fields, P. V. Burke, and R. J. Whitaker. 2009. Biogeography of the *Sulfolobus islandicus* pan-genome. *Proc. Natl Acad. Sci. USA* 106:8605–8610.
- Sachs, J. L., and E. L. Simms. 2006. Pathways to mutualism breakdown. *Trends Ecol. Evol.* 21:585–592.
- Sachs, J. L., U. G. Mueller, T. P. Wilcox, and J. J. Bull. 2004. The evolution of cooperation. *Q. Rev. Biol.* 79:135–160.
- Sachs, J. L., A. C. Hollowell, and J. E. Russell. 2011. Evolutionary instability of symbiotic function in *Bradyrhizobium japonicum*. *PLoS ONE* 6:e26370.
- Saffo, M. B. 2014. Mutualistic symbioses. In: *eLS*. John Wiley & Sons Ltd, Chichester. <http://www.els.net> [doi: 10.1002/9780470015902.a0003281.pub2]
- Scornavacca, C., F. Zickmann, and D. H. Huson. 2011. Tanglegrams for rooted phylogenetic trees and networks. *Bioinformatics* 27:i248–i256.
- Shapiro, B. J., J. Friedman, O. X. Cordero, S. P. Preheim, S. C. Timberlake, G. Szabo, et al. 2012. Population genomics of early events in the ecological differentiation of bacteria. *Science* 336:48–51.
- Six, D. L. 2009. Climate change and mutualism. *Nat. Rev. Microbiol.* 7:686.
- Smil, V. 2002. Nitrogen and food production: proteins for human diets. *Ambio* 31:126–131.
- Spaink, H. P., A. Konodorosi, and P. Hooykaas. 1998. *The rhizobiaceae: molecular biology of model plant-associated bacteria*, 1st ed. Kluwer Academic Publishers, Dordrecht, Netherlands.
- Sprent, J., and J. Sutherland. 1987. Some aspects of the biology of nitrogen-fixing organisms. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 317:111–129.
- Stacey, G., R. Burris, and H. Evans. 1992. *Biological nitrogen fixation*. Chapman and Hall, New York.
- Stachowicz, J. J. 2001. Mutualism, facilitation, and the structure of ecological communities. *Bioscience* 51: 235–246.
- Stallforth, P., D. A. Brock, A. M. Cantley, X. Tian, D. C. Queller, J. E. Strassmann, et al. 2013. A bacterial symbiont is converted from an inedible producer of beneficial molecules into food by a single mutation in the *gacA* gene. *Proc. Natl Acad. Sci. USA* 110:14528–14533.
- Stamatakis, A. 2014. RAxML Version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. *Bioinformatics* 30:1312–1313.
- Sterner, R. W., and J. J. Elser. 2002. *Ecological stoichiometry: the biology of elements from molecules to the biosphere*. Princeton Univ. Press, Princeton, NJ.
- Swofford, D. L. 2003. *PAUP*. Phylogenetic Analysis Using Parsimony (*and Other Methods)*. Version 4. Sinauer Associates, Sunderland, Massachusetts.
- Tilman, D. 1987. Secondary succession and the pattern of plant dominance along experimental nitrogen gradients. *Ecol. Monogr.* 57:189–214.

- de Villemereuil, P., J. A. Wells, R. D. Edwards, and S. P. Blomberg. 2012. Bayesian models for comparative analysis integrating phylogenetic uncertainty. *BMC Evol. Biol.* 12:102–116.
- Vincent, J. 1970. A manual for the practical study of root-nodule bacteria. *Int. Biol. Program. Handb.* 15:164.
- Vitousek, P. M. 1997. Human domination of earth's ecosystems. *Science* 277:494–499.
- Vitousek, P. M., J. D. Aber, R. W. Howarth, G. E. Likens, P. A. Matson, D. W. Schindler, et al. 1997. Human alteration of the global nitrogen cycle: sources and consequences. *Ecol. Appl.* 7:737–750.
- Vos, M., and X. Didelot. 2009. A comparison of homologous recombination rates in bacteria and archaea. *ISME J.* 3:199–208.
- Weese, D. J., K. D. Heath, B. T. M. Dentinger, and J. A. Lau. 2015. Long-term nitrogen addition causes the evolution of less-cooperative mutualists. *Evolution* 69:631–642.
- Wernegreen, J. J., and M. A. Riley. 1999. Comparison of the evolutionary dynamics of symbiotic and housekeeping loci: a case for the genetic coherence of rhizobial lineages. *Mol. Biol. Evol.* 16:98–113.
- Wernegreen, J. J., E. E. Harding, and M. A. Riley. 1997. Rhizobium gone native: unexpected plasmid stability of indigenous *Rhizobium leguminosarum*. *Proc. Natl Acad. Sci. USA* 94:5483–5488.
- West, S. A., E. T. Kiers, E. L. Simms, and R. F. Denison. 2002. Sanctions and mutualism stability: why do rhizobia fix nitrogen? *Proc Biol Sci.* 269:685–694.
- West, S. A., A. S. Griffin, and A. Gardner. 2007. Evolutionary explanations for cooperation. *Curr. Biol.* 17:R661–R672.
- Wielgoss, S., J. E. Barrick, O. Tenaillon, S. Cruveiller, B. Chane-Woon-Ming, C. Medigue, et al. 2011. Mutation rate inferred from synonymous substitutions in a long-term evolution experiment with *Escherichia coli*. *G3: Genes - Genomes - Genetics* 1:183–186.
- Wilson, A. J., D. Réale, M. N. Clements, M. M. Morrissey, E. Postma, C. A. Walling, et al. 2010. An ecologist's guide to the animal model. *J. Anim. Ecol.* 79:13–26.
- Youngblut, N. D., M. Dell'aringa, and R. J. Whitaker. 2013. Differentiation between sediment and hypolimnion methanogen communities in humic lakes. *Environ. Microbiol.* 16:1411–1423.
- Young, J. P. W., L. C. Crossman, A. W. B. Johnston, N. R. Thomson, Z. F. Ghazoui, K. H. Hull, et al. 2006. The genome of *Rhizobium leguminosarum* has recognizable core and accessory components. *Genome Biol.* 7:R34.
- Zaneveld, J. R., D. R. Nemergut, and R. Knight. 2008. Are all horizontal gene transfers created equal? Prospects for mechanism-based studies of HGT patterns. *Microbiology* 154:1–15.

Supporting Information

Additional Supporting Information may be found in the online version of this article:

Figure S1. Tanglegram of ITS/nifH.

Figure S2. Tanglegram of ITS/nodD.

Figure S3. Tanglegram of nifD/nifH.

Figure S4. Tanglegram of nifD/nodD.

Figure S5. Tanglegram of nifH/nodD.

Figure S6. Bullseye plot showing partner quality mapped to the fan phylogenies of *ITS*, *nifH*, and *nodD-A*. Traits, from inside out: shoot mass, leaf number, stolon number, and chlorophyll content. Darker colors represent values that are further away from the mean, with blue indicating below the mean (-) and red indicating above the mean (+).